

**DEVELOPMENT OF A MONOLITH BASED -
CHROMATOGRAPHIC PURIFICATION
SYSTEM FOR PLASMID DNA
VACCINE AGAINST THE
GROUPE IRIDOVIRUS**

TAMAR KANSIL

**PERPUSTAKAAN
UNIVERSITI MALAYSIA SABAH**

**THESIS SUBMITTED IN PARTIAL FULFILLMENT FOR
THE DEGREE OF MASTER OF SCIENCE**

**BIOTECHNOLOGY RESEARCH INSTITUTE
UNIVERSITI MALAYSIA SABAH
2016**



BORANG PENGESAHAN STATUS TESIS

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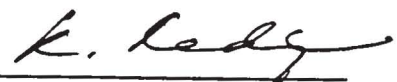
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(Dr. Clarence M. Ongkudon)



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DECLARATION

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6th September 2016



Tamar Kansil
MZ1311005T

PERPUSTAKAAN
UNIVERSITI MALAYSIA SABAH

CERTIFICATION

NAME : TAMAR KANSIL
MATRIC NO. : MZ1311005T
TITLE : DEVELOPMENT OF A CHROMATOGRAPHIC
PURIFICATION SYSTEM FOR GROUPER'S
IRIDOVIRUS PLASMID DNA VACCINE
DEGREE : MASTER OF SCIENCE
DATE OF VIVA : 18th JULY 2016

DECLARED BY;

1. **SUPERVISOR**
Dr. Clarence M. Ongkudon
2. **CO-SUPERVISOR**
Assoc. Prof. Dr. Kenneth F. Rodrigues

Signature



CERTIFICATION

NAME : **TAMAR KANSIL**
MATRIC NO. : **MZ1311005T**
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1. SUPERVISOR
Dr. Clarence M. Ongkudon

Signature

2. CO-SUPERVISOR
Assoc. Prof. Dr. Kenneth F. Rodrigues

ACKNOWLEDGEMENT

I would like to take this privilege to thank God for His great blessings throughout my entire master's degree project. My deep gratitude goes to Dr. Clarence M. Ongkudon, my research supervisor, for his strong motivation, encouragement, guidance and valuable comments that helped me in my postgraduate's study. My sincere appreciation also goes to Assoc. Prof. Dr. Kenneth F. Rodrigues, my research co-supervisor who has guided, motivated and gave valuable advices to me during my research endeavour. Also, no forgetting Puan Marlenny Jumadi, En. Mony Mian, Puan Vidarita Maikin, and Puan Azian Ag. Besar for assisting me in the laboratory whenever I needed them the most. I would also like to extend my sincere appreciation to my fellow postgraduate friends, Chan Yi Wei, Ahmad Zaidi, Fernandes, Haniza, and others for making my research experience in Biotechnology Research Institute of University Malaysia Sabah a memorable one. Last but not least, I could not thank enough my family for their continuous supports, prayers, financial assistances, and encouragements throughout my entire research.

Tamar Kansil
6th September 2016

ABSTRACT

Routine research-scale production of plasmid DNA vaccine relies heavily on commercial kits that are expensive due to the application of RNase, including the use of multiple elution steps and single-use columns which are subjected to lot-to-lot quality deficiency. This research focused on developing an innovative plasmid DNA purification strategy by using a non-functionalized polymethacrylate monolith in centrifugal-based purification technique coupled with a modified lysis procedure. The effect of temperature on pore size distribution was first analyzed to determine the suitability of the monolith for plasmid DNA purification. Initially, the plasmid DNA was purified via monolithic chromatography using an automated chromatography system based on ionic interaction to determine the optimal chromatographic conditions such as flow rate and elution condition. The centrifugal-based monolithic purification was then conducted by mimicking the elution conditions used in the automated chromatography system. The main obstacle encountered during the research was the limited versatility when using non-functionalized polymethacrylate monoliths to purify plasmid DNA from crude cell extracts. In most cases, a reasonable amount of RNA was found to co-elute with plasmid DNA. This greatly affected the purification efficiency as the separation was presumably relied on size exclusion as the only mean of separation. However, we discovered that the monolith was inherently charged even without a functional group immobilization. Apparently, an electrostatic interaction between plasmid DNA and charged matrix had occurred. This was evident by the gel electrophoresis of samples obtained after several elutions using deionized water. There was no noticeable RNA or plasmid DNA observed on the gel indicating that an interaction between nucleic acids and the monolith had taken place. The overall purification methodology comprised pretreatment of lysed cells with sulphate ions followed by step-wise elution of plasmid DNA using monolithic column via centrifugation (300 x g) with NaCl as the elution buffer. The first elution step involved the use of low concentration of saline buffer (0.2 M NaCl) to remove the remaining RNA from the pretreated cell extracts with some of the plasmid DNA were being compromised. The second elution step involved the use of high concentration (1.0 M NaCl) buffer to elute the remaining bound plasmid DNA. The study showed that 70% porogen content in the polymerization mixture gave a minimum heat build-up of 64°C and a homogenous pore size distribution of 3µm – 4µm. In this research, it was evident that the pretreatment of lysed cells with 0.8 M sulphate salt solution resulted in the removal of more than 80% RNAs with a minimal amount of plasmid DNA being compromised. This method allowed the removal of the remaining RNA with a high recovery of plasmid DNA during the first step elution using monolithic centrifugation technique (two-step elution). The discovery of a chemically induced RNA removal method based on the bias selection of sulphate salt ion and the establishment of a centrifugal-based monolithic purification platform would have a great impact on the production of plasmid DNA vaccine for grouper's Iridovirus. The developed method eliminates the use of an expensive enzyme RNAase making it economically favourable. The monolithic purification column can also be used repeatedly with consistent outputs thus enabling a high-throughput production of Iridovirus plasmid DNA vaccine for *in vivo* testing.

ABSTRAK

Perkembangan Sistem Penuliran Kromatografi untuk Iridovirus Plasmid Vaksin DNA Kerapu

Penghasilan rutin vaksin DNA plasmid berskala penyelidikan banyak bergantung kepada kit komersil yang mahal disebabkan oleh penggunaan 'RNA-ase', di samping ia juga melibatkan pelbagai langkah pengeluaran dan penggunaan kolum kromatografi kolom yang terhad kepada sekali sahaja dan ini mendorong kepada penurunan dari segi kualiti produk. Kajian ini tertumpu kepada pembangunan strategi untuk penuliran DNA plasmid secara inovatif dengan menggunakan monolit poli(metakrilat) tanpa kumpulan berfungsi di dalam teknik penuliran berasaskan pengemparan yang digabungkan dengan prosedur lisis yang terubahsuai. Kesan suhu ke atas taburan saiz liang dianalisa untuk menentukan kesesuaian monolit bagi penuliran DNA plasmid. Pada mulanya, penuliran menggunakan monolit berasaskan pengemparan telah dijalankan dengan mengaplikasikan kadar aliran dan parameter pengeluaran yang digunakan dalam sistem kromatografi-automatik. Dalam kebanyakan kajian, sejumlah RNA didapati mengeluat bersama DNA selepas proses penuliran. Keadaan ini telah menghadkan kaedah-kaedah penuliran kepada penuliran berasaskan saiz molekul sebagai prinsip pengasingan yang tunggal. Hasil kajian ini menunjukkan bahawa tiada RNA atau plasmid DNA yang kelihatan pada gel selepas beberapa langkah pengeluaran menggunakan air tanpa ion dan ini menerangkan bahawa terdapat interaksi antara asid nukleik dan monolit. Penuliran keseluruhan terdiri daripada rawatan awal sel-sel terlis dengan ion sulfat diikuti dengan langkah berperingkat, pengeluaran DNA plasmid menggunakan kolum monolit melalui pengemparan (300 x g) dengan NaCl sebagai larutan penimbal. Langkah pertama pengeluaran melibatkan penggunaan penimbal yang mempunyai kepekatan garam rendah (0.2 M NaCl) untuk mengeluarkan sisa RNA dari ekstrak sel yang telah melalui rawatan awal dan sebahagian DNA plasmid yang turut hilang pada peringkat ini. Langkah pengeluaran kedua melibatkan penggunaan penimbal yang mempunyai kepekatan garam tinggi (1.0 M NaCl) untuk mengeluarkan sisa DNA plasmid yang terikat. Kajian menunjukkan 70% kandungan porogen dalam campuran pempolimeran memberikan bacaan haba terbentuk yang minimum sebanyak 64°C dan taburan saiz liang yang sekata dalam julat antara 3 µm – 4 µm. Hasil keputusan juga menunjukkan rawatan awal sel lisis dengan 0.8 M ion sulfat dapat menyingkirkan sebanyak 80 % RNA dengan sedikit kehilangan DNA plasmid. Kaedah ini dapat menyingkirkan sisa RNA dengan pengumpulan semula DNA plasmid yang tinggi semasa langkah pengeluaran pertama menggunakan teknik pengemparan monolitik. Penemuan kaedah penyingkiran RNA aruhan secara kimia berasaskan pemilihan ion sulfat secara berat sebelah dan pembangunan kaedah penuliran menggunakan monolit berasaskan emparan akan memberi impak yang besar ke atas penghasilan vaksin DNA plasmid terhadap Iridovirus kerapu. Kaedah yang dibangunkan ini akan dapat menggantikan penggunaan enzim 'RNA-ase' yang mahal dan menyebabkan ianya menjadi pilihan. Kolum penuliran monolit ini juga boleh digunakan berulang kali dengan hasil yang konsisten, dan dengan itu, membenarkan penghasilan vaksin DNA plasmid Iridovirus dengan daya pemprosesan yang tinggi untuk ujian in vivo.

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- Figure 4.22: Optimal concentration of first step elution solution. Gel electrophoresis of pMCS2.1-IRIDOVIRUS of *E. coli* isolated via monolithic (non-functionalized) chromatography purification with 0.2 M – 0.3 M NaCl as the 1st step elution solution, and 1.0 M as the 2nd step elution solution. Analysis was performed in 1 % agarose at 100 V for 0.7 h. 50
- Figure 4.23: Chromatograms of 0.2 M – 0.3 M NaCl runs. Chromatogram of *E. coli* plasmid DNA extraction using Next Generation 50

Chromatography System (BIORAD) via alkaline lysis. 0.2 M – 0.3 M NaCl are used during the first peak elution followed by 1.0 M NaCl for second peak elution. Flow rate of 1.0 mL/min.

Figure 4.24: Recovery percentage (%) of plasmid DNA (pDNA) and RNA at 0.21 M, 0.25 M and 0.29 M NaCl during second peak elution. 51

Figure 4.25: The comparison of number of frits used in the conical column. Preliminary results of gel electrophoresis of pMCS2.1-IRIDOVIRUS of *E. coli* isolated with innovative conical monolithic column in centrifugal format. Plasmid DNA elution was carried out using deionized water. Analysis was performed in 1 % agarose of 60 mL TBE x 1 buffer at 100 V for 0.7 h. 53

LIST OF ABBREVIATIONS

GI	-	Grouper iridovirus
HSV	-	Hemorrhagic septicemia virus
IPNV	-	Infectious pancreatic necrosis virus
IHN	-	Infectious haematopoietic necrosis virus
ISAV	-	Infectious salmon anaemia virus
Nm	-	Nanometre
RPS-	-	Relative percentage survival
DNA	-	Deoxyribonucleic acid
RNA	-	Ribonucleic acid
CMV	-	Cytomegalovirus
KHV	-	Koi herpes virus
IgM	-	Immunoglobulin M
MHC	-	Major histocompatibility complex
CD8+	-	Cluster of differentiation 8 +
CPG	-	5'-C-phosphate-G-3'
GM-CSF	-	Granulocyte-macrophage colony-stimulating factor
µg	-	Microgram
µl	-	Microliter
IM	-	Intramuscular injection
IFN	-	Interferon
VHS	-	Viral hemorrhagic septicemia
IL	-	Interleukin
Th2	-	Type 2 T helper
H	-	Hyperosmotic infiltration
DI	-	Direct immersion
gDNA	-	Genomic DNA
pDNA	-	Plasmid DNA
VLP	-	Virus-like particles
BEVS	-	Baculovirus expression vector
IC	-	Insect cell
SEC	-	Size exclusion

AC	-	Affinity chromatographic
AEX	-	Anion-exchange chromatography
D_{pore}	-	Pore radius
ε_p	-	Particle porosity
A_{internal}	-	Specific surface area
CIM	-	Convective interaction media
mg	-	Miligram
mL	-	Mililitre
L	-	Litre
cm	-	centimetre
Min	-	Minute
EDMA	-	Ethylene glycol dimethacrylate
GMA	-	Glycidyl methacrylate
AIBN	-	Azobisisobutyronitrile
NaCl	-	Sodium chloride
SDS	-	Sodium dodecyl sulfate
MeOH	-	Methanol
EDTA	-	Ethylenediaminetetraacetic acid
RPM	-	Revolutions per minute
Tris-HCL	-	Tris-Hydrochloride
CH₃COOK	-	Potassium acetate
SEM	-	Scanning electron microscope
NGC	-	Next generation chromatography
RCF	-	Relative centrifugal force
Kb	-	Kilobase
TBE	-	Tris-borate-EDTA
PDM	-	pUC-based plasmid DNA semi-defined medium
OD	-	Optical density
LB	-	Lysogeny broth
Hr	-	Hour
Vs	-	Versus
kV	-	Kilovolt
T_{poly}	-	Polymerization temperature
T_{max}	-	Maximal temperature

AC	-	Affinity chromatographic
AEX	-	Anion-exchange chromatography
D_{pore}	-	Pore radius
ε_p	-	Particle porosity
A_{internal}	-	Specific surface area
CIM	-	Convective interaction media
mg	-	Miligram
mL	-	Mililitre
L	-	Litre
cm	-	centimetre
Min	-	Minute
EDMA	-	Ethylene glycol dimethacrylate
GMA	-	Glycidyl methacrylate
AIBN	-	Azobisisobutyronitrile
NaCl	-	Sodium chloride
SDS	-	Sodium dodecyl sulfate
MeOH	-	Methanol
EDTA	-	Ethylenediaminetetraacetic acid
RPM	-	Revolutions per minute
Tris-HCL	-	Tris-Hydrochloride
CH₃COOK	-	Potassium acetate
SEM	-	Scanning electron microscope
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Hr	-	Hour
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kV	-	Kilovolt
T_{poly}	-	Polymerization temperature
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