

**PRODUCTION AND CHARACTERIZATION OF PU ERH TEA FROM LOCAL TEA LEAVES**

**WONG YIU NYUK**

**DISSERTATION SUBMITTED AS PARTIAL FULFILLMENT OF  
THE REQUIREMENTS FOR THE DEGREE OF  
BACHELOR OF SCIENCE WITH HONOURS**

**INDUSTRIAL CHEMISTRY PROGRAMME  
SCHOOL OF SCIENCE AND TECHNOLOGY  
UNIVERSITI MALAYSIA SABAH**

**2010**



**UMS**  
UNIVERSITI MALAYSIA SABAH

UNIVERSITI MALAYSIA SABAH



BORANG PENGESAHAN STATUS TESIS@

JUDUL: Production and Characterization of Pu Erh tea from local Tea leaves.

Ijazah: Bachelor's science (Honors)

SESI PENGAJIAN: 07/08

Saya WONG YU NYUK

(HURUF BESAR)

mengaku membenarkan tesis (LPS/Sarjana/Doktor Falsafah)\* ini disimpan di Perpustakaan Universiti Malaysia Sabah dengan syarat-syarat kegunaan seperti berikut:

1. Tesis adalah hakmilik Universiti Malaysia Sabah.
2. Perpustakaan Universiti Malaysia Sabah dibenarkan membuat salinan untuk tujuan pengajian sahaja.
3. Perpustakaan dibenarkan membuat salinan tesis ini sebagai bahan pertukaran antara institusi pengajian tinggi.
4. \*\*Sila tandakan ( / )

SULIT

(Mengandungi maklumat yang berdarjah keselamatan atau kepentingan Malaysia seperti yang termaktub di dalam AKTA RAHSIA RASMI 1972)

TERHAD

(Mengandungi maklumat TERHAD yang telah ditentukan oleh organisasi/badan di mana penyelidikan dijalankan)

TIDAK TERHAD

*[Signature]*

(TANDATANGAN PENULIS)

PERPUSTAKAAN UNIVERSITI MALAYSIA SABAH

Disahkan oleh

*[Signature]*

NURULAIN BINTI ISMAIL  
LIBRARIAN

(TANDATANGAN PUSKUTAWAN)

Alamat Tetap: Lot 160, Qg 107  
Kudat, Kluang, Sarawak

\_\_\_\_\_  
Nama Penyelia

Tarikh: 7/5/10

Tarikh: \_\_\_\_\_

- CATATAN:
- \* Potong yang tidak berkenaan.
  - \*\* Jika tesis ini SULIT atau TERHAD, sila lampirkan surat daripada pihak berkuasa/organisasi berkenaan dengan menyatakan sekali sebab dan tempoh tesis ini perlu dikelaskan sebagai SULIT dan TERHAD.
  - @ Tesis dimaksudkan sebagai tesis bagi Ijazah Doktor Falsafah dan Sarjana socara penyelidikan, atau disertasi bagi pengajian socara kerja kursus dan penyelidikan, atau Laporan Projek Sarjana Muda (LPSM)

PERPUSTAKAAN UMS



\* 1000353872 \*



## DECLARATION

I declare that the works presented in this dissertation are based on my own research. Sources of finding reviewed herein have been duly acknowledged.

09 APRIL 2010



---

WONG YIU NYUK

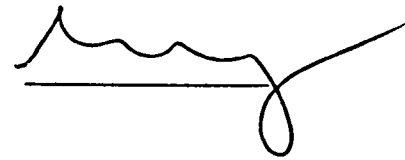
BS07110141

## VERIFICATION

Name           Wong Yiu Nyuk  
Title           Production and Characterization of Pu erh Tea from Local Tea Leaves

### SIGNATURE

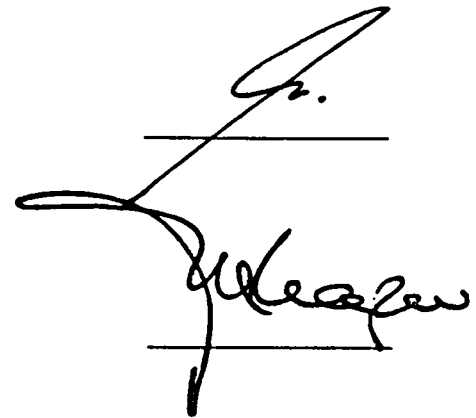
1. SUPERVISOR  
(ASSOC. PROF. DR. HOW SIEW ENG)



2. EXAMINER 1  
(DR. NOUMIE SURUGAU)



3. EXAMINER 2  
(MS. RUBIA IDRIS)



4. DEAN  
(PROF. DR. MOHD HARUN ABDULLAH)



## **ACKNOWLEDGEMENT**

This dissertation arose as a final year project to fulfil part of the requirement for Bachelor of Science degree in Industrial Chemistry. By the time, I have worked with a great number of people whose contribution in assorted ways to the research. It is a pleasure to express my gratitude to them all in my humble acknowledgement.

First of all I would like to express my sincere gratitude to my supervisor, Assoc. Prof. Dr. How Siew Eng who has provided a great help in constructive criticism on my study, valuable suggestions, continuous encouragement in supervising throughout the completion of the dissertation.

Other than that, I would also like to thank Mr. Khoo Yau Liang, Ms. Ch'ng Ai Ying, and Ms. Thong Wei Li for their valuable advices, teaching, and guidance in the screening assay.

Thirdly, I would like to thank my teamwork partners of Osadhi Natural Products Research Group and also my fellow coursemates (especially Ho Pui Shing, Lee Sun Nee, Lew Yih, Yap Phooi Yee) for their helps, supports and friendship.

Finally, this dissertation is dedicated to my father Wong Kong Jin and my mother Ling Moi Moi, for their love, financial supports and helps.



# PENYEDIAAN DAN PENCIRIAN TEH PU ERH TEMPATAN

## ABSTRAK

Teh Pu erh telah menjadi salah satu minuman kegemaran para pengguna disebabkan oleh ciri-ciri kesihatan seperti, keupayaan anti oksida, penurunan kolesterol, penurunan tekanan darah and perlindungan tisu berantara. Sejumlah empat teh Pu erh komersial (T31, T32, T33, T34) dibandingkan dengan teh buatan tempatan Pu erh muda and matang (T35,T36). Jumlah kandungan polifenol, flavonoid, aktiviti anti oksida, antimikrobial dan perencat kinase telah dinilai bagi setiap teh Pu erh. T36 mencatat peratusan hasil ekstrak yang paling tinggi manakala T34 mempunyai jumlah kandungan flavonoid yang paling tinggi dalam ekstrak dengan menggunakan cara  $AlCl_3$ . Nilai  $IC_{50}$  bagi T35 dan T36 dalam kajian ini ialah 0.032 mg/mL dan 0.034 mg/mL. Keputusan ini menunjukkan potensi teh Pu erh yang lebih tinggi berbanding dengan BHT (Butylates hydroxyl toluene) dalam penyekatan radikal bebas pada kepekatan 0.10 mg/mL. Semua teh Pu erh telah menunjukkan aktiviti antimikrobial terhadap *Staphylococcus aureus* and *Bacillus cereus*, *Vibrio parahaemolytics*, *Listeria Monocytogenes*. Manakala hanya the Pu erh muda dan matang buatan tempatan (T35, T36) menyekat penumbuhan *Enterobacter sakazaki*. Acid gallic, catechin, caffeine, EGCG, dan ECG telah dikesan dalam semua teh Pu erh muda (T31, T35, T36) dengan menggunakan RP-HPLC. Oleh itu, the Pu erh muda adalah sumber agen antioxidan yang kuat dan berpotensi untuk mencegah penyakit yang berkaitan dengan radikal bebas.

## ABSTRACT

Pu erh teas are of interest to the consumers because of its antioxidant properties, ability to reduce cholesterol, lower blood pressure and protect connective tissue. Four commercial Pu erh teas, raw Pu erh 2003 (T31), raw Pu erh 2006 (T32), mature Pu erh 2005 (T33) and mature Pu erh 2007 (T34) were used to compare with the freshly prepared raw and mature Pu erh produced by using Sabah tea leaves (T35, T36). The total polyphenol and flavanoid, DPPH radical scavenging, antimicrobial and anti kinases effects were determined to examine the tea infusion quality. T36 possessed the highest percentage yields of crude extract (6.048 g) and T34 possessed the highest total flavonoids content (AlCl<sub>3</sub> method) in crude extract in tea samples (12.9 mg QE/g). The IC<sub>50</sub> value of T35 and T36 on DPPH radical scavenging was found to be 0.032 mg/mL and 0.034 mg/mL respectively, which were significantly higher than Butylated hydroxyl toluene (IC<sub>50</sub> value 0.13 mg/mL). All the Pu erh tea extracts inhibited the growth of *Staphylococcus aureus* and *Bacillus cereus*, *Vibrio parahaemolyticus*, *Listeria Monocytogenes*. However, only freshly prepared raw and mature Pu erh tea (T35, T36) inhibited the growth of *Enterobacter sakazaki*. Gallic acid, catechin, caffeine, EGCG and ECG were detected in tea extracts of all the raw Pu erh (T31, T32, T35) as analyzed using RP-HPLC. Therefore, raw pu erh is a powerful source of antioxidant beverage and may be used in preventing free radical-related diseases.

# CONTENTS

	Page
DECLARATION	ii
VERIFICATION	iii
ACKNOWLEDGEMENT	iv
ABSTRAK	v
ABSTRACT	vi
LIST OF CONTENTS	vii
LIST OF TABLES	ix
LIST OF FIGURES	x
LIST OF PHOTOS	xi
LIST OF SYMBOLS AND ABBREVIATIONS	xii
<b>CHAPTER 1 INTRODUCTION</b>	<b>1</b>
1.1 Background study	1
1.2 Pu erh tea and health	2
1.3 Objectives	2
1.4 Scope of study	3
<b>CHAPTER 2 LITERATURE REVIEW</b>	<b>4</b>
2.1 Introduction	4
2.2 Taxanomy	5
2.3 Morphology	
2.4 Classification of Pu erh tea	7
2.5 Chemical composition of Pu erh tea and its bioactivity	9
2.5.1 Tea polyphenols	10
(a). Tea flavonoids	11
(b). Flavonols	13
(c). Theaflavins and Thearubigins	13
(d). Phenolic acid	15
2.5.2 Statins	16
2.5.3 $\gamma$ - aminobutyric acid GABA	16
2.5.4 Tea alkaloid	17
(a). Caffeine	17
(b). Theobromine and Theophylline	18
2.5.5. Theanine	20
2.6 Pu erh tea Manufacturing Process	21
2.7 Antioxidant Properties of Tea Polyphenol	22
2.7.1. Scavenging Reactive Oxygen Species	23
2.7.2. Antioxidant Properties of Pu erh tea	24
2.8 Antimicrobial activity of Pu erh tea	24
2.9 Tea polyphenol againts MAP kinase Signaling Pathways	25
2.10 Health effect of Pu erh tea	26
2.10.1. Anti-obesity	26
2.10.2 Inhibition of SARS-CoV 3C-like Protease Activity by Theaflavin-3,3'-digallate (TF3)	27



<b>CHAPTER 3 METHODOLOGY</b>	<b>28</b>
3.1 Sample collection	29
3.2 Sample drying	29
3.3 Fermentation	29
3.4 Sample extraction	31
3.5 Total polyphenols test	32
3.6 Total flavonoids test	32
3.7 Antioxidant test	32
3.8 Antimicrobial activity test	33
3.8.1 Preparation of media	33
3.8.2 Bacteria Culture Media	33
3.8.3 Fermentation of Bacterial Strains	33
3.8.4 Preparation of Discs	33
3.8.5 Antimicrobial screening	34
3.9 Antikinas activity test	34
3.9.1 MKK1 <sup>P386</sup> Screening System	34
(a) Cultivation of Yeast Strain H10068	35
(b) Fermentation of Yeast Strain H10068	36
(c) MKK1 <sup>P386</sup> Inhibitor Screening Assay	36
3.9.2 MKK1 <sup>P386</sup> -MSG5 Screening System	36
(a) Cultivation of Yeast Strain H10069	36
(b) Fermentation of Yeast Strain H10069	37
(c) MKK1 <sup>P386</sup> -MSG5 Inhibitor Screening Assay	38
3.10 RP-HPLC analysis	38
<b>CHAPTER 4 RESULT AND DISCUSSION</b>	
4.1 Sample extracts	40
4.2 Total polyphenols content	40
4.3 Total flavonoids content	42
4.4 Antioxidative activity	43
4.5 Antimicrobial activity	45
4.6 Antikinas activity	49
4.7 RP-HPLC analysis on polyphenols	51
<b>CHAPTER 5 CONCLUSION</b>	<b>56</b>
<b>REFERENCES</b>	<b>57</b>
<b>APPENDIX</b>	<b>58</b>
APPENDIX A	61
APPENDIX B	
APPENDIX C	
APPENDIX D	
APPENDIX E	
APPENDIX F	

## LIST OF TABLES

Table No.		Page
2.1	Classification of Pu erh tea	7
4.1	Percentage yield of tea extracts	40
4.2	Total polyphenols content in crude extract	41
4.3	Total flavonoids content in crude extract	42
4.4	Inhibition percentage (%) of free radical DPPH	44
4.5	IC <sub>50</sub> of free radical DPPH	44
4.6	Antimicrobial screening for freshly prepared Pu erh tea extracts	45
4.7	Antimicrobial screening for commercial Pu erh tea extracts	46
4.8	MKK1 <sup>P386</sup> screening for Pu erh tea extracts	50
4.9	MKK1- MSG5 screening for Pu erh tea extracts	50
4.10	Compounds detected in freshly prepared raw Pu erh tea extract (T35).	51
4.11	Compounds detected in freshly prepared mature Pu erh tea extract (T36).	52
4.12	Compounds detected in commercial raw Pu erh 03 (T31).	53
4.13	Compounds detected in commercial raw Pu erh 06 (T32).	53
4.14	Compounds detected in commercial mature Pu erh 05 (T33).	54
4.15	Compounds detected in commercial mature Pu erh F8525 (T34).	55

## LIST OF FIGURES

Figure No.		Page
2.1	Chemical structures of flavonoids in tea	12
2.2	Structures of flavonols and flavonol glycosides	13
2.3	Structure of gallic acid	14
2.4	Structural formula of lovastatin	16
2.5	Structural formula of gamma-aminobutyric acid	17
2.6	Synthesis of caffeine from xanthosine	18
2.7	Bacterial and higher plant caffeine catabolism pathways	19
2.8	Schematic description of Pu erh tea manufacturing	21
4.1	HPLC chromatogram of freshly prepared raw Pu erh tea extract (T35)	51
4.2	HPLC chromatogram of freshly prepared mature Pu erh tea extract (T36)	52
4.3	HPLC chromatogram of commercial raw Pu erh 03 (T31)	52
4.4	HPLC chromatogram of commercial raw Pu erh 06 (T32)	53
4.5	HPLC chromatogram of commercial mature Pu erh 05 (T33)	54
4.6	HPLC chromatogram of commercial mature Pu erh F8525(T34)	54

## LIST OF PHOTOS

Photo No.		Page
2.1	Fresh tea leaves ( <i>C. sinensis</i> ) in Sabah tea plantation	6
3.1	Yunnan Chi Tse Beeng Cha (2003)	28
3.2	Yunnan Chi Tse Beeng Cha (2006)	28
3.3	Fengpai Qizi Beng Cha(2007)	29
3.4	Jin Gua Gong Cha (2005)	29
3.5	Fermentation of Pu erh tea on straw mat	30
3.6	Freshly prepared mature Pu erh tea sample	30
3.7	Freshly prepared raw Pu erh tea sample	31
3.8	Rotary evaporator	31
3.9	High performance liquid chromatography (HPLC)	31
4.1	Screening for <i>S.aureus</i> inhibitors from the Pu erh tea crude extracts	47
4.2	Screening for <i>Vibrio parahaemolyticus</i> inhibitors from the Pu erh tea crude extracts	47
4.3	Screening for <i>Bacillus cereus</i> inhibitors from the Pu erh tea crude extracts	48
4.4	Screening for <i>Enterobacter sakazaki</i> inhibitors from the Pu erh tea crude extracts	48
4.5	Screening for <i>Listeria monocytogenes</i> inhibitors from the Pu erh tea crude extracts	49
4.6	Screening for MKK1 inhibitors from Pu erh tea extracts	50

## LIST OF SYMBOLS AND ABBREVIATIONS

=	Equal
>	More than
<	Less than
±	Plus, minus
~	Approximately
%	Percentage
°	Degree
°C	Degree Celsius
AD	Anno Domini
BC	Before Christ
BOPF	Broken Orange Pekoe Fannings
cm	centimetre
Da	Dalton
DAD	Diode-Array Detector
etc	etcetera
g	gram
h	hour
i.e.	That is
kg	kilogram
LC <sub>50</sub>	Lethal Concentration of 50
mg	milligram
mg/g	milligram per gram
mg/mL	milligram per milli litre
min	minute
mL	milli Litre
mm	millimetre
nm	nanometre
ppm	Part per million
µg	microgram
µL	micro litre
N	North
S	South
US	United State

## CHAPTER 1

### INTRODUCTION

#### 1.1 Background study

Pu erh tea, a well-known traditional Chinese tea is originated from the districts of Xi-shuang-ban-na, Si mao, and Lan-chuang-jiang valley, Yunnan Province, China. The famous tea mountains in Yunnan are Youle Shan, Nannuo Shan, Yibang Shan and Bada Shan. There are two major types of Pu erh which are raw (sheng) and mature (shu) Pu erh, depending on the processing method and the degree of fermentation. Within the raw Pu erh category, we find various grades of loose leaf sun-dried green teas made with Yunnan's broad leaf varieties. Mature Pu erh's referred to as "the cooked type" due to the fact that dried green tea leaves are transformed via natural fermentation. Yunnan province produces the vast majority of pu erh tea. Tea mountains with adequate rainfall, good drainage and high elevation met the requirement for tea plantation (Graham, 1999). Sabah Mount Kinabalu which has similar climate and geographical condition with Yunnan tea mountains shows the potential of producing Pu erh tea from local tea leaves.

#### 1.2 Pu erh tea and health

Pu erh tea is of interest to the consumers because of its quality and health care function and distinct from other kinds of tea. Unlike other teas, the quality of Pu erh is not easy to evaluate, it is highly affected by the production process which involves fermentation. Sano *et al.* (1986) shows that longer the fermentation period of the Pu erh tea has better function in lipid reducing than the younger preparation.



Researches had been done on the polyphenol fraction in Pu erh tea on its health beneficial properties, such as suppressing fatty acid synthase expression (Chiang *et al.*, 2006), acting as an inhibitor of lipid and non-lipid oxidative damage (Duh *et al.*, 2004) and also exhibiting metal-binding ability, reducing power, scavenging effect for free radical (Jie *et al.*, 2006) and potential antimicrobial effect (Wu *et al.*, 2007).

### 1.3 Objectives

The production of high quality of Pu erh tea requires precise information on the diversity available and also the biochemicals which contributes toward the tea quality. Pu erh tea is characterized by physiochemical. The objectives of this study are:

- a) To produce Pu erh tea from local Sabah tea leaves.
- b) To analyse the polyphenol and flavonoids content of Pu erh tea produced from local Sabah tea leaves.
- c) To evaluate the antioxidant activity of Pu erh tea produced from Sabah tea leaves.
- d) To evaluate the antimicrobial activity of Pu erh tea against foodborne microbial.
- e) To evaluate antikinases inhibition activity of Pu erh tea.
- f) To compare the locally produced Pu erh tea with the Yunnan Pu erh tea leaves.

### 1.4 Scope of study

Fresh tea leaves were plucked from tea plants (*Camellia sinensis*) at Sabah Tea Plantation, Ranau, Sabah and were produced into raw (sheng) Pu erh and mature (shu) Pu erh. The milled leaves sample was extracted with 70% aqueous methanol (200 mL) (Sharma *et al.*, 2005). The extracted samples were cooled and filtered. The filtrate was evaporated with a rotary evaporator around 40 – 42 °C and then concentrated to powder using freeze dryer.

The sample extracts were analyzed to determine the total polyphenols and flavonoids content with classical Folin and Ciocalteu method (Turkmen *et al.*, 2006)

using gallic acid (standard phenolic compound) as a calibrant and  $\text{AlCl}_3$  colorimetric method (Huang *et al.*, 2006) respectively.

Three types of bioactivities of Pu erh tea tested included antioxidant, antimicrobial, and antikinase activity. The antioxidative activity was evaluated using a free radical 1,1-diphenyl-2-picrylhydrazyl (DPPH) inhibition assay (Katalinic *et al.*, 2006). The bacteria used in the antimicrobial test were *Staphylococcus aureus*, *Bacillus cereus*, *Salmonella typhimurium*, *Salmonella enteritidis*, *Enterobacter sakazaki*, *Listeria Monocytogenes*, *Yersina Enterocolitica*, *Vibrio parahaemolyticus* and *Escherichia coli*. Antikinases activity was elevated by MKK1<sup>P386</sup> screening system. The antikinases screening systems involved genetically modified yeast strains, targeting on protein kinases.

The main constituents of Pu erh tea, four major catechin, gallic acid and caffeine are analyzed by using HPLC (Zuo *et al.*, 2002).



## CHAPTER 2

### LITERATURE REVIEW

#### 2.1 Introduction

Tea was discovered around 5000 to 6000 years ago during the "Shen-Nong" era of ancient China (Zhen *et al.*, 2002). The centre of origin of tea is in southwest of China. Tea began as a medicine and grew to beverage. The world first commentary on tea was written by Lu Yu entitled *The Book of Tea* which was later exerting a profound influence after its appearance. Tea drinking spread widely and rapidly and its health benefits were documented. As listed in one of the greatest masterpieces of traditional Chinese medicine pharmacology Compendium of Materia Medica (Ben Cao Gang Mu), tea acts as a medicine as an antidote to herbal poisons (Zhen *et al.*, 2002).

Tea production including cultivation, harvesting and processing had developed rapidly since in Tang Dynasty (618 – 906 A.D). Tea is made by processing the tea leaves (*Camellia sinensis* O. kun-tze var. *assamica* Kitamura) (Jen *et al.*, 2008). Tea is classified into several types according to different processing method, typically green tea (nonferment tea), oolong tea (partially fermented tea), black tea (fully fermented tea) and Pu erh tea (produce by microbial fermentation of green tea) (Jen *et al.*, 2008).



Pu erh tea is one of China's most unusual tea with long history which is originated from the districts of Xi-shuang-ban-na, Si-mao, and Lan-chuang-jiang valley, Yunnan Province, China. Pu erh tea is named after an important town in southern Yunnan and also the largest market for tea trade located just north of Simao in the Xi-shuang-ban-na district of Yunnan. The ancient tea-horse road is a significant part of the international trade of tea in history, which starts from Sichuan and Yunnan Province in Southwest China, runs along the eastern foothills and deep canyons of several major rivers in between the six famous tea mountains.

## 2.2 Taxonomy

The taxonomic hierarchy of tea (*C. sinensis*) is classified as (ITIS, 2007):

Kingdom: Plantae  
Subkingdom: Tracheobionta  
Division: Magnoliophyta  
Class: Magnoliopsida  
Subclass: Dilleniidae  
Order: Theales  
Family: Theaceae  
Genus: *Camellia*  
Species: *Camellia sinensis*

## 2.3 Morphology

*C. sinensis* can be a shrub or evergreen tree up to 16 m tall. Leaves alternate, exstipulate, lanceolate to obovate, up to 30 cm long, 2–5 cm broad, pubescent, sometimes becoming glabrous, serrate, acute or acuminate; flowers 1–3, in axillary or subterminal cymes, deflexed, 2–5 cm broad, aromatic, white or pinkish, actinomorphic, sepals and petals 5–7, pedicels 5–15 mm long; stamens numerous; ovary 3–5 carpellate, each carpel 4–6 ovulate; capsules depressed-globose, brownish, lobate, to 2 cm broad, valvate, with 1–3 subglobose seeds in each lobe; approximately 500 seeds/kg (Duke & Atchley, 1984). Sabah tea leaves were shown in Photo 2.1.

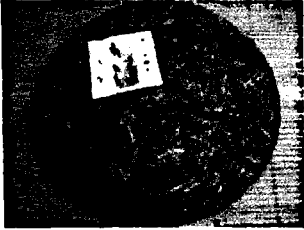




**Photo 2.1:** Fresh tea leaves (*C. sinensis*) in Sabah tea plantation.


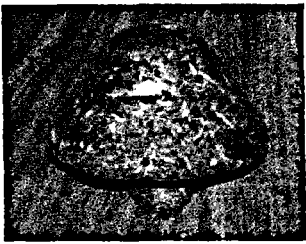

## 2.4 Classification of Pu erh tea

Pu erh tea can be classified in many ways by its shapes, processing methods, region, cultivation, grade, and season. Pu erh tea is commercially sold in three types of products: loose pu erh tea, pressed pu erh tea and pu erh tea bags (Jen *et al.*, 2008). However, Pu erh tea is compressed into a variety of shapes. The classification of Pu erh tea is shown in Table 2.1(Liu, 2005).

**Table 2.1 Classification of Pu erh tea**

Name	Description
 Cake (Bingcha)	A round, flat, disc shaped tea which its size ranges from 100 g to 5 kg. It is also known as seven units cake tea (qi zi bing cha) due to the fact that seven of the cake are packaged together at a time for sale or transport.
 Bowl (Tuocha)	A convex knob-shaped tea with size ranging from 3 g to 3 kg.
 Brick (Zhuancha)	A thick rectangular block of tea ranging from 100g to 1 kg.

**Table 2.1 continue**

 <p>Square (Fangcha)</p>	<p>A flat square of tea, usually in 100 g or 200g sizes which often contain words that are pressed into the square.</p>
 <p>Mushroom (Jincha)</p>	<p>A mushroom shape of pressed pu erh tea, usually in 250g or 300g.</p>
 <p>Gold melon (jingua)</p>	<p>A pumpkin-like shape tea.</p>

Pu erh tea can be classified into two main types according to its degree of fermentation. A green tea like pu erh tea which is commonly known as raw Pu erh is process without fermentation or storage under high humidity. The fermented Pu erh is produced by microbial fermentation of piled fresh loose tea and stored at room temperature which is known as mature Pu erh (Jen *et al.*, 2008).

There are three different tea cultivation methods which is important in classifying Pu erh tea. Some tea plants are planted in a plantation bushes (guanmu) which the seeds or cutting of wild tea are planted in relatively low altitudes and flatter terrain. Wild arbor trees are tea plants from older plantation that were cultivated in the previous generation that have gone feral due to lack of care. Wild tree (gushu) is tea from wild tree which grown without human intervention and are at the highest valued Pu erh teas. These teas have deeper and more complex flavors (Liu, 2005).

## 2.5 Chemical composition of Pu erh tea and its bioactivity

The composition of tea varies with species, season, and horticultural conditions and particularly with degree of fermentation during the manufacturing process (Chen *et al.*, 2009). Pu erh teas undergo a full fermentation stage by using microorganism like *Aspergillus niger*. During the fermentation, tea catechins in Pu erh are oxidized and condensed to other large polyphenolic molecules such as theaflavins and thearubigins. This oxidation gives a low content of catechin in Pu erh tea compare to green tea and semi-fermented oolong tea (Zuo *et al.*, 2002).

The microorganism in Pu erh tea oxidized tea polyphenols more completely than the enzymatic oxidation process which occurs in black tea, resulting in lower concentration of tea polyphenols and tea catechins (Xie *et al.*, 2009). During black tea processing, fresh tea leaves are rolled and cut before drying so that tea polyphenols in tea leaves come into contact with the tea polyphenol oxidizes and then oxidized in the consequences of fermentation process. However, during Pu erh tea fermentation process, fresh tea leaves are fixed by heat in a drum to inactivate polyphenol oxidizes. The fixed tea leaves is then rolled and partially dried and piled up in humid conditions for a week. The tea polyphenol is more intensively oxidized by the action of microorganism (Mo *et al.*, 2008).

### 2.5.1 Tea polyphenol

Tea polyphenols, previously called tea tannins, account for 30% of the dry weight of the fresh tea leaves. Catechins are predominant form of polyphenols which account for 12 – 24% of dry weight. Besides catechins, flavanol, and their glycosides, anthocyanidin and leucoanthocyanidin, phenolic acid and despidies also present. These phenolic compounds are directly or indirectly associated with the characteristics of tea, including its color, taste, and aroma. The polyphenol contents known to be present in Pu erh tea samples, such as gallic acid, (-) epicatechin (EC), (-) epigallocatechin (EGC), (-) epigallocatechin gallate (EGCG) and epicatechin gallate (ECG), catechin (C), myricetin, quercetin, and kaempferol. (Lu *et al.*, 2008).

#### 2.5.1(a) Tea flavonoids

Tea flavonoids are largely responsible for the distinctive taste and color of tea. The common flavonoids in tea are the flavan-3-ols (flavanols or flavans). The flavan-3-ols subclasses are ranked by degree of polymerization. The catechins are monomers which consists of (-) epicatechin (EC), (-) epigallocatechin (EGC), (-) epigallocatechin gallate (EGCG) and epicatechin gallate (ECG). However, the theaflavins are dimmers, such as theaflavin, theaflavin 3-gallate, theaflavin 3'-gallate, theaflavin 3,3'-digallate), and the derived tannins the arubigins are oligomers of unknown structure. Other flavonoids, including the flavonols (quercetin, kaempferol, myricetin) and flavones (apigenin and luteolin), are also present but lesser amount than flavan-3-ols (Peterson *et al.*, 2004). Flavonoids are colourless, water-soluble compound which impart bitterness and astringency to green tea infusion (Wang *et al.*, 2000). Total flavonoids content could be used to indicate the quality potential of tea, with high content being related to high quality (Magoma *et al.*, 2000). The characteristics of manufactures tea mostly are associated directly or indirectly with modifications to the flavonoids. During green tea manufacture, most catechins and other polyphenols are preserved owing to inactivation of the endogenous enzymes by dry heating or steaming at the initial step.

A vast body of scientific researches and studies has suggested that catechins are responsible for the majority of the potential health benefits attribute to tea consumption. Among the catechins, EGCG is most effective in reacting with most reactive oxygen species. This chemical structure contributes to effective antioxidant activity of catechins including the vicinal dihydroxyl or trihydroxyl structure which can chelate metal ions and prevent the generation of free radicals. This structure also allows electron delocalization, conferring high reactivity of quench free radicals. Free radicals occurring in the environment can trigger chain reaction which may cause oxidative damage to sensitive biological structures, such as DNA or cell membranes, and subsequently result in cancer, heart diseases. Studies in animal models suggest that EGCG may reduce some of the harmful effects following to exposure of UV radiation, and also inhibit cervical cancer cell growth (Nagle *et al.*, 2006). Epicatechin gallate (ECG) is reported to inhibit growth of some of the cancer cell lines (Vergote *et al.*, 2002) but not their normal counterparts.

Other catechins are also present in smaller amounts: gallocatechin, epigallocatechin digallate, 3-methylepicatechin gallate, catechin gallate, and gallocatechin gallate (Luczaj *et al.*, 2005). Figure 2.1 shows the chemical structure of major flavonoids in tea.



## REFERENCES

- Abe, M., Takaoka, N., Idemoto, Y., Takagi, C., Imai, T., & Nakasaki, K. 2008. Characteristics Fungi Observed in the Fermentation Process of Pu erh, *International Journal of Food Microbiology*, **124**: 199-203.
- Atoui, A. K., Mansouri, A., Boskou, G. & Kefalas, P. 2005. Tea and herbal infusions: Their antioxidant activity and phenolic profile. *Food Chemistry* **89**: 27-36.
- Ashihara, H., & Crozier, A. 2001. Caffeine: a well known but little mentioned compound in plant science. *TREND in Plant Science*, **6**: 407-413.
- Benzie, I.F.F., & Strain, J.J., 1996. The Ferric Reducing Ability of Plasma (FRAP) as a Measure of Antioxidant Power": The FRAP Assay. *Analytical Biochemistry*, **239**: 70-76.
- Chan, E.W.C., Lim, Y.Y., & Chew, Y.L. 2006. Antioxidant activity of *Camellia sinensis* leaves and tea from lowland plantation in Malaysia. *Food Chemistry* **102**: 1214-1222.
- Chen, C. N., Lin, P.C., Huang, K.K, Chen, W.C., Hsieh, H.P., Liang, P.H, Hsu, & T.A. 2005. Inhibition of SARS-CoV 3c-like Protease Activity by Theaflavin-3,3\_-digallate (TF3), *eCAM*, **2**: 209-215.
- Chen, Y.S., Liu, B.L., & Chang, Y.N. 2009. Bioactivities and sensory evaluation of Pu erh teas made from three tea leaves in an improved pile fermentation process. *Journal of Bioscience and Bioengineering*.
- Chiang, C. T., Weng, M. S., Lin-Shiau, S. Y., Kuo, K. L., Tsai, Y. J. & Lin, J. K. 2006. Pu erh tea supplementation suppresses fatty acid synthase expression in the rat liver through down regulating Akt and JNK signalings as demonstrated in human hepatoma HepG2 cells. *Oncology Research* , **16**: 119-128.
- Duh, P. D., Yen, G. C., Yen, W. J., Wang, B. S. & Chang, L. W. 2004. Effects of pu-erh tea on oxidative damage and nitric oxide scavenging. *Journal of Agricultural and Food Chemistry* , **52**: 8169-8176.
- Duke, J. A. & Atchley, A. A. 1984. Proximate analysis, in: B.R. Christie (Ed.), *The Handbook of Plant Science in Agriculture*, CRC Press.
- Fernandez, P. L., Pablos, F., Martin, M. J. & Gonzales, A. G. 2002. Study of catechin and xantine tea profiles as geographical tracers. *Journal of Agricultural and Food Chemistry* , **50**: 1833-1839.
- Graham, H. N. 1999. Tea. In J. F. Frederick (Ed). *Wiley encyclopedia of food science and technology*, 2<sup>nd</sup> ed. 2292-2305.

- Hou, Y., Shao, W.F., Xiao, R., Xu, K.L., Ma, Z.Z., Johnstone, B.H., & Du, Y.S., 2009. PU erh tea aqueous extract lower atherosclerotic risk factors in a rat hyperlipidemia model. *Experimental Gerontology*, **44**: 434-439.
- Huang, Y. C., Chang, Y. H., & Shao, Y. Y. 2006. Effects of Genotype and Treatment on the Antioxidant Activity of Sweet Potato in Taiwan. *Food Chemistry*, **98**: 529-538.
- ITIS. 2007. *Integrated Taxonomic Information System Report: Camellia sinensis (L.) O. Kuntze*. North America.
- Ito, E., Crozier, A., & Ashihara, H. 1997. Theophylline metabolism in higher plants. *Biochimica et Biophysica Acta*, **1336**:323-330
- Jen, K.L., & Shahidi, F. 2008. *Tea and Tea Product, Chemistry and Health-promoting Properties* CRC Press 11-13.
- Jeng, K.C., Chen, C.S., & Fang, Y.P. 2007. Effect of microbial fermentation on content of statin, GABA, and polyphenols in Pu-erh tea. *Journal of Agricultural and Food Chemistry*, **55**: 8787-8792.
- Jie, G. L., Lin, Z., Zhang, L. Z., Lu, H. P., He, P. M. & Zhao, B. L. 2006. Free radical scavenging effect of Pu-erh tea extracts and their protective effect on oxidative damage in human fibroblast cells. *Journal of Agricultural and Food Chemistry*, **54**: 8058-8064.
- Katalinic, V., Milos, M., Kulisic, T., & Jukic, M. 2006. Screening of 70 medicinal plant extracts for antioxidant capacity and total phenols. *Food Chemistry*, **94**(4): 550-557.
- Kuroda, Y., & Hara, Y. 1999. Antimutagenic and anticarcinogenic activity of tea polyphenols. *Mutation Research*, **436**: 69-97
- Lee, B.L., Ong, & C.N. 2000. Comparative analysis of tea catechins and theaflavins by high performance liquid chromatography and capillary electrophoresis. *Journal of Chromatography*. **1119**: 439-447.
- Lin, J. K., Lin, C. L., Liang, Y. C., Lin-Shiau, S.Y. & Juan, I. M. 1998. Survey of catechins, gallic acid, and methylxanthines in green, oolong, pu-erh and black teas. *Journal of Agricultural and Food Chemistry* **46**: 3635-3642.
- Liu, Q.J. 2005. *China Pu erh Tea*. Tea products. Guangdong Travel & Tourism Press.
- Luczaj, W. & Skrzydlewska, E. 2005. Antioxidative properties of black tea. *Preventive Medicine*, **40**: 910-918.
- Magoma, G. N., Wachira, F. N., Obanda, M., Imbuga, M., & Agong, S. 2000. The use of catechins as biochemical markers in diversity studies of tea (*Camellia sinensis*). *Genetic Resources and Crop Evolution*, **47**: 107-114.

- Mo, H., Zhu, Y. & Chen, Z. M. 2008. Microbial fermented tea – a potential source of natural food preservatives. *Trends in Food Science and Technology*, **19**: 124-130.
- Mo, H. Z., Xu, X. Q., Yan, M. C. & Zhu, Y. 2005. Microbiological analysis and antibacterial effects of the indigenous fermented Puer tea. *Agrology Food Industry Hi-Tech*, **16**: 16-18.
- Nagle, C.M., Bain, C.J., Webb, P.M. 2006. Cigarette smoking and survival after ovarian cancer diagnosis . *Cancer Epidemiol Biomarkers* ,**15**:2557-2560
- Ngure, F.M., Wanyoko, J.K., Mahungu,S.M., & Shitandi,A.A., 2009. Catechins depletion patterns in reation to theaflavin and thearubigins formation. *Food Chemistry*, **115**: 8-14.
- Parekh, J., Jadela, D. & Chanda, S. 2005. Efficacy of aqueous and methanol extracts of some medicinal plants for potential antibacterial activity. *Turkish Journal of Biology* **29**: 203-210
- Peterson, J., Dwyer, J., Bhagwat, S., Haytowitz, D., Holden, J., Eldridge, A. L., Beecher, G. & Aladesami, J. 2005. Major flavonoids in dry tea. *Journal of Food Composition and Analysis* , **18**: 487-501.
- Qian, Z.M., Guan, J.G., Yang, F.Q., & Li, S.P. 2008. Identification and Quantification of Free Radical Scavengers in Pu erh Tea by HPLC-DAD-MS Coupled Online with 3,3'-Azinobis (3-Ethylbenzthiazolinesulfonic Acid) Dlammonium Salt Assay. *Journal of Agricultural and Food Chemistry*, **56**:11187-11191.
- Sakanaka, S., Juneja, L. R., & Taniguchi, M. 2000. Antimicrobial Effects of Green Tea Polyphenols on Thermophilic Spore-Forming Bacteria. *Journal of Bioscience and Bioengineered*, **90**: 81-85
- Sano, M., Takahashi, Y., Yoshino, K., Nakamura, Y., Tomita, I., Oguni, I., & Konomoto, H. 1995. Effect of Tea (*Camellia sinensis* L.) on Lipid Peroxidation in Rat Liver and Kidney; a comparison of green tea and black tea feeding. *Biology Pharmaceutical Bull*, **18**: 1006-1008.
- Sharma, V., Gulati, A., Ravindranath, S.D., & Kumar, V. 2005. A simple and convenient method for analysis of tea biochemicals by reverse phase HPLC. *Journal of Food Composition and Analysis*, **18**: 583-594
- Syu, K.Y., Lin, C.L., Huang, H.C., & Lin, J.K. 2008. Determination of Theanine, GABA and Other Amino Acid in Green, Oolong, Black, and Pu erh tea with Dabsylation and Hign-Perforamnce Liquid Chromatography. *Journal of Agricultural and Food Chemistry*, **56**: 7637-7643.
- Vergote, D., Olive, C.C., Chopin, V., Toillon, R.A., Rolando, C., Hondermarck, H., & Bourhis. 2002. (-)-Epigallocatechin (EGC) of green tea induces apoptosis of

human breast cancer cells but not of their normal counterparts. *Breast Cancer Research and Treatment*, **76**: 195–201.

- Turkmen, N., Sari, F. & Velioglu, Y. S. 2006. Effects of extraction solvents on concentration and antioxidant activity of black and black mate tea polyphenols determined by ferrous tartrate and Folin-Ciocalteu methods. *Food Chemistry*, **99**: 835-841.
- Wang, B. S., Yu, H. M., Chang, L. W., Yen, W. J., & Duh, P. D. 2008. Protective effects of pu-erh tea on LDL oxidation and nitric oxide generation in macrophage cells. *Lebensmittel-Wissenschaft und-Technologie*, **41**: 1122-1132.
- Wang, H. F., Helliwell, K. & You, X.Q. 2000. Isocratic elution system for the determination of catechins, caffeine and gallic acid in green tea using HPLC. *Food Chemistry*, **68**: 115–121.
- Wu, S.C., Yen, G.C., Wang, B.S., Chiu, C.K., Yen, W.J., Chang, L.W., Duh & P.D. 2007. Antimutagenic and antimicrobial activities of pu-erh tea. *Lebensmittel-Wissenschaft und-Technologie*, **40**:506-512
- Xie, G.X., Ye, M., Wang, Y.G., Ni, Y., Su, M.M., Huang, H., Qiu, M.F., Zheng, X.J., Chen, T.L, Jia, W. 2009. Characterization of Pu erh Tea Using Chemical and Metabolism Profiling Approaches. *Journal of Agricultural and Food Chemistry*, **57**: 3046-3054.
- Yang, D.J., Hwang, & L.S., 2006. Study on the conversion of three natural statins from lactone forms to their corresponding hydroxy acid forms and their determination in Pu-erh tea. *Journal of Chromatography*, **1119**: 277–284.
- Yang, X.R., Ye, C.X., Xu, J.K., & Jiang, Y.M. 2007. Simultaneous analysis of purine alkaloids and catechins in *Camellia sinensis*, *Camellia ptilophylla* and *Camellia assamica* var, *kucha* by HPLC. *Food Chemistry*, **100**: 1132-1136.
- Zhen, Y.S., Chen, Z.M., Cheng, S. J., Chen, M. L. 2002. *Tea: Bioactivity and Therapeutic Potential*. CRC Press, 267.
- Zuo, Y., Chen, H., Deng, Y. 2002. Simultaneous Determination of Catechins, Caffeine and Gallic Acids in Green, Oolong, Black and Pu-erh Teas using HPLC with a Photodiode Array Detector. *Talanta*, **57**: 307-316.