# THE DEVELOPMENT OF SENSORY ORGANS IN AFRICAN CATFISH, Clarias gariepinus DURING LARVAL TO JUVENILE STAGE

### **MAZFARINA BINTI MUSTAFA**

# THIS DISSERTATION IS SUBMITTED IN PARTIAL FULFILLMENT OF THE REQUIREMENT FOR THE BACHELOR OF SCIENCE WITH HONOURS

PERPUSTAKAAN UNIVERSITI MALAYSIA SABAR

# AQUACULTURE PROGRAMME SCHOOL OF SCIENCE AND TECHNOLOGY

## UNIVERSITI MALAYSIA SABAH

**APRIL 2008** 



PUMS99:1

### UNIVERSITI MAŁAYSIA SABAH

BORANG PENGESAHAN STATUS TESIS@	
JUDUL THE DEVELOPMENT OF AFRICAN CATFISH, CLAMAS	
ganepinus DURING LARVAL TO OUVENILE STAGE	*
UAZAH: SARJANA NUDA SAINS AKUAKULTUR	
SAYA MAZFARINA BT MUSTAFA (HURUF BESAR) SESI PENGAJIAN: 2005/2005	
mengaku membenarkan tesis (LPSM/Sarjana/Doktor Falsafah) ini disimpan di Perpustakaan Universiti Malaysia Sabah dengan syarat-syarat kegunaan seperti berikut:-	4
<ol> <li>Tesis adalah hakmilik Universiti Malaysia Sabah.</li> <li>Perpustakaan Universiti Malaysia Sabah dibenarkan membuat salinan untuk tujuan pengajian sahaja.</li> <li>Perpustakaan dibenarkan membuat salinan tesis ini sebagai bahan pertukaran antara institutsi pengajian tinggi.</li> </ol>	-
4. Sila tandakan (/)	
SOLIT       (Nonganoungi maktumat yang termaktub di dalam         Kepentingan Malaysia seperti yang termaktub di dalam         AKTA RAHSIA RASMI 1972)         TERHAD         (Mengandungi maklumat TERHAD yang telah ditentukan oleh organisasi/badan di mana penyelidikan dijalankan)	
TIDAK TERHAD Disahkan Oleh	
(TANDATANGAN PENULIS) (TANDATANGAN PENULIS)	N SIA SABAH
Alamat Tetap: LOT 60, KG. BIAH JUN PEROL, 16010, KOTA BHARY, KELANTAN Nama Penyelia	
Tarikh: 30 APRIL 2008 Tarikh:	
<ul> <li>CATATAN:- *Potong yang tidak berkenaan.</li> <li>**Jika tesis ini SULIT atau TERHAD, sila lampirkan surat daripada pihak berkuasa /organisasi berkenaan dengan menyatakan sekali sebab dan tempoh tesis ini perlu dikelaskan sebagai SULIT dan TERHAD.</li> <li>@Tesis dimaksudkan sebagai tesis bagi Ijazah Doktor Falsafah dan Sarjana secara penyelidikan atau disertai bagi pengajian secara kerja kursus dan Laporan Projek Sarjana Muda (LPSM).</li> </ul>	



I declare that this dissertation is the result of my own independent work except where otherwise stated.

30<sup>th</sup> APRIL 2008

MAZFARINA MUSTAFA HS2005-1755



### **AUTHENTICATION**

I/We, members of the Dissertation Supervisory Committee, certify that this dissertation has fulfilled all the research requirements for the Bachelor Degree in Science with Honours, has been verified and is qualified for examination.

Members of Dissertation Supervisory Committee

Signature

Yeh Anda

2. **EXAMINER 1** 

Dr. Yukinori Mukai

SUPERVISOR

1.

(Muhammad Kli lyed Husein

3. EXAMINER 2 (Masern Tomaka.)

Branka

4. DEAN OF SST

Supt/Ks. Prof. Madya. Dr. Shariff A. Kadir

S. Omang

UNIVERSITI MALAYSIA SABAH

### ACKNOWLEDGEMENTS

Praise be to Allah for it is with His Grace that this dissertation has been completed successfully.

I would like to express my sincere grateful acknowledgement to my supervisor of my final year project, Dr. Yukinori Mukai for his invaluable suggestions, guidance, encouragement, support and patience throughout my study period. The wide range of reference he suggested was thoroughly helpful.

I would like to thank Y. Bhg. Prof. Dato. Dr. Mohd. Noh bin Dalimin, Vice-Chancellor of University of Malaysia Sabah, Prof. Dr. Saleem Mustafa, Director of Marine Borneo Research Institute, Prof. Dr. Shigeharu Senoo for his support and teachings, to Mr. Muhammad Ali Syed Hussein for his understanding and kindness, all BMRI lecturers, BMRI and UMS hatchery staff and RAs, for their encouragement and assistance.

My sincere appreciation is also extended to Ms. Audrey Daning Tuzan, Ms. Shahirah Yahaya, Ms. Dayana and Mr. Lim Leong Seng for their support and helpful teaching in complete my final year project. I wish to heartily express my indebtness to all my dear colleagues and friends especially Mr. Abdul Manan, Mr. Zulfa, Ms. Surianun, Ms. Siti Farhana, Ms.Nurhidayahanum, Ms. Siti Fairus, Ms. Atiqah, Mr. Benjamin Teo, Mr. Tan Nai Han, and Mr. Beh Cheng Yang for their assistance and valuable inputs directly or indirectly for the successful completion of the dissertion.

Last but not least, I would like to express my heartfelt gratitude to my dearest family, ma, Saniah Abdullah; abah, Mustafa Yusoh; brothers, Wadi, Firdaus and Lukman and my sisters, Wani, Eira, and Ema for their love, emotional support, patience, encouragement and support throughout my difficult period. Love you all very much.



### ABSTRAK

Kajian ini menjelaskan perkembangan organ deria larva keli Afrika, Clarias gariepinus dan bertujuan untuk memahami ekologinya serta memperbaiki kaedah pengkulturan larva. Telur diperoleh dari Institut Penyelidikan Marin Borneo dan larva keli Afrika dikultur dari peringkat penetasan sehingga juvenil. Penyampelan dilakukan setiap hari dan perkembangan organ derianya diperhatikan dengan menggunakan mikroskop cahaya. Larva diberi makan rotifer, Artemia nauplii dan serbuk makanan tiruan. Panjang keseluruhan larva keli Afrika pada peringkat 20, 25 dan 30 hari adalah 16.85±1.53mm, 19.17±1.48mm dan 24.19±2.61mm (purata ± S.D.). Dalam usia ini, larva telah mempunyai mata berpigmen. Nisbah cone-nuclei bertambah dengan pertumbuhan larva. Nisbah mencapai 1.0 semasa larva berusia 20 hari. Peningkatan nisbah cone-nuclei menunjukkan kewujudan sel rod di dalam retina keli Afrika. Peningkatan ketebalan lapisan nuklear luar dengan pertumbuhan larva menunjukkan peningkatan dalam kepadatan rod. Sebagai hasil daripada pertambahan kelebaran lapisan pigment epithelium, lapisan sel penglihatan dan lapisan nuklear luar. kawasan dasar menjadi lebih tebal berbanding dengan kawasan lain di dalam retina kerana nuklear sel penglihatan di kawasan dasar menjadi lebih panjang. Berkait dengan kelakuannya, larva keli Afrika sentiasa tinggal di kawasan tengah dan dasar tangki. Mereka menunjukkan fototaksis negatif. Kewujudan sel rod menunjukkan larva keli Afrika lebih suka tinggal di persekitaran yang gelap. Pertumbuhan organ olfaktori meningkat dengan pertumbuhan larva menunjukkan larva keli Afrika bertindak balas dengan rangsangan kimia pada peringkat awal larva. Larva menunjukkan kelakuan kemotaksis yang kuat apabila larva diberi makan dengan serbuk makanan tiruan. Reseptor perasa dijumpai di bahagian barbel, mulut, rongga mulut dan di bahagian ranker insang menunjukkan larva keli Afrika dapat memperbaiki kelakuan pemakanannya kerana reseptor perasa membenarkan larva untuk memilih antara jenis makanan. Larva keli Afrika juga menunjukkan reotaksis positif yang kuat. Pertumbuhan deria organ yang disertai dengan perubahan kelakuan membawa implikasi penting untuk ekologi larva di habitat semulajadinya dan pengkulturan larva keli Afrika.



### ABSTRACT

### DEVELOPMENT OF SENSORY ORGANS IN AFRICAN CATFISH, Clarias gariepinus DURING LARVAL TO JUVENILE STAGE

This study was conducted to clarify the development of the sensory organs with larval growth in African cafish, Clarias gariepinus for the purpose of understanding its ecology as well as to improve the larval rearing techniques. Eggs were collected from Borneo Marine Researched Institute and the larvae of African catfish were reared from hatching to juvenile stage, sampled every day and observed under light microscope for the development of the sensory organs. Larvae were fed with rotifers, Artemia nauplii and artificial powder feed. The 20, 25 and 30-day-old larvae were 16.85±1.53mm, 19.17±1.48mm and 24.19±2.61mm (mean ± S.D) in total length (TL). During these days after hatching, the eye already deeply pigmented. The nucleicone ratio was increase with larval growth. The ratio exceeds 1.0 when the fish were 20-day-old. The cone-nuclei ratio was increase showed that the appearance of rods cell in the retina of the African catfish. The thickness of the outer nuclear layer in the retina also increases with larval growth showing an increase in rod density. As a result of the width added from the pigmented epithelium layer, visual cell layer and outer nuclei layer, the bottom area became thicker than other areas of the retina because the visual cell nuclei of the bottom area became longer than those in other areas. Related to its behaviour, African catfish larvae mostly stayed at the middle and bottom part of the tanks and they showed negative phototaxis. The appearance of rods cell indicates that the larvae of African catfish preferred to stay in dark environment. The development of olfactory organs increased with larval growth showed that African catfish response to chemical stimuli in the early larval stage. They showed strong chemotaxis behaviour when they were fed with artificial feed powder. The taste buds were found in barbels, mouth, buccal cavity and gill rankers showed that the African catfish larvae can improve their feeding behaviour because taste buds allow larvae to choose among food items. The African catfish larvae also showed the positive rheotaxis behaviour in this larval stage. The development of the sensory organs is accompanied by behavioural changes that have important implications for larval ecology at their natural habitat and African catfish larval rearing.



## CONTENTS

Pages

TITLE	
DECLARATION	ii
AUTHENTICATION	iii
ACKNOWLEDGEMENT	iv
ABSTRAK	v
ABSTRACT	vi
CONTENTS	vii
LIST OF TABLES	x
LIST OF FIGURES	xi
LIST OF PHOTOS	xiii
LIST OF ABBREVIATIONS	xiv
CHAPTER 1 INTRODUCTION	
1.1 Aquaculture Industry in Malaysia	1
1.2 African catfish, Clarias gariepimus	2
1.3 Problems on Larval Rearing of African Catfish	4
1.4 Studies on Sensory Organ and Development and B	ehaviour 5
Changes in African Catfish Juveniles	
1.5 Objectives of Study	6
CHAPTER 2 LITERATURE REVIEW	
2.1 Sensory System	7
2.1.1 Eyes	7
2.1.2 Olfactory Organ	9
2.1.3 Taste Buds	10
2.1.4 Lateral Line	11
2.1.5 The Inner Ear	13
CHAPTER 3 MATERIALS AND METHODS	
3.1 Culturing African catfish in UMS hatchery	14
3.1.1(A) Brood Fish Selection	14
3.1.1(B) Brood Fish measurement	15
3.1.2 Hormone Injection	18
3.1.3 Egg Stripping	18
3.1.4 Eggs Incubation	21
3.1.5 Larval Rearing	22
3.2 Sampling and Measuring	23
3.3 Observations	24
3.3.1 Morphology Observation	24
3.3.2 Behaviours Observation	24
3.4 Histology Experiment	25
3.4.1 Fixation	25
3.4.2 Dehydration	26
3.4.3 Clearing	27
3.4.4 Impregnation	28
3.4.5 Embedding	30



	3.4.6	Blocking	31
	3.4.7	Cutting	32
	3.4.8	Staining	33
	3.4.9	Mounting	34
CHAI	PTER 4	RESULTS	
4.1	Morph	ological changes and growth of African catfish	35
4.2	Behav	ioural Changes	37
4.3	Develo	opment of Sensory Organs	40
	4.3.1	Eye	40
	4.3.2	Olfactory organ	46
	4.3.3	Taste Buds	49
CHA	PTER 5	DISCUSSION	
5.1	Morph	nological Changes	53
5.2	Chang	ges of Behaviours	55
	5.2.1	Phototaxis	55
	5.2.2	Chemotaxis	56
	5.2.3	Rheotaxis	57
5.3	Devel	opment of Sensory Organs	58
	5.3.1	Eyes	58
	5.3.2	Olfactory Organ (Chemoreceptor)	59
	5.3.3	Taste Buds (Chemoreception)	60
CHA	PTER (	6 CONCLUSION	62
REF	ERENC	CES	64
APP	ENDIX		75



# LIST OF TABLES

No. Table		Pages
3.1	Selected brood fish measurement.	15
3.2	Dosage used for males brood fish.	17
3.3	Dosage used for females brood fish.	17
3.4	The amount of eggs from three female brood fish	20
3.5	The total number of eggs from three female broodfish.	20
3.6	Fertilization rate and hatching estimation.	20
4.1	Diagram of the relationship between the behavioural changes and the ontogenic development of the sense organs of African catfish, <i>Clarias gariepinus</i> larvae.	52

• • •



## **LIST OF FIGURES**

а ....

# No. Figure

4.1	Growth of the African catfish, Clarias gariepinus larvae.	35
4.2	Schematic diagram of behaviour in 20 to 30-day-old larvae of	38
	African catfish, Clarias gariepinus.	
4.3	Schematic diagram of behaviour in 20 to 30-day-old of African	39
	catfish.	
4.4	Eye diameter of African catfish.	40
4.5	The ratio of eye diameter with total length of African catfish.	41
4.6	Development of retina in African catfish. A, 20-day-old;	42
	B, 25-day-old; C, magnification from B; D, 30-day-old.	
4.7	Width of retina layers of the eyes of African catfish.	43
4.8	Width from the pigmented epithelium to outer nuclear layer	. 44
	of the eyes of African catfish.	
4.9	Change in the nuclei-cone ratio in the retina of African catfish.	45
4.10	Change in thickness of outer nuclear layer in the retina of	45
	African catfish.	
4.11	Development of olfactory organ in African catfish.	46
	A, 20-day-old; B, 25-day-old; D, 30-day-old. Scale bar,	
	A, 50 μm; B-C, 100μm.	
4.12	The number of lamellae of African catfish.	47
4.13	Width of olfactory pit of the African catfish.	48
4.14	Sum of lamellae length of African catfish.	49
4.15	The taste buds of 30-day-old African catfish, Clarias	50
	gariepinus larvae.	
4.16	The taste buds of 20-day-old African catfish, Clarias	51
	gariepinus larvae.	



# LIST OF PHOTOS

No. Photo		Pages
1.1	African catfish, Clarias gariepinus.	2
2.1	Eye, transverse section = Scale bar, $335\mu m$	7
2.2	Olfactory organ in fish.	9
2.3	Section of taste buds in epithelium tissues.	10
2.4	Free neuromast from lateral line system.	11
3.1	Brood fish selection.	14
3.2	Hormone injection under the pectoral fin.	16
3.3	Eggs stripping from female brood fish.	18
3.4	The distribution of fertilized eggs.	21
3.5	The 10-tonne tank is using for larval rearing.	22
3.6	Samples for histological experiment.	26
3.7	The different percentage of alcohol.	26
3.8	Xylene solution for clearing.	27
3.9	The wax bath machine used for melting the paraffin wax.	28
3.10	The equipment used for the embedding process.	30
3.11	Specimens were attached to wooden block.	31
3.12	The microtome used to make cross-section ribbons.	32
3.13	A series of reagents involved in the staining process.	33
3.14	The prepared mounted slide glasses ready to be observed.	34
4.1	20-day-old larvae of African catfish. Scale bar, 1mm	36
4.2	25-day-old larvae of African catfish. Scale bar, 1mm	36
4.3	30-day-old larvae of African catfish. Scale bar, 1mm	37



# LIST OF ABBREVATIONS

	an arrest the second second
cm	centimetre
μm	micrometre
d	day
dAH	days after hatching
DO	dissolved oxygen
kg	kilogramme
g	gramme
x	multiply
$\lambda_{\max}$	wavelength maximum
L	litre
ml	mililitre
mm	millimetre
no.	number
ppt	part per thousand
TL	total length
BL	body length
SL	standard length
HL	head length
BH	body height
BR	body round
BWidth	body width
BWt	body weight
1	per
S.D	standard deviation
μm	micrometre
%	percentage
°C	degree celcius
±	more or less
IUkg <sup>-1</sup>	international unit per kilogramme
ONL	outer nuclear layer
INL	inner nuclear layer
PE	pigmented epithelium
PR	photoreceptor cell
L	lens
hc	horizontal cell
ipl	inner plexiform layer
gl	ganglion layer
vc	visual cell



#### **CHAPTER 1**

#### INTRODUCTION

### 1.1 Aquaculture Industry in Malaysia

Aquaculture industry in Malaysia considered as new, contributing nearly 12% of total landings. Production from the aquaculture was recorded at 169,449 metric tonnes valued at RM 1,047.2 million in 2001. The production is from a culture area of 23,227 ha. In Third National Agricultural Policy (NAP3) it is expected that the production would increase to 600,000 metric tonnes by 2010. To achieve this target aquaculture has to be streamlined in terms of investment of capital, technology and human resources. Thus, in NAP3 greater attention has been to aquaculture production through commercialization of the sector with private and corporate sectors participation.

The environmentalists have constantly criticized aquaculture development as a sector of non-sustainable and environmental degradation. So, the industry possess critical constrains in its development. The key issues are limitation of land resources and the water bodies as uses for the other purposes. Other issues are pollution and environmental degradation, healthy and quality of seeds, disease, high production costs, indiscriminate use of chemicals, drugs and antibiotics. Government has identified certain areas for sustainable aquaculture production and makes it



environmental friendly. These are, (i) zoning of potential aquaculture areas (AIZ), (ii) legislation for aquaculture, (iii) guidelines for aquaculture development, (iv) code of practice for aquaculture, (v) farm accreditation and (vi) better extension services.

### 1.2 African Catfish, Clarias gariepinus



Photo 1.1: African catfish, Clarias gariepinus.

Catfishes constitute a large group of primarily freshwater fishes which are widely distributed throughout the world. They reach their greatest diversity in the continents spanning the equator, namely South America, Africa and Asia. They are especially diverse in the larger rivers such as the Amazon and Zaire, in each of which several hundred species are found. Catfish are classified in the order Siluriformes which includes such familiar fishes as the bullheads, squeakers, electric catfish, sea barbel and armoured catfishes, as well as less familiar forms such as the doradids, plotosids,



pimelodids and callichthyids. There are about 2000 species of catfishes in the world (about 8% of the total number of fishes). Most African catfishes are either too small or too difficult to culture or would encounter too much consumer resistance to be successful aquaculture candidates.

There are only three African Siluroidea families which contain some species which could be considered suitable for food fish aquaculture; one of which is *Clarias gariepinus*. The distinguishing characteristics of *Clarias gariepinus* are: Head large and bony with small eyes. Dorsal and anal fins long. No adipose fin. Pectoral fin with stout serrated spine, used for defence or "walking" overland. Mouth terminal, large. Four pairs of barbels present. Colour varies from sandy-yellow through gray to olive with dark greenish-brown markings, belly white. Well developed suprabranchial organ present. The principal feature which distinguishes *Heterobranchus longifilis* from *Clarias gariepinus* is its prominent adipose fin.

*Clarias gariepinus* can best described as an omnivore, often feeding on vegetable matter, aquatic invertebrates, small fish and detritus. Though it normally survives on dissolved oxygen, it comes to the surfaces and breathes atmospheric air when the oxygen concentration becomes low. A high degree of hardiness, the ability to feed on a variety of foodstuff, good growth and survival in poorly oxygenated water have made it an attractive fish for aquaculture. It is also resistant to diseases. So, the survival rate of this species is considered high but it is depends on environmental factors and cultural methods.



The species can also grow in brackish water in salinities of 10 ppt and survive in salinities up to 29 ppt. The species is one of the well-known freshwater species in Malaysia. It tastes good and Malaysian of all races, like to eat this fish. It can be cooked in Malay, Chinese and Indian dishes.

### 1.3 Problems on Larval Rearing of African Catfish

Although African catfish is suitable for mass production, there are several problems faced in culturing it. Main problem is they show cannibalism behaviour. This problem occurred because of feed sufficiency. When there are not enough feed, they will eat their own kind. This caused the amount of fish cultured to decrease and the percentage of survival rate does not high. So, the production of the seeds can not be maximised. It is important to improve larval rearing methods in African catfish to develop the aquaculture industry.

Other problem is African catfish faced the air bubbles in their stomach. It happens in the juvenile stage of African catfish. The air in the stomach can see be seen clearly. Their swimming behaviour would change. The air in the stomach prevent them to swim as usual because they had lost their balance in swimming. They gather at the surface of the tank and their activity will decrease. So, we have to find a way how to prevent this problem. We have to study the morphological features, their behaviour and also their sensory organs to successfully culture of this species.

In larval to juvenile stage, they are very sensitive and easily die. It is called critical period for the fish. We called this stages is critical period because during this



time their organs growth rapidly. Their organs fully develop during this period. They easily get stress because of handling, feeding management or water management. So, the suitable condition is very important to maintain the health of the fish. Regular observation must be done to ensure the condition of the fish.

# 1.4 Study on Sensory Organ and Development and Behaviour Changes in African Catfish Juveniles

This study is conducted to find possible relationships between developmental change in morphology of sense organs and behaviour during early stages of African catfish, *Clarias gariepinus*.

Accompanying changes in behaviour such as feeding, schooling, phototactic and rheotactic behaviour, of the fish with growth were observed. For the histological observation of sense organs, the fish were fixed in Bouin's solution. Employing standard methods, embedded fish were cut 6  $\mu$ m thick transversally and longitudinally. All sections were stained by standard method and observed under a microscope. This study is supported by The Ministry of Higher Education under Fundamental Research Grant. The research title: Studies on development of sensory organs and behavioural changes in the larvae of Grouper and other species of aquaculture importance. The project code is FRG0002-ST-1/2006.



All the three factors are correlative; behavior, sensory organ and morphological development. The objectives are:

- 1) This study is conduct to understand the morphological changes in African catfish juveniles.
- 2) To understand how behaviour changes with larval growth.
- To study the function of sensory organ in African catfish that is related to its behavior.



### **CHAPTER 2**

### LITERATURE REVIEW

2.1 Sensory system

### 2.1.1 Eyes



**Photo 2.1**: Eye, transverse section (Formalin, H&E, Bar = 335). 1, cornea; 2, lens; 3, iris; 4, retina; 5, optic nerve; 6, choroid body; 7, sclera. Scale bar, 335µm

The eyes of most teleosts are oriented and desirable so that light can enter from almost any direction, above, below, forward, or behind. In front, a fish view an object with



both eyes to create binocular vision and improved depth perception. Many species can use ocular movements to adjust size of the binocular field. Most fishes can see objects with only one eye, where depth perception is personably compromised.

Light entering the eye is focused on the retina, a spherical lens that moves away from or toward retina to accommodate objects near or far. The central circuitry from the brain extends through the optic nerve to the retina, which is composed of layer distinctive neural cells (ganglion, bipolar, photoreceptors). The major classes of photoreceptors are cones and rods. Vision under bright conditions (photopic vision) immediated by cones, and rods take over the visual function when light levels are low (scotopic vision). Both types of photoreceptors operate by photoactivation of visual pigments. Visual pigments generally respond to a broad range of wavelengths but have a peak of sensitivity, identified as the wavelength of maximum absorption,  $\lambda_{max}$ . up to four classes of visual pigments, each with a different  $\lambda_{max}$  can be found in the different types of cones. This is the basis for color vision. Species with a single pigment discriminate objects from the background by differences in contrast; wavelengths closer to their  $\lambda_{max}$  appear brighter than other wavelengths.





Photo 2.2: Olfactory organ in fish

The olfactory organ in fishes, as in other vertebrates, is located in the snout, where water, often aided by a groove or skin flap, passes over receptors in the epithelium, which is arranged in folds or lamellae, often resembling a rosette (Kleerekoper, 1969). Water passes through the organ either by movements of cilia located on supporting cells in the ephithelium or by muscular pumps associated with accessory sacs (Bardach and Villars, 1974; Doving *et al.*, 1977). The ultrastructure of the epithelium, bulb, and tract of the olfactory system is reviewed in detail by Hara (1993). For teleosts larvae, several studies have demonstrated that the receptor cells in the olfactory epithelium develop quickly (summarized by Doving and Knutsen, 1993) The sensory regions of the epithelium, in the majority of teleosts studied, contain at least two types of bipolar neurons, ciliated and microvillous, which are classified on the basis of their dendrites.



Differences in physiological function among the cell types are unclear. The axons of these bipolars, forming the olfactory nerve fibers, lead to the rostral end of the olfactory bulb, where they spread across its surface. The bulb, thought to process the bulk of olfactory signals (Bruckmoser, 1973), projects via olfactory tract to the telencephalon and minor components to the ventral diencephalon (Finger, 1988). Olfactory acuity appears to be correlated with the numbers and arrangement of olfactory lamellae in the nasal as well as the area covered by sensory supporting units in the epithelium (Bardach and Villars, 1974).

### 2.1.3 Taste buds



Photo 2.3: Section of taste buds in epithelium tissues.

Fish taste buds, located in the epidermis; vary between 30 and 80  $\mu$ m in length and 20 and 50  $\mu$ m in width. Generally, two types of microvillous cells, along with a basal cell, are routinely described in the taste buds of teleosts. One of the former, the so-called light cell, possesses a large club-shaped microvillus at its apical surface and is generally accepted as a gustatory receptor cell because of its synaptic contacts with



nerve processes within the taste bud (Caprio, 1988). The other type of cell, the socalled dark cell, possesses the numerous small microvilli and has been considered a supporting cell. Reutter (1978) found, however, that dark cells in catfishes also form synapses with nerve terminals within taste buds. Basal cells, two to five per taste bud, are located in the base of the bud, resting against the basal lamina. Their function is unknown, but they may identify gustatory receptor activity through synapses with both light and dark cell, at least in ameirid catfishes (Reutter, 1982).

Although the gross morphology of the taste buds of fishes is similar, caution about generalizations regarding vertebrate taste buds is suggested based on differences in their ultrastructure (Reutter and Witt, 1993). Taste buds are generally distributed within the oral cavity, pharynx, and gill rakers.

### 2.1.4 Lateral line



Photo 2.4: Free neuromast from lateral line system.



#### REFERENCES

Arnold, L. R. 1974. Rheotropism in Fishes. Biol. Rev. 49: 515-576.

- Atema, J. 1977. Functional Separation of Smell and Taste in Fish and Crustaceans. In: Olfaction and Taste 6: 165-174
- Audrey, D.T., 2007. Development of Sensory Organs and Changes of Behaviour in Larvae of Mouse Grouper, Cromileptes altivelis. Borneo Marine Research Institute, Universiti Malaysia Sabah, Kota Kinabalu, Sabah. (MSc. Thesis).
- Bagarinao, T., and Hunter, J.R. (1983). The visual feeding threshold and action spectrum of northen anchovy (Engraulis mordax) larvae. Calif. Coop. Oceanic Fish. Invest. Rep. 24: 245-254.
- Baidya A. P., 2004. Oocyte Final Maturation, Egg Development and Effects of Stripping Timings on Egg Quality in Pangasius hypophthalmus and Clarias gariepinus. Borneo Marine Research Institute, Universiti Malaysia Sabah, Kota Kinabalu, Sabah. (Ph.D Thesis).
- Balon, E.K. (ed.) 1985. Early life histories of fishes: New developmental, ecological and evolutionary perspectives. Dev. In Env. Biol. Fish. 5, Dr W. Junk Publishers, Dordrecht. 280 pp.



- Bardach, J.E., and Villars, T. (1974). The chemical senses of fishes. In: "Chemoreception in marine organisms." (P.T. Grant and A.M. Mackie, eds.), pp. 49-104. Academic Press, New York.
- Blaxter, J.H.S. (1986). Development of sense organs and behaviour of teleost larvae with special reference to feeding and predator avoidance. Trans. Am. Fish. Soc. 155: 98-114
- Blaxter, J.H.S. (1987). Stucture and development of the lateral line. Biol. Rev. 62: 471-514.
- Blaxter, J.H.S. and Fuiman, L.A., 1989. Function of the free neuromasts of marine teleost larvae. In: The Mechanosensory Lateral Line: Neurobiology and Evolution, Coombs, S., Goner, P. and Munz, H., eds. Springer-Verlag, New York, pp. 481-499.
- Bond C. E. 1996. Biology of Fishes. 2<sup>nd</sup>. Ed. Sounders College Publishing. pp. 519-520.
- Bond C. E. 1996. Biology of Fishes. 2<sup>nd</sup>. Ed. Sounders College Publishing. pp. 524-525.
- Bruckmoser, P. (1973). Beziehungen zwischen Stuktur und Funktion in der Evolution des Telencephalon. Verh. d. Deutschen Zool. Ges. 66: Jahresvers. 219-229.



- Caprio, J. (1988). Peripheral filters and chemoreceptors cells in fishes. In "Sensory biology of aquatic animals." (J. Atema, R.R. Fay, A.N. Popper, and W.N. Tavolga, eds.), pp. 313-338, Spinger-Verlag, New York.
- Cobcroft, J. M. and Pankhurst, P. M. 2003. Sensory Organ Development in Cultured Striped trumpeter larvae, *Latris lineate*: Implication for Feeding Behaviour. *Marine and Freshwater Research* 54: 669-692
- Colson, D.J., Patek, S.N., Brainerd, E.L., and Lewis, S.M. (1998). Sound production during feeding in *Hippocampus* seahorses (Syngnathidae). *Env. Biol. Fishes* 51: pp. 221-229
- Corwin, J.T. (1981a). Audition in elasmobranches. In: "Hearing and sound communication in fishes." (W.N. Tavolga, A.N. Popper and R.R. Fay, eds.), pp. 81-102, Springer-Verlag, New York.
- Corwin, J.T. (1981b). Peripheral auditory physiology in the lemon shark: Evidence of parallel otolithic and non-otholitic sound detection. J. Comp. Physiol. 142A: 379-390.
- Corwin, J.T., (1989). Functional anatomy of the auditory system in sharks and rays. J. Exp. Zool. Suppl. 2: 62-74.
- Culling, C.F.A., 1974. Handbook of Histopathological and Histochemical Techniques (including museum techniques) Third Edition. Butterworth & Co. (Publishers) Ltd., Great Britain. Pg 29-220.



- de Graaf, G., and Janssen, H., 1996. Artificial Reproduction and Pond Rearing of the African Catfish <u>Clarias gariepinus</u> in Sub – Saharan Africa – A Handbook. Food and Agriculture Organization, Netherlands.
- Disler, N.N., 1971. Lateral Line Sense Organs and their Importance in Fish Behavior (translated by Mills, H. and Yariv, M.), Israel Program for Science Translation, Jerusalem.
- Diyana, R., 2007. The Development of Sensory Organs and Changes of Behaviour in Larvae of Patin, Pangasius hypophthalmus. Borneo Marine Research Institute, Universiti Malaysia Sabah, Kota Kinabalu, Sabah.
- Doving, K.B., Dubois-Dauphin, M., Holley, A., and Jourdan, F. (1977). Functional anatomy in the olfactory organs of fish and the ciliary mechanisms of water transport. Acta Zool. (Stockh.) 58: 245-255.
- Doving, K.B., and Knutsen, J.A. (1993). Chemokinesis in marine fish larvae. In: "Physiological and biochemical aspects of fish development." (B.T. Walther and H.J. Fyhn, eds.), pp. 139-145. university of Bergen, Norway.
- Fine, M.L., Winn, H.E., and Olla, B. (1977). Communication in fishes. In: "How animals communicate" (T.A Sebeok, ed.), pp. 472-518. Indiana Univ. Press, Bloomington.



Finger, T.E. (1988). Organization of chemosensory systems within the brains of bony fishes. In: "Sensory biology of aquatic animals." (J. Atema, R.R. Fay, A.N. Popper, and W.N. Tavolga, eds.), pp. 339-363. Springer-Verlag, New York.

- Fuiman, L.A., 2004. Changing structure and function of the ear and lateral line system of fishes during development. American Fisheries Society Symposium, 40: pp. 117-144.
- Foto sensory organ ; Source from Miss Diyana bt Ramli. Free neuromast and olfactory organs of patin, *Pangasius hypopthalmus*, Borneo Marine Reseach Institute, University Malaysia Sabah, Kota Kinabalu, Sabah. Pp. 46-49.
- Gozlan E. Rodolphe, Gordon H. Copp and Jean-Noel Tourenq, 1999. Comparison of
   Growth Plasticity in the Laboratory and Field, an Implications for the Onset of
   Juvenile Development in Sofie, Chondrostoma toxostoma. Journal of
   Environmental Biology of Fishes 56: 153-165
- Guthrie, D. M., 1986. Role of Vision in Fish Behaviour, p. 75-113. In: The Behaviour of Teleost Fishes, T. J.Pitcher (ed.). Johns Hopkins Univ. Press, Baltimore
- Hara, T.J. (1993). The role of olfaction in fish behaviour. In: "The behaviour of teleost fishes." (T.J. Pitcher, ed.), pp. 171-199. Chapman and Hall, New York.



- Huet, M., 1995. Faizah Shaharom, Hassan Hj. Mohd Daud, and Siti Khadijah Daud (translator). Buku Teks Mengkultur Ikan Pembiakan dan Pemeliharaan Ikan. Dewan Bahasa dan Pustaka, Kuala Lumpur. Pg: 230 – 238.
- Iwai, T. (1965). Notes on the cupulae of free neuromasts in larvae of the goldfish. Copeia 1965:379.
- Iwai, T. (1967). Structure and development of lateral line cupulae in teleost larvae. In: "Lateral Line Detectors." Ed by P. Cahn, Indiana Univ. Press, Bloomington, pp 27-44.
- Iwai, T., 1972a. Feeding of teleost larvae: a review. La Mer, 10: pp. 71-82.
- Iwai. T., 1972b. on the free neuromasts of some teleost larvae. Japanese Journal of Ichthyology, 19: pp. 307-311.
- Iwai, T. 1980. Sensory Anatomy and Feeding of Fish Larvae. In: Fish Behaviour and its use in the Capture and Culture Species. Bardach, J. J. Magnuson, R. C. May and J. M. Reinhart.) pp. 124-145. (International Center for Living Aquatic Resources Management: Manila, Philippines.)
- Jones, W.R and Janssen, J., 1992. Lateral line development and feeding behaviour in the mottled sculpin, *Cottus bairdi (Scorpaeniformes: Cottidae)*. Copeia, 1992: pp. 485-492.



Kawamura, G. 1984. The Sense organs and Behaviour of Milkfish Fry in Relation to collection Techniques In: Juarian, J. V., Ferraris, R. P. and Benitez, V. (eds). Advance in Milkfish Biology and Culture. Island Publishing House, Manila. 69-85.

- Kawamura, G. and Ishida, K. 1984. Changes in Sense Organ Morphology and Behaviour with Growth in the Flounder, *Paralichthys olivaceus*. Bulletin of the Japanese Society of Scientific Fisheries 51(2): 155-165.
- Kawamura, G. and Washiyama N. 1989. Ontogenic Changes in Behaviour and Sense Organ, Morphogenesis in Largemouth Bass and *Tilapia nilotica*. *Transactions* of the American Fisheries Society **118**: 203-21.
- Kawamura, G., Masuma, S., Tezuka, N., Koiso, M., Jinbo, T. and Namba, K. 2003.
  Morphogenesis of Sense Organs in bluefin tuna, *Thunnus orientalis*. In: Haward, I., Anne, B.S. (eds). Proceedings of the 26<sup>th</sup> annual larval fish conference, 123-135.
- Kleeropter, H. 1969. Olfaction in Fishes. Indiana University Press, Bloomington.
- Kozloski, J., and Crawford, J.D. (1998). Functional neuroanatomy of auditory pathways in the sound-producing fish *Pollimyrus*. J. Comp. Neurol. 401: 227-252.
- Ladich, F. (1997). Agonistic behaviour and significance of sounds in vocalizing fish. Mar. Fresh. Behav. Physiol. 29: 87-108



- Liang, X.F., Liu, J. K., and Huang B. Y. 1998. The Role of Sense Organs in the Feeding Behaviour of Chinese Perch. Department of Biology, Jinan University, Guangzhou, China.
- Lyon, E.P. 1904. On Rheotropism in Fishes. Am. J. Physiol. 12: 149-161
- Marshall, N. B. 1966. The Life of Fishes. Universe Books, New York.
- Mukai, Y., Kobayashi H., Yoshikawa, H. (1992). Development of free and canal neuromasts and their directions of maximum sensitivity in the larvae of ayu, *Plecoglossus altivelis. Japan J. Ichthyol* 38: 411-417. (In Japanese with English abstract.
- Mukai, Y. and Kobayashi, H., 1994. Development of free neuromasts in larvae cyprinid fish. *Memoir of Faculty of Agriculture*, Kinki University, 27: pp. 1-14.
- Mukai, Y., Yoshikawa H., Kobayashi H. (1994). The relationship between the length of the cupulae of free neuromasts and feeding ability in larvae of the willow shiner, Gnathopogon elongates caerulescens (Teleostei, Cyprinidae). J. Exp. Biol. 197: 399-403.
- Mukai, Y., 2006. Role of free neuromasts in larval feeding of willow shiner Gnathopogon elongates caerulescens Teleostei, Cyprinidae. Fisheries Science, 72, pp. 705-709.



- Myrberg, Jr., A.A. (1981). Sound communication and interception in fishes. In " Hearing and sound communication in fishes." (W.N. Tavolga, A.N. Popper and R.R. Fay, eds.). pp. 395-425. Springer-Verlag, New York.
- Omura, Y., Shiozawa, S. and Tabata, K. 2004. proliferation of Rod cell on the Mature retina of Japanese Flounder Paralichthys olivaceus. Fisheries Science 70: 80-86.
- Platt, C. (1988). Equilibrium in the vertebrates: signals, senses, and steering underwater. In: "Sensory biology of aquatic animals". (J. Atema, R.R. Fay, A.N. Popper, and W.N. Tavolga, eds.), pp. 783-809. Springer-Verlag, New York.
- Fay, R.R., and Popper, A.N., (1980). Structure and function in teleost auditory systems. In: "Comparative studies of hearing in vertebrates" (A.N. Popper and R.R. Fay, eds.), pp. 3-42. Springer-Verlag, New York.
- Popper, A.N. (1977). A scanning electron microscopic study of the sacculus and lagena in the ears of fifteen species of teleost fishes. J. Morph. 153: 397-417
- Popper, A.N., and Coombs, S. (1980a). Acoustic detection by fishes. In "Environmental physiology of fishes." (M.A. Ali, ed.), pp. 403-430. Plenum Press, New York. (1983).



- Popper, A.N., and Coombs, S. (1980b). Auditory mechanisms in teleost fishes. Am. Sci. 68: 429-440.
- Popper, A.N. (1983). Organization of the inner ear and auditory processing. In: "Fish neurobiology and behaviour, vol. 1: brain stem and sense organs." (R.G. Northcutt and R.E. Davies, eds.), pp. 125-178. Univ. Mich. Press, Ann Arbor.
- Reutter, K. (1978). Taste organ in the bullhead (Teleostei). Adv. Anat. Embryol. Cell Biol. 55: 1-98.
- Reutter, K. (1982). Taste organ in the barbel of the bullhead. In: "Chemoreception in fishes." (T.J. Hara, ed.), pp. 77-91. Elsevier, New York.
- Reutter, K., and Witt, M. (1993). Morphology of vertebrate taste organs and thir nerve supply. *In*: "Mechanisms of taste transduction." (S.A. Simon and S.D. Roper, eds.), pp. 29-82. CRC Press, Boca Raton.
- Schneider, H. (1961). Neue Ergebnisse der Lautforschung bei Fischen. Naturwiss. 48: 513-518.
- Smith, R.J.F. (1992). Alarm signals in fishes. Rev. fish Biol. Fish. 2: pp. 33-63
- Sorensen, P. W. and Carpio 1998. Hormones, Pheromones and chemreception, p. 199-228. In: Fish Chemoreception, T. J. Hara (ed.). Chapman and Hall, London.



- Tagliafierro, G. and Zaccone, G. 2001. Morphology and Immuno histochemistry of Taste Buds in Bony Fishes. In: Kapoor, B. G. and Hara, T. J. (eds). Sensory Biology of Jawed Fishes: New Insight. Science Publishers, Inc. USA. 335-346.
- Tavolga, W.N. (1960). Sound production and underwater communication in fishes. In "Animal sounds and communication." (W.E. Lanyon and W.N. Tavolga, eds.), pp. 93-136. A.I.B.S., Washington, D.C.
- Wootton, R.J., 1992. Fish ecology. Black Academic & Professional, Great Britain. Pg 31 – 85.
- Yamamoto, M. 1982. Comparative Morphology of the Peripheral Olfactory Organ in Teleosts, p. 39-59. In: Chemoreception in fishes. T. J. Hara (ed.). Elsevier, Amsterdam.
- Webb, J.F. 1989. Gross morphology and evaluation of the mechanoreceptive lateralline system in teleost fishes. *Brain, Behav. Evol.* 33: pp. 34-53.

