EFFECTIVENESS OF VINEGAR AND SUCROSE IN ROOM TEMPERATURE ON THE VASE LIFE OF POSTHARVEST ROSES

SABITRA A/P RATHRVELOO

DISSERTATION SUBMITTED IN PARTIAL FULFILLMENT OF THE REQUIREMENTS FOR THE DEGREE OF BACHELOR OF AGRICULTURE SCIENCE WITH HONOURS

HORTICULTURE AND LANDSCAPING PROGRAMME SCHOOL OF SUSTAINABLE AGRICULTURE UNIVERSITI MALAYSIA SABAH 2011



UNIVERSITI MALAYSIA SABAH

_	
ſ	BORANG PENGESAHAN STATUS TESIS
ľ	JUDUL: EFFECTIVENESS OF VINEGAR AND SUCROSE IN ROOM
ł	TEMPERATURE ON THE VASE LIFE OF POSTHARVEST
1	ROSES
	IJAZAH: BACHELOR OF AGRICULTURE SCIENCE WITH HONDURS (HG 35)
	SAYA: <u>SABITRA RATHRUELOO</u> SESI PENGAJIAN: 2-2010/2011 (HURUF BESAR)
	Mengaku membenarkan tesis * (LPSM/ Sarjana/Doktor Falsafah) ini disimpan di Perpustakaan Universiti Malaysia Sabah dengan syarat-syarat kegunaan seperti berikut:-
	 Tesis adalah hakmilik Universiti Malaysia Sabah. Perpustakaan Universiti Malaysia Sabah dibenarkan membuat salinan untuk tujuan pengajian sahaja. Perpustakaan dibenarkan membuat salinan tesis ini sebagai bahan pertukaran antara institusi pengajian tinggi. Sıla tandakan (/)
	SULIT (Mengandungi maklumta yang berdarjah keselamatan atau kepentingan Malaysia seperti yang termaktub di AKTA RAHSIA RASMI 1972)
	TERHAD (Mengandungi maklumat TERHAD yang telah ditentukan oleh organisasi/badan di mana Penyelidikan dijalankan)
	TIDAK TERHAD
	PERPUSTAKAAN SERAU INIVERSITI MALAYSIA SABAD UNIVERSITI MALAYSI
	b sobetro. Ann
	(TANDATANGAN PENULIS)
	Alamat Tetap: J 8190 TAMAN KESANG INDAH, JJDCO Setolah Pertanjan Lestari
	JASIN, MELAKA. Universiti Malaysia Sabah
	(NAMA PENYELIA dan cop)
	Tarikh: 9/05/11 Tarikh: 9/5/11
	 Potong yang tidak berkenaan. ** Jika tesis ini SULIT atau TERHAD, sila lampirkan surat daripada pihak yang berkuasa/organisasi berkenaan dengan menyatakan sebab dan tempoh tesis ini perlu dikelaskan sebagai SULIT atau TERHAD. Tesis dimaksudkan sebagai tesis bagi Ijazah Doktor Falsafah dan Sarjana Secara penyelidikan atau disertasi bagi pengajian secara kerja kursus dan Laporan Projek Sarjana Muda (LPSM)
L	



DECLARATION

I hereby declare that this dissertation is based on my original work except for citations and quotations which have been duly acknowledged. I also declare that no part of this dissertation has been previously or concurrently submitted for a degree at this or any other university.

SABITRA A/P RATHRVELOO BR07110048 20 APRIL 2011



En Lum Mok Sam Ketua Program Program Hortikultur dan Landskap Sekolah Pertanian Lestari Universiti Malaysia Sabah Jalan UMS 88400 Kota Kinabalu Sabah

Tuan

PENYERAHAN KUASA TANDATANGAN DALAM BUKU DISSERTASI BAGI PELAJAR SABITRA A/P RATHRVELOO (BR07110048)

Dengan segala hormatnya perkara di atas adalah dirujuk.

Untuk makluman tuan, saya adalah pemeriksa dissertasi bagi pelajar Sabitra A/P Rathrveloo (BR07110048) pada Semester 2 Sesi 2010/2011. Saya telah menyemak dan berpuashati dengan pembetulan yang telah dilaksana oleh pelajar tersebut. Walau bagaimanapun, atas sebab-sebab yang tidak dapat dielakkan, saya tidak dapat menurunkan tandatangan saya dalam dissertasi pelajar tersebut.

Saya dengan ini menyerahkan kuasa tandatangan saya kepada Dr. Mohamadu Boyie Jalloh yang juga selaku Ketua Program Pengeluaran Tanaman, untuk menandatangani dissertasi pelajar tersebut.

Sekian, harap maklum.

Yang benar

Mohammad Amizi Bin Ayob

BOHANAD AMIZI 1914 AY Senior Lecturer / Academic Adyle School Sustainable Agricultura Universiti Malausia Sebeh

Sk: Dr. Suzan Benedick, Timbalan Dekan (A&HEP) Dr. Mohamadu Boyie Jalloh Dr. Abdul Rahim Awang dan Cik Chee Fong Tyng, Penyelaras Porjek Tahun Akhir



ACKNOWLEDGEMENT

An endeavor over long period can be successful only with advice and guidance of many well wishers. I am highly indebted to my supervisor Pn. Rosmah Murdad, for her assistance and constant source of encouragement. Furthermore I would thank my supervisor for sparing time to go through every tiny detail and give her valuable suggestions to make this project and report a success and also for sparing her valuable time to extend help in every steps of my project work. I would also like to thank the staff of School of Sustainable Agriculture for their generous guidance. I would also like to thank to those who were involved directly or indirectly in helping me completing this project and may your charity and goodwill will be blessed. Last but not the least I would like to thank my friends and family for their help in every way for the success of this project report.



ABSTRACT

The objective of this study was to evaluate the effect of vinegar and sucrose as a potential floral preservative that can extend the vase life of postharvest roses at room temperature $(25\pm5^{\circ}C)$. The tested treatments were distilled water (control), 0.5% of vinegar, 1.0% of vinegar, 2% of sucrose, 4% of sucrose, 2% of sucrose and 0.5% of vinegar, 4% of sucrose and 0.5% of vinegar, 2% of sucrose and 1.0% of vinegar, and 4% of sucrose and 1.0% of vinegar. The treatments were arranged according to Complete Randomized Design (CRD). Five replicates were prepared for each treatment and replicates were stored in room temperature ($25\pm5^{\circ}C$). The water loss and vase life data of the cut roses were observed and recorded daily. The bacteria was cultured on the 7th day of experiment. Day 7 was selected as a control day. The data on water loss was compared across the treatments using SPSS software by performing One-way ANOVA. The results suggested that vinegar and sucrose significantly affected the vase life of cut roses (p<0.05). Treatments with 2% of sucrose and 0.5% of vinegar was the best treatment to extent the vase life of the cut roses at room temperature ($25\pm5^{\circ}C$) to 20 days without existence of bacteria .



KEBERKESANAN CUKA DAN SUKROSA DALAM SUHU BILIK TERHADAP JANGKA HAYAT BUNGA KERATAN ROS

ABSTRAK

Kajian ini dijalankan untuk mengetahui kesan keberkesanan cuka dan sukrosa sebagai pengawet bunga yang bagus dalam suhu bilik terhadap jangka hayat bunga keratan ros. Rawatan-rawatan yang dikaji adalah air suling, 0.5% cuka, 1.0% cuka, 2% sukrosa, 4% sukrosa, 2% sukrosa dengan 0.5% cuka, 4% sukrosa dengan 0.5% cuka, 2% sukrosa dengan 1.0% cuka, dan 4% of sukrosa dengan 1.0% cuka. Semua rawatan disusun mengikut Rekabentuk Rawak Lengkap (CRD). Setiap rawatan direplikasi sebanyak lima kali dan disimpan di suhu bilik (25±5 °C). Jangka hayat dan kuantiti air yang hilang diperhatikan setiap hari dan data tersebut dicatatkan. Pengkulturan bakteria dilakukan pada hari ketujuh kajian. Hari ketujuh dipilih sebagai hari kawalan. Data kehilangan kuantiti air dianalisis dengan menggunakan perisian SPSS (analisis ANOVA Satu-hala). Keputusan yang diperolehi mencadangkan keratan bunga ros yang dirawat dengan 2% sukrosa + 0.5% cuka, merupakan rawatan yang terbaik dimana keratan bunga ros mencapai jangka hayat selama 20 hari pada suhu bilik (25±5 °C), (p<0.05) tanpa kehadiran bakteria.



TABLE OF CONTENTS

Content	Page
DECLARATION	ii
VERIFICATION	iii
AKNOWLEDGEMENT	iv
ABSTRACT	V
ABSTRAK	vi
TABLE OF CONTENTS	vii
LIST OF TABLES	ix
LIST OF FIGURES	х
LIST OF SYMBOLS, UNITS, AND ABBREVATION	xii
LIST OF FORMULAE	xiii

CHAP	TER 1	INTRODUCTION	1
1.1	Introduction		1
1.2	Problem State	ement	4
1.3	Justification		4
1.4	Objectives		4
1.5	Hypotheses		5

СНАРТ	ER 2	LITERATURE REVIEW	6
2.1	Taxono	my of Rose Flower	6
2.2	History	of Rose Flower	7
	2.2.1	Roses in the Ancient World	8
2.3	Posthar	vest Consideration for Roses	9
	2.3.1	Quality of Cut Flower	9
	2.3.2	Light	10
	2.3.3	Temperature	10
	2.3.4	Water Stress	11
2.4	Chemic	al Applications (Floral Preservatives)	13
	2.4.1	Silver Thiosulfate (STS)	13
	2.4.2	Sucrose	13
2.5	Vinega	r	15
2.6	Vase L	ife	16
2.7	Visual	Appearance	17
2.8	Bacteri	a in Vase Solution	17
2.9	Seneso	cence and Abscission of Cut Flower	18
	a)	Leaf Senescence	18
	b)	Petal Senescence	19



CHAP	FER 3	MATERIALS AND METHODS	20
3.1	Experir	nent Location	20
3.2	Materia	als	20
	3.2.1	Plant Material	20
	3.2.2	Chemicals	20
3.3	Flower	Preparation	21
3.4	Param	eters	22
	3.4.1	Determination of Vase Life	22
	3.4.2	Determination of Water Loss	23
	3.4.3	Determination of Number of Bacteria in Vase Solution	24
3.5	Treatn	nents	25
3.6	Experi	mental Design	26
3.7	Data A	Analysis	27
CHAD	TED 4	RESULTS	28
	Effort	of Vinegar and Sucrose on Water loss of Cut Rose Flowers	20
4.2	Effort	of Vinegar and Sucrose on Vace Life of Cut Rose Flowers	20

4.2 Effect of Vinegar and Sucrose on Vase Life of Cut Rose Flowers
4.3 Effect of Vinegar and Sucrose on the Number of bacteria in vase
35 solution

СНАРТ	ER 5 DISCUSSION	37
5.1	ntroduction	37
5.2	ffect of Vinegar and Sucrose on Water Loss of Cut Rose Flowers	37
5.3	ffect of Vinegar and Sucrose on Vase Life of Cut Rose Flowers	39
5.4	Effect of Vinegar and Sucrose on the Number of bacteria in vase solution	41
CHAP	ER 6 CONCLUSION	44

REFERENCES	45
APPENDIX	52



LIST OF TABLE

Table		Page
3.1	Criteria used to determine the end of rose cut flower vase life	22
3.2	Concentrations of vinegar and sucrose used for vase solution at	25
	room temperature (25 \pm 5 ^o C)	
5.1	Amount of vinegar for standard preservative solution	40



LIST OF FIGURES

Figure	e	Page
2.1	Fresh cut rose flowers	7
2.2	Vinegar	15
3.1	Design layout of cut roses arrangement at room temperature ($25\pm5^{\circ}C$)	26
4.1	Mean water loss (%) of cut rose flowers of at room temperature ($25\pm5^{\circ}$ C)	29
4.2	The effect of vase solution on overall water loss percentages of cut rose	31
	flowers at room temperature (25±5 °C)	
4.3	Mean vase life of cut rose flowers of at room temperature (25±5 $^{\circ}$ C)	33
4.4	Effect of vinegar and sucrose on the number of bacteria in vase	35
	solution at room temperature (25±5 °C)	
4.5	Presence of bacteria on nutrient agar (A) T1-Distilled water; (B) T4- 2%	36
	of sucrose; (C) T5-4% of sucrose	



LIST OF SYMBOL, UNITS AND ABBREVIATIONS

ANOVA	Analysis of Variance
BA	benzyladenine
Chi	Chlorophyll
ClO ₂	Chlorine dioxide
DDMH	dimethyl-thydanton
HQC	hydroxyquinoline citrare
PCD	Programmed Cell Death
PGR	Plant Growth Regulator
RH	Relative Humidity
RM	Ringgit Malaysia
SSA	School of Sustainable Agriculture
STS	Silver thiosulfate
UMS	Universiti Malaysia Sabah
SPSS	Statistical Package for Social Science



LIST OF FORMULAE

Formulae

Percentage of water loss = (R1-R2) X 100% 3.1

Whereas;

R1=Weight of vase solution with cut flower (Day 1)

R1

R2=Weight of vase solution with cut flower (Day 2)

3.2 Number of bacteria

> Number of CFU Number of CFU 24 Volume plated (mL) x total dilution used mL

*CFU= Colony-forming unit, volume plated= the volume of inoculum used to spread in agar plate, total dilution used= dilution factor (e.g 10¹)



Page

23

CHAPTER 1

INTRODUCTION

1.1 Introduction

Rose is a perennial flower shrub or vine of the genus *Rosa*, within the family Rosaceae, that contains over 100 species and comes in a variety of colours. The species form a group of erect shrubs, and climbing or trailing plants, with stems that are often armed with sharp prickles. Most are native to Asia, with smaller numbers of species native to Europe, North America, and northwest Africa. Natives, cultivars and hybrids are all widely grown for their beauty and fragrance. There are many types of roses. They are, wild roses, old garden roses like, Alba, Gallica, Damask, Moss, Potland, Hybrid Musk, and modern garden roses like, Hybrid tea, Pernetiana, Polyantha, and landscape roses

Rose, a universally celebrated flower, has been used as a garden plant since the dawn of civilization. Rose is a symbol of perfection, elegance, romance and love. It was called "The Queen of Flowers" firstly by Greek poetess in her "Ode to the Rose" (Muhammad *et al.*, 1996). Roses (*Rosa hybirida*) belong to family Rosaceae and Genus *Rosa* which contains more than 150 species and 1400 cultivars (Synge, 1971). Rose enjoys superiority over all other flowers being extensively used for decorative purposes and is prized for its delicate nature, beauty, charm and aroma. Rose is recognized for their high economic value, which are used in agro-based industry especially in



LINIVERSITI MALAYSIA SABAH

cosmetics and perfumes. Additionally, roses play a vital role in the manufacturing of various products of medicinal and nutritional importance. However, the main idea of rose plant cultivation is to get the cut flowers, which greatly deals with the floricultural business (Butt, 2003).

Cut flowers are parts of plants, characteristically including the blooms or "inflorescences" and some attached plant materials, but not including roots and soil. Fresh cut flowers are highly perishable because they maintain only limited lifesupporting processes by taking water up through their stems. The United States is the third-largest producer of cut flowers in the world; production was valued at \$424 million in 2001, down from \$472 million in 1997. Almost all domestic production serves the U.S. market, which has seen increasing demand for cut flowers since the late 1980s. The United States has one of the most diverse cut flower markets, with all types of fresh flowers arriving from all over the world. Roses are the leading fresh cut flower produced and consumed in the United States (based on value) and, in 2001, accounted for almost 30 percent of fresh cut flower shipments. Fresh cut flowers are used for decorative purposes such as vase arrangements and bouquets at formal events; designs for weddings; gifts on occasions such as Mother's Day, Valentine's Day, in times of illness, and at holidays such as Christmas and Easter; corsages and boutonnieres; and informal displays to beautify homes and public places (Joanna Bonarriva, 2003).

The cut flower industry in Malaysia is a relatively recent development compared to other agricultural enterprises. From its rather humble origins as a hobby industry, the cut flower industry in Malaysia has developed into a very viable commercial enterprise with the most marked growth in the mid-Eighties. In fact it has shown such tremendous growth in the last decade that production has increased tenfold and export twelve-fold in response to local and foreign demands. The trend is expected to continue in the future with growing affluence of the local population and that of the developed countries as well as improved market opportunities. The cut flower market consists of three important components which are temperate flowers, orchids and other lowland flowers. In general, the area of cultivation of cut flowers in Malaysia is determined by the climate and topography of the land. For instance, highlands such as the Cameron Highlands are the major growing areas of temperate flowers. Other cut



flowers adapt better to the hot humid conditions in the lowlands with orchids constituting the major share of the production (Jong *et al.*, 2000).

In the National Agriculture Policy (1992-2010) and the Seventh Malaysia Plan (1996-2000), cut flowers have been identified as a priority group of crops with good potential to meet the growing domestic and international demand and to generate higher income for producers. It has been estimated that at least 1,000 farmers are involved in the floriculture industry that contributed RM 370 million to the economy in 1995. Orchids contributed 40 percent of total value of production, followed by temperate cut flowers (33 percent) and ornamental plants (27 percent) (Jong *et al.*, 2000).

Although the contribution of cut flowers to the total of Malaysian exports is not significant, the annual growth rate over the last year was 24%. If the forecast of 10% annual growth rate of floriculture production holds true for the 7th Malaysia Plan period (1996-2000), the floriculture industry will enjoy the highest annual growth rate compared to other major commodities including rubber and palm oil over the same period (Mohd. Ridzuan *et al.*, 2000).

It is clearly stated that, nowadays cut flowers especially cut roses are very demanding. Among horticultural crops, cut flowers and other ornamentals have perhaps the highest value and are the most perishable. Their high respiration rates, rapid deterioration, and susceptibility to damage require the utmost care to maintain quality during postharvest handling. In postharvest handling, the most important thing that we must consider is, the preservative solution or floral preservative that we placed the cut flowers. The use of preservative solution is considered a common practice for the storage of floral stems. These treatments allow to control ethylene synthesis, pathogen development, maintenance of hydric and respiration balance, to contribute to colour conservation, floral buttons induction and latter to complete their development (Halevy and Mayak, 1981). For these reasons, many floral preservative contain germicides, ethylene synthesis inhibitors, growth regulators, some mineral compounds, and carbohydrates that are essential to extend the vase life of cut flowers (Halevy and Mayak, 1981). In my research I used commercial vinegar and sucrose as preservative solution to extend the vase life of cut roses.



3

1.2 Problem Statements

There are many factors involved in keeping good quality of cut flowers, but the most important factors are appropriate storage temperature, flower preservatives, ethylene inhibitor, and inhibitor of microorganisms (Scott *et al.*, 1994). Floral preservatives are very important to increase the vase life and maintain good quality of cut rose flowers (Roberts *et al.*, 1993). Growers and florists do not give much attention on this matter because floral preservatives are expensive and not easily available in the market. To overcome this problem, they (especially florists) need flower preservatives that are easily available and cheaper.

1.3 Justification

This study has been designed to evaluate the effectiveness of vinegar and sucrose in room temperature on the vase life of postharvest roses which has potential to be good flower preservative. This is one of the initiative to replace the expensive flower preservative in the market where vinegar and sucrose can be flower preservative which is economical and easily available. The importance of this study is to suggest a good flower preservative (vinegar + sucrose) where florists and growers can prepare by their own with simple steps without spending much money.

1.4 Objective

The objective of this study is:

I. To evaluate the effect of vinegar and sucrose as a potential flower preservative that can extend the vase life of postharvest roses at room temperature.



1.5 Hypotheses

- H_0 = There are no significant differences in the effect of vinegar and sucrose on postharvest roses in room temperature.
- H₁ = There are significant differences in the effect of vinegar and sucrose on
 postharvest roses in room temperature.





CHAPTER 2

LITERATURE REVIEW

2.1 Taxonomy of Rose Flower

Roses for the longest time have enjoyed the honour of being the most popular flowers in the world. The reason for popularity of the rose flower may be its wide variety in terms of colour, size, fragrance and other attributes. There are many types of rose flowers. They are, Alba, Gallica, Damask, hybrid tea roses, hybrid musk, floribunda and more. Flowers may be solitary or panicled. Many native species have five petals and numerous stamens. The ovary is inferior and develops into fleshy fruit or hip, which can become colourful yellow to red when ripe (Dole and Wilkins, 2005). Figure 2.1 shows picture of a fresh cut rose flower. Below are the scientific classification of rose:

Kingdom	: Plantae
Division	: Magnoliophyta
Class	: Magnoliopsida
Order	: Rosales
Family	: Rosaceae
Subfamily	: Rosoideae
Genus	: Rosa

(Source: http://en.wikipedia.org/wiki/Rose)





Figure 2.1	Fresh cut rose flower
Source:	http://www.google.com.my/imglanding/rose
Bar = 1 cm	

2.2 History of Rose Flower

The rose apparently originated in Central Asia about 60 to 70 million years ago, during the Eocene epoch, and spread over the entire Northern Hemisphere. Early civilizations, including the Chinese, the Egyptians, the Phoenicians, the Greeks, and the Romans, appreciated roses and grew them widely as long as five thousand years ago.

About 500 B.C. Confucius wrote of roses growing in the Imperial Gardens and noted that the library of the Chinese emperor contained hundreds of books about roses. It is said that the rose gardeners of the Han dynasty (207 B.C.-A.D. 220) were so obsessed with these flowers that their parks threatened to engulf land needed for producing food, and that the emperor ordered some rose gardens plowed under.



The oldest rose identified today is *Rosa gallica*, also known as the French rose, which once bloomed wild throughout central and southern Europe and western Asia, and still survives there. Although the exact origin of *Rosa gallica* is unknown, traces of it appear as early as the twelfth century B.C., when the Persians considered it a symbol of love (Bose and Yadav, 1989).

2.2.1 Roses in the Ancient World

Descending from *Rosa gallica* is *Rosa damascena*, the damask rose, whose well-known fragrance has been part of rose history since the rose first appeared in about 900 B.C. About 50 B.C. a North African variant called *Rosa damascena semperflorens*, the 'Autumn Damask', thrilled the Romans because it bloomed twice a year - a trait previously unknown to them. The 'Autumn Damask', which has been traced back to at least the fifth century B.C., is believed to be a cross between *Rosa gallica* and *Rosa moschata*, the musk rose. Until European merchants discovered the tea and China roses in the Orient many centuries later, this rose would be the only repeat bloomer known to the Western world.

Another important early rose was *Rosa alba*, the 'White Rose of York'. Made famous as the emblem of the House of York during the fifteenth-century Wars of the Roses, this five-petal rose is actually far older, dating to before the second century A.D. It probably originated in the Caucasus and travelled west by way of Greece and Rome. *Rosa alba* and its relatives, known as albas, are believed to have descended from some combination of *Rosa gallica*, *Rosa damascena*, *Rosa canina*, and *Rosa corymbifera*.

The early Phoenicians, Greeks, and Romans all grew and traded in roses, which they brought with them as they travelled and conquered. As a result, roses spread throughout the Middle East and elsewhere in the Mediterranean.

The Greek scientist and writer Theophrastus, cataloging roses known about 300 B.C., described their flowers as having anywhere from five to one hundred petals. He was the first known detailed botanical description of a rose. Alexander the Great, king of Macedonia around this time, grew roses in his garden and is credited for introducing



8

cultivated roses into Europe. He may have had something to do with rose growing in Egypt, too.

In ancient Rome patricians tended rose gardens at their homes, and public rose gardens were a favourite place to pass a summer afternoon. Records show that there were two thousand public gardens in Rome before its fall in A.D. 476. The poet and satirist Horace complained about the short sightedness of the Roman government for allowing rose gardens to be planted where the land should have been used for wheat fields and orchards (http://mirrorofisis.freeyellow.com).

2.3 Postharvest Consideration for Roses

Problem of the cut rose flower begin at harvest; if cut too early, the rose may contain little store food. Generally rose should be cut at the stage which the flowers will continue to develop for the final consumer. There are two short-term problems of the cut rose; water loss increases at harvest and water moves into the leaves from the stems where air enters the stem base. The first problem causes wilting while the second one limits the rate of water uptake when the flowers are placed in the water. Hydration of cut flowers in deep ,which is the harvest water problem can be solved by adding acidifier to the water (Durkin, 1980).

2.3.1 Quality of Cut Flower

Fresh flowers and ornamental plants differ from edible and other agricultural products because edible and other agricultural products will be consumed or processed within few days after harvest. But, fresh flowers are expected to maintain their aesthetic value as long as possible. Quality in horticultural products is assessed by the relative value of several characteristics which considered together, will determine the acceptability of the product to the buyer and the customers (Nelson, 2003).



2.3.2 Light

Light can influence postharvest performance of roses. Cut roses exposed to constant light or 12 hours of light per day lost 5 times more water than roses kept in darkness (Halevy and Mayak, 1981). This, it is thought, was due to stomatal opening caused by the light. Light intensity directly influences the efficiency of photosynthesis, which determines the carbohydrate contents of the cut flowers. Petal colour is also affected by shading or low light intensity, and thus depends on the availability of carbohydrates in surrounding tissues, resulting in tissue blueing in the presence of low light intensity greatly reduces plant and consequently flower quality due to leaf and flower abscission (Cushman *et al.,* 1994).

2.3.3 Temperature

Temperature is the most important factor that cause damages to cut flowers. Cut flowers have shown that flowers are often exposed to damaging in high temperatures (Maxie *et al.*, 1974). The suitable temperature for most perishable horticultural commodities to last longer is near at 0 °C, where low temperatures reduce both metabolic processes and microbial growth rate and germination (Van Doorn and de Witte, 1991). Poor temperature management during transport of cut flowers is largely the result of inadequate pre-cooling and transport under non-refrigerated conditions Several researchers have shown the negative effects of improper storage temperatures⁴ on vase life of a range of cut flowers (Cevallos and Reid, 2001).

Since temperature is the major factor affecting the cut flower life, emphasis is laid on cool temperature. Cooling is essential in order to reduce the metabolic changes such as enzyme activity, and to slow down the maturation of flower or foliage. The cooling period required for flowers is generally depends upon difference between the temperature of flowers and the room temperature, quantity of flowers to be cooled, species of flowers and efficiency of the refrigeration (Teixera da silva, 2003).



10

The optimum temperature depends on the species, cultivar, and the stage of maturity, production environment and duration of storage (Wills, 2007). Temperature, relative humidity, air composition and air circulation in storage environment influence the postharvest lasting quality of cut flower (Dole and Wilkin, 2005).

2.3.4 Water Stress

Water stress, has a detrimental effect on cut flowers, causing wilting that ends their vase life. Three factors are involved in maintaining a balance of water in the cut flower. They are uptake and transport of water, loss of water through stomates or transpiration, and retention of water by plant tissues. Uptake and transport of water can be inhibited by several factors. Microbial growth, and some metabolites produced by these bacteria, can block the xylem in the lower part of the stem. The bacteria suspected to be the biggest problems are *Pseudomonas sp.* and members of the enterobacteriaceae found in tap water (van Doorn and de Witte, 1997).

Loss of water, through stomates and transpiration, will cause wilting if this loss exceeds water uptake. Cut roses kept in the light lost five times more water than those kept in the dark because the stomates were open in the light (Halevy and Mayak, 1981). The ability of plant tissues to hold water is affected by osmotic potential and by membrane integrity. Sugar added to vase water of cut flowers prolongs flower life by helping to maintain the osmotic potential in the plant tissue and by adding metabolic fuel.

Water balance is a major factor determining quality and longevity of cut flowers. It is influenced by water uptake and transpiration, being the balance between these two processes (Teixera da silva, 2003). When the amount of transpiration exceeds the volume of water uptake, water deficit and wilting develop (Halevy and Mayak, 1981). Low water uptake is often due to occlusions located mainly in the basal stem end (He *et al.*, 2006), and microbes are a common cause of stem end blockage (van Doorn, 1997). Many agents have been used in cut flower vase solutions to extend vase life by improving water uptake. These include silver nitrate (Fujino *et al.*, 1983), aluminum sulphate (Ichimura and Shimizu-Yumoto, 2007) and 8-hydroxyquinoline



11

REFERENCES

- Acock, B. and Nichols, R. 1979. Effect of sucrose on water relation of cut, senescing carnation Flowers. *Annals of Botany* **44**: 221-230.
- Acquaah, G. 2002. *Horticulture; Principle and Practices*. Edition 2. United States of America: Prentice Hall.
- Bleeksma, H.C. and van Doorn, W.G. 2003. Embolism in rose stems as a result of vascular bud break, yield, and vase life of cut roses. *Horticultural Science* 28: 621-622.
- Bose, T. K. and Yadav, L.P 1989. *Commercial Flower*. First edition. India: Department of Horticulture.
- Burdett, A.N. 1970. The cause of bent neck in cut roses. *Journal of the American* Society for Horticultural Science **95**: 427–431.
- Butt, S.J. 2003. A Review on prolonging the vase life of Roses. *Pakistan Rose Annual.* Published by Pakistan National Rose Society, 49-53.
- Butt, S.J. 2005. Extending the vase life of roses (*Rosa hybirida*) with different preservatives. *Anatomy of Agriculture Biology* **7**: 1, 91-99.
- Casadoro, G., Trainotti, L. and Tomasin, C. A. 1999. Expression of abscission-related endo-β-1, 4 glucanases. In: Kanellis, A.K., Chang, C., Klee, H., Bleecker, A.B., Pech, J.C., Grierson, D. (Eds.), *Biology and biotechnology of the plant hormone ethylene II.* Kluwer Academic Publishers, Dordreecht 243-247
- Cevallos, J.C. and Reid, M.S 2001. Effectof storage temperature on respiration and vase life of selected flowers. *Postharvest Biology and Technology*.
- Cushman, L.C., Pemberton, H.B., Miller, J.C. and Kelly J.W. 1994. Interactions of Flower Stage, Cultivar, and Shipping Temperature and Duration Affect Pot Rose Performance. *Horticultural Science* **33**: 736-740
- De Stigter, H.C.M. 1980. Water balance of cut and intact Sonia rose plants. *Plant Physiology* **99:** 131–140.
- De Witte, Y. and Van Doorn, W.G. 1988. Identification of bacteria in the vase water of roses, and the effect of the isolated strains on water uptake. *Scientia Horticulturae* **35**: 285-291.
- Dixon, M.A. and Peterson, C.A. 1989. A re-examination of stem blockage in cut roses. Scientia Horticulturae **38**: 277–288



- Dole, J.M. and Wilkins, H.F. 2005. *Floriculture: Principles and Species*. Second edition. United States of America : Prentice Hall.
- Durkin, D. J. 1980. Factors affecting hydration of cut flowers. *Acta Horticulturae* **113**: 109-117.
- Durkin, D.J. 1980. *A Manual of Greenhouse Rose Production*. Michigan: Roses incoprated.
- Durkin, D.J. 1979. Effect of Millipore filtration, citric acid, and sucrose on peduncle water potential of cut rose flower. *Journal of the American Society for Horticultural Science* **104**: 860–863.
- Eason, J.R., de Vre, L.A., Somerfield, S.D. and Heyes, J.A. 1997. Physiological changes associated with *Sandersonia aurantiaca* flower senescence in response to sugar. *Postharvest Biology and Technology* **12**: 43–50.

Estelle, M. 2001. Proteases and cellular regulation in plants. Plant Biology 4: 254-260

- Evans, R., Zheng, J.M. and Reid, M.S. 1996. Structural and environmental factors affecting the postharvest life of cut roses. *Acta Horticulturae*. **424**: 169–173.
- Florack, D.E.A., Willem J. S. and Bosch, D. 1996. Toxicity of Peptides to Bacteria Present in the Vase Water of Cut Roses. *Postharvest Biology and Technology* 8: 285-291
- Fujino, D.W., Reid, M.S. and Kohl, H.C. 1983. The water relations of maidenhair fronds treated with silver nitrate. *Scientia Horticulturae* **19**: 349–355.
- Gilman, K.F. and Steponkus, P.L. 1972. Vascular blockage in cut roses. *Journal of the American Society for Horticultural Science* **97:** 662–667.
- Goszczynska, D.M. and Rudnicki, R.M. 1988. Storage of cut flowers. *Horticultural Reviews* **10**: 35-62.
- Halevy, A.H., Byrne, T.G., Kofranek, A.M., Farnham, D.S., Thompson, J.F.and Hardenburg, R.E., 1978. Evaluation of postharvest handling methods for transcontinental truck shipments of cut carnations, chrysanthemums and roses. *Journal of the American Society for Horticultural Science* **103**: 151–155.
- Halevy, A.H. and Mayak, S. 1979. Senescence and postharvest physiology of cut flowers, Part 1. *Horticultural Reviews* **1**: 205-236.
- Halevy, A.H. and Mayak, S. 1981. Senescence and postharvest physiology of cut Flowers, Part 2. *Horticultural Reviews* **3**: 59-143.
- Halevy, A.H. 1976. Treatment to improve the water balance of cut flowers. *Acta Horticulturea* **64:** 223-230.



- Han, S.S. 1992. Role of sucrose in bud development and vase life of cut *Liatris spicata* (L.) Willd. *Horticultural Science* **27**: 1198–1200.
- Han, S.S. 2003. Role of sugar in the vase solution on postharvest flower and leaf guality of oriental lily 'Stargazer'. *Horticultural Science* **38**: 412–416.
- He, S., Joyce, D.C., Irving, D.E., Faragher, J.D. 2006. Stem end blockage in cut Grevillea 'Crimson Yul-lo' inflorescences. *Postharvest Biology and Technology* **41**: 78–84.
- Heuser, C. W. and Evensen, K.B. 1986. Cut flower longevity of peony. *Journal of the American Society for Horticultural Science* **111**: 896-899.
- Ho, L.L. and Nichols, R. 1977. Translocation of i4C-sucrose in relation to changes in carbohydrate content in rose corollas cut at different stages of development. *Annals of Botany.*, **41**: 227-242.
- Hoogerwerf, A. and Van Doom, W.G. 1992. Numbers of bacteria in aqueous solutions used for postharvest handling of cut flowers. *Postharvest Biology and Technology* 1: 295 304.
- Ichimura, K. and Hisamatsu, T. 1999. Effects of continuous treatment with sucrose on the vase life, soluble carbohydrate concentrations, and ethylene production of cut snapdragon flowers. *Journal of the Japan Society for Horticultural Science* **68**: 61–66.
- Ichimura, K., Kojima, K. and Goto, R. 1999. Effects of temperature, 8-hydroxyquinoline sulphate and sucrose on the vase life of cut rose flowers. *Postharvest Biology and Technology* **15:** 33–40.
- Ichimura, K.and Shimizu-Yumoto, H. 2007. Extension of the vase life of cut rose by treatment with sucrose before and during simulates transport. *Bulletin of National Institute Floriculture Science* **7**: 17–27.
- Ichimura, K., Suto, K. 1999. Effects of the time of sucrose treatment on vase life, soluble carbohydrate concentrations and ethylene production in cut sweet pea flowers. *Plant Growth Regulator* **28**: 117–122.

Joanna Bonarriva. USITC. 2003. Industry & Trade Summary, Cut flower.

- Jones, R.B. and Hill, M. 1993. The effect of germicides on the longevity of cut flowers. Journal of the American Society for Horticultural Science **118**: 350–354.
- Jong, L.H., Saad, Mohd. Ridzuan and Hamir, N.A. 2000. Cut Flower Production in Malaysia. *Malaysian Agricultural Research and Development Institute* (MARDI).



- Kaltaler, R.E.L. and Steponkus, P.L. 1974. Uptake and metabolism of sucrose in cut roses. *Journal of the American Society for Horticultural Science* **99**: 490–493.
- Kaltaler, R.E.L. and Steponkus, P.L. 1976. Factors affecting respiration in cut roses. Journal of the American Society for Horticultural Science **101**: 352–354.
- Kuiper, D., Ribot, S., Van Reenen, H.S.and Marissen, N. 1995. The effect of sucrose on the flower bud opening of 'Madelon' roses. Scientia *Horticulturae* **60**: 325–336.
- Liao, I.J., Lin, Y.H., Huang, K.L., Chen, W.S. and Cheng, Y.M. 2000. Postharvest life of cut life and flower and bud opening in *Polianthes tuberosa* using gibberellic acid and sucrose. *Australian Journal of Experimental Argiculture* **41**: 1227–1230.
- Markhart, A.H. and Harper, M.S. 1995. Deleterious effects of sucrose in preservative solutions on leaves of cut roses. *Horticultural Science* **30**: 1429-1432
- Marousky, F.J. 1969. Vascular blockage, water absorption, stomatal opening, and respiration of cut Better times roses treated with 8- hydroxyquinoline citrate and sucrose. *Journal of the American Society for Horticultural Science* **94**: 223–226.
- Maxie, E.C., Farnham, D.S., Mitchell, F.G. and Sommer, N.F. 1974. Temperature management effects on quality of carnation flowers and rosebuds. *California Agriculture* **28**: 6-7.
- Mayak, S., Halevy, A. H., Sagie, S., Bar-Yoseph, A and Bravdo. B. 1974. The water balance of cut rose flowers. *Plant Physiology*. **31**: 15-22.
- Mayak, S. and Kofranek, A. 1976. Altering the sensitivity of carnation flowers to ethylene *Journal of the American Society for Horticultural Science* **101**: 503-506.
- Muhammad, S.M., Hiroyasu, S. and Shahzad, N. 1996. *Diversity in roses*. National Agriculture Research Center, Islamabad, pp: 1-2.
- Nelson, P.V. 2003. *Greenhouse operation and Management*. Ed.6. Prentice Hall, United States of America.
- Nijsse, J., Van Meeteren, U. and Keijzer, C.J. 2000. Air in xylem vessels of cut flowers. *Acta Horticulturae*. 517, 479–486.
- Nooden, L.D., Guiamet, J.J. and John, L. 1997. Senescence Mechanism. *Physiologia* plantarum **101**: 746-753
- Ohkawa, K., Kasahara, Y. and Suh, J. 1999. Mobility and effects on vase life of silver containing compounds in cut rose flowers. *Horticultural Science* **34**: 112–113.
- Parups, E.V. and Chan, A.P. 1973. Extension of vase-life of cut flowers by use of isoascorbate-containing preservative solutions. *Journal of the American Society for Horticultural Science* **98**: 22–26.



- Patil, K. and Singh, J. 2007. Effects of Temperature and Light and Dark Conditions on Wilting of Cut Rose. *Postharvest Biology and Technology* **14**: 65–70
- Paull, R. E. 1982. Anthurium vase life evaluation criteria. *Horticultural Science* **17**: 606-607.
- Paull, R. E., Chen, N. J. and Deputy, J. 1985. Physiological changes as sociated with senescence of cut anthurium flowers. *Journal of the American Society for Horticultural Science* **110**: 156-162.
- Pertwee, J. 1995. *The Production and Marketing of Roses*. 2nd edition. Pathfast Publication Essex, England.
- Poovaiah, B. W. 1988. Calcium and senescence, *Senescence and aging in plants. Academic Press. San Diego, CA*: 369-389
- Poovaiah, B.W., Glenn, G.M. and Reddy, A.S.N. 1988. Calcium and fruit softening: physiology and biochemistry. *Horticultural Reviews* **10**: 107-152.
- Prince, T.A. and Tayama, H.K. 1988. Preservatives and fresh cut-flower longevity. *Ohio Florists' Association Bulletin* **706**: 34
- Put, H. and Clerkx, A.C.M. 1988. The infiltration ability of micro-organisms Bacillus, Fusarium, Kluyverumyces and Pseudomonas spp. into xylem vessels of Gerbera cv. 'Fleur' and Rosa cv. 'Sonia' cut flowers: a scanning electron microscope study. Journal of Applied Bacteriology 64: 515-530.
- Put, H. and Jansen, L. 1989. The effects on the vase life of cut *Rosa cultivar* 'Sonia' of bacteria added to the vase water. *Scientia Horticulturae* **39**: 167-179.
- Put, H. and Van der Meyden, T. 1988. Infiltration of *Pseudomonas putida* cells, strain 48, into xylem vessels of cut Rosa cv. 'Sonia'. *Journal of Applied Bacteriology* 64: 197-208.
- Put, H. 1990. Micro-organisms from freshly harvested cut flower stems and developing during the vase life of chrysanthemum, gerbera and rose cultivars. *Scientia Horticulturae* **43**: 129-144.
- Put, H.M.C. 1986. Investigations into the influence of the microflora from stems of cut flowers on the vase life of *Rosa cv.* Sonia, *Gerbera cv.* Fleur, and *Chrysanthemum cv.* Spider. *Acta Horticulturae* **181**: 415-418.
- Ranwala, A.P. and Miller, W.B. 2000. Preventive mechanisms of gibberellin and light on temperature-induced leaf senescence in Lilium cv. Stargazer. *Poshtharvest Biology and Technology* **19:** 85-92.



- Reid, M.S., Evans, R.Y. and Dodge, L.L. 1989. Ethylene and silver thiosulfate influence opening of cut rose flowers. *Journal of the American Society for Horticultural Science* 114: 436-440.
- Roberts, G.L., Tsujita, M.J. and Dansereau, B. 1993. Supplemental light quality affects rose flowers as affected by silver thiosulfate and sucrose. *Botanical Bulletin of Academia Sinica* **41**: 299–303.
- Roberts, S. B. 2000. "High-glycemic index foods, hunger, and obesity" *Nutrition Reviews* **58**: 163-169.
- Rod Jones. 2001. Caring for Cut Flowers. Australia: Landlinks Press
- Saichol, F. and Piluek, C. 1994. *Effect of Silver Nitrate and Silver Thiofosulfate on Vase Life of Cut Roses*. Thailand: Department of Horticulture.
- Salinger, J. P. 1975. Criteria for the evaluation of postharvest senescence of cut flowers. *Acta Horticulturae* **41**: 207-215.
- Scott, D., Boyle, T.H. and Han, S.S. 1994. Floral development and flower longevity in *Rhipsalidopsis* and *Schlumbergera* (Cactaceae). *Horticultural Science* 29: 898-900.
- Su,W.R., Huang, K.L., Chang, P.S. and Chen, W.S. 2001. Improvement of postharvest vase life and flower and bud opening in *Polianthes tuberosa* using gibberellic acid and sucrose. *Australian Journal of Experimental Agriculture* **41**: 1227–1230.
- Synge, P.M. 1971. The dictionary of rose in colour. 1st Edition., Madison Square Press, New York, ISBN-10: 0448025043 pp: 191.
- Teixeira da Silva, J.A. and Nhut, D.T. 2003. Thin Cell Layers (TCLs) and Floral Morphogenesis, Floral Genetics and *in vitro Flowering*. *Online Journal of Biological Science* **4**: 406-442
- Teixera da Silva, J.A. 2003. The cut Flower: Postharvest considerations. *Online Journal* of Biological Science **4**: 406-442
- Torre, S. and Fjeld, T. 2001. Water loss and postharvest characteristics of cut roses grown at high or moderate relative air humidity. *Scientia Horticulturae* **89:** 217-226
- Van Doorn, W.G. and Vojinovic, A. 1996. Petal Abscission in Rose Flower: Effects of Water Potential, Light Intensity and Light Quality. *Annals of Botany* **78**: 619-623
- Van Doorn, W.G. and Witte, D. 1991. Effect of dry storage on bacterial counts in the stems of cut flowers. *Horticulturae science* **26**: 1521- 1522



- Van Doorn, W.G. and Perik, R.R.J. 1990. Hydroxy-quinoline citrate and low pH prevent vascular blockage in the stems of cut rose flowers by reducing the number of bacteria. *Scientia Horticulturae* **115**: 979-981
- Van Doorn, W.G. and Vaslier, N. 2002. Wounding-Induced Xylem Occlusion in Stems of Cut Chrysanthemum Flowers: Roles of Peroxidase and catechol oxidase. *Postharvest Biology and Technology* 26: 275–284
- Van Doorn, W.G. 1990. Aspiration of air at the cut surface of rose stems and its effect on the uptake of water. *Journal of Plant Physiology*. **137**: 160–164.

Van Doorn, W.G. 1997. Water Relations of Cut Flowers. Horticultural reviews 18: 1-85.

- Van Doorn, W.G., Buis, H.C.E.M and De Witte, Y. 1986. Effect of exogenous bacterial concentrations on water relations of cut flowers. II. Bacteria in the vase solution. *Acta Horticulturea* 181: 463-467.
- Van Doorn, W.G.and Cruz, P. 2000. Evidence for a wounding-induced xylem occlusion in stems of cut chrysanthemum flowers. *Postharvest Biology and Technology* **19**: 73–83.
- Van Doorn, W.G., de Witte, Y. and Perik, R.R.J. 1990. Effect of Antimicrobial Compounds on the Number of Bacteria in Stems of Cut Rose Flowers. *Journal of Applied Bacteriology* 68: 117–122.
- Van Doorn, W.G., Schurer, K. and De Witte, Y. 1989. Role of endogenous bacteria in vascular blockage of cut rose flowers. *Journal of Plant Physiology* **134**: 375–381.
- Van Doorn, W.G. and Vojinovic. 1996. Petal Abscission in Rose Flowers: Effects of Water Potential, Light Intensity and Light Quality. *Botany* **78**: 619-623
- Wills, R.H.B. 2007. Postharvest: An Introduction to the Physiology and Handling of Fruit, Vegetables and Ornamentals. Fifth Edition. Australia : University of New South Wales Press.
- Yamada, K., Ito, M., Oyama, T., Nakada, M., Maesaka, M and Yamaki, S. 2006. Analysis of Sucrose Metabolism During Petal Growth of Cut Roses. *Postharvest Biology and Technology* 35: 76-84
- Zagory, D. and Reid, M.S. 1986. Role of vase solution microorganisms in the life of cut flowers. *Journal of the American Society for Horticultural Science* **111**: 154–158.
- Zieslin, N., kohl, H.C., Kofranek, A.M and Halevy, A.H. 1978. Changes in the water status of cut roses and its relationship to bent-neck phenomenon. *Journal of the American Society for Horticultural Science* **103 (2):** 176-179.

