

ISOLATION OF POTENTIAL ENDOPHYTIC BACTERIA OF OIL PALM  
AGAINST *GANODERMA BONINENSE*

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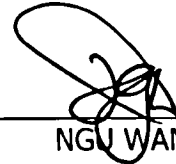
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## ABSTRACT

Fifteen culturable endophytic bacteria were isolated from symptomless roots of oil palm sampled from oil palm plantation site located at the School of Sustainable Agriculture, Sandakan campus of Universiti Malaysia Sabah. Characterizations of the bacterial isolates were carried out based on morphological characteristics, including colony colour, elevation, margin and the appearance displayed on nutrient agar and potato dextrose agar media. Dual culture test was carried out on PDA media to select isolate with antagonistic character against *Ganoderma boninense*. Results revealed degrees of inhibition toward mycelial growth of *G. boninense* vary accordingly to the bacterial isolates. Among the fifteen isolated endophytic bacteria, only two bacterial isolates (P<sub>15</sub> and P<sub>18</sub>) showed potential inhibitory effect against *G. boninense* with their percentage inhibitions of radial growth (PIRG) recorded were more than 50% in both dual culture and culture filtrate tests. There was significantly difference in inhibitory effect ( $P < 0.05$ ) for dual culture test and in culture filtrate test. The isolated P<sub>15</sub> and P<sub>18</sub> were further investigated in greenhouse trial. Results from *in planta* greenhouse trial further suggested that isolate P<sub>15</sub> and P<sub>18</sub> were capable to increase vegetative growth of oil palm seedlings while providing early protection against *G. boninense* infection. They play a role in keeping the *G. boninense* population below threshold for Basal Stem Rot (BSR) initiation by restricting its entry and movement in the seedlings.

# MENGESAN BAKTERIA ENDOFITIK DARIPADA KELAPA SAWIT YANG BERKESAN MELAWAN *GANODERMA BONINENSE*

## ABSTRAK

Lima belas bakteria endofitik telah diasingkan daripada akar kelapa sawit yang sihat daripada ladang kelapa sawit yang terletak di Sekolah Pertanian Lestari, kampus Sandakan, Universiti Malaysia Sabah. Pencirian bakteria telah dijalankan berdasarkan ciri-ciri morfologi, termasuk warna koloni, permukaan koloni dan corak koloni yang dipaparkan pada agar nutrien dan media agar PDA. Ujian dwi-kultur telah dijalankan pada media PDA untuk memilih bakteria endofitik dengan watak antagonis terhadap *Ganoderma boninense*. Keputusan menunjukkan darjah perencatan terhadap pertumbuhan mycelial *G. boninense* adalah berbeza antara bakteria. Antara lima belas bakteria endofitik yang diasingkan, hanya dua bakteria (P<sub>15</sub> dan P<sub>18</sub>) menunjukkan potensi perencatan terhadap *G. boninense* dengan peratusan perencatan pertumbuhan mycelial (PIRG) yang lebih daripada 50% dalam dua kedua-dua ujian dwi-kultur dan ujian turasan kultur. Terdapat perbezaan secara ketara ( $P < 0.05$ ) dalam kedua-dua ujian yang dijalankan. Seterusnya, bakteria endofitik telah diuji terhadap anak benih sawit. Keputusan dari rumah hijau (*in planta*) mencadangkan bahawa endofitik bakteria P<sub>15</sub> dan P<sub>18</sub> mampu untuk meningkatkan pertumbuhan vegetatif benih kelapa sawit di samping memberikan perlindungan awal terhadap jangkitan *G. boninense*. Bakteria ini memainkan peranan dalam mencegah *G. boninense* berkembang dengan menyekat kemasukan dan pergerakan dalam benih.

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## LIST OF SYMBOLS, UNITS AND ABBREVIATIONS

t ha <sup>-1</sup>	Tonnes per hectare
°C	Celsius
%	Percentage
β	Beta
γ	Gamma
Fe <sup>3+</sup>	Iron ion
kg	kilogram
m	meter
mL	milliliter
μL	microliter
mins	minutes
cm	centimeter
nm	nanometer
rpm	Revolutions per minute
v/v	Volume to Volume
ACC	1-aminocyclopropane-1-carboxylase
BRI	Biotechnology Research Institute
BSR	Basal Stem Rot
cfu	Colony forming unit
CO <sub>2</sub>	Carbon dioxide
CONH <sub>2</sub>	Carboxamide group
COOH	Carboxyl group
CRD	Complete Randomized Design
DDR	2,3-deepoxy-2,3-didehydrorhizoxin
DNA	DeoxyriboNucleic Acid
DSI	Disease Severity Index
G+C	Guanine-plus-Cytosine
HCN	Hydrogen Cyanide
HSD	Honestly Significant Different
IAA	Indole-3-acetic acid
ISR	Induced Systemic Resistance
LPS	Lipopolysaccharides
MPOB	Malaysia Palm Oil Board
NA	Nutrient Agar
PCA	Phenazine-1-carboxylic acid
PCN	Phenazine-1-carboxamide
PDA	Potato Dextrose Agar
PGPB	Plant Growth Promoter Bacterium
PGPR	Plant Growth Promoting Rhizobacteria
PIRG	The percentage inhibition of radial growth
PLT	Pyoluteorin
PR	Pathogenesis-related
PRN	Pyrrrolnitrin
PVC	Polyvinyl Chloride
PVD	Pyoverdine
rDNA	Ribosomal DeoxyriboNucleic Acid
rRNA	Ribosomal Ribonucleic Acid
SA	Salicylic Acid

## LIST OF SYMBOLS, UNITS AND ABBREVIATIONS

SAR	Systemic Acquired Resistance
SDW	Sterile Distilled Water
SPSS	Statistical Package for the Social Science
SSA	School of Sustainable Agriculture
TCBS	Thiosulphate citrate bile salt sucrose (II)
UMSKS	University Malaysia of Sabah Campus Sandakan
UV	Ultra violet

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## CHAPTER 1

### INTRODUCTION

#### 1.1 Introduction

Oil palm (*Elaeis guineensis* Jacq.) is the most crucial species in the genus of *Elaeis* which belongs to the family of *Palmae*. Oil palm is indigenous to West Africa where it distribute in large range of area from Sierra Leona, Liberia, the Ivory Coast, Ghana and Cameroon to the equatorial regions of the Republics of Congo and Zaire. The first introduction of African oil palm seedlings were planted in the Botanic Garden in Bogor in 1848 (Teoh, 2002). The first commercial oil palm plantation was established in Sumatra, Indonesia. In 1911, the oil palm seeds were brought by Fauconnier and planted in Rantau Panjang Estate in Selangor, Malaysia (Teoh, 2002).

Today, oil palm is the most important plantation crop in Malaysia. It produces with an average yield of approximately 4 t ha<sup>-1</sup> annually. Oil palm industry contributed substantially to the Malaysian's economy. The oil palm exportation reached about RM31 billion increased by 8.54% or RM2.44 billion in 2006 compared to 2005 (Najmie, 2011). According to Basiron (2007), the areas of oil palm plantation had increased from 54,000 hectares in 1960 to 4.05 million hectares in 2005. This represents a compound annual growth of 10.06%. While, the production from oil palm industry increased from 94,000 tones in 1960 to 15 million tones in 2005, reflecting a compound annual growth of 11.93% per year (Basiron, 2007).

In Malaysia, the oil palm is blessed by being largely disease free, but suffering from one major disease known as the Basal Stem Rot (BSR) caused by *Ganoderma boninense*. It is the major disease in oil palm plantation with no known effective cure at present. The economic life span of oil palm is 25-30 years. BSR can kill more than

80% of stands by the time they are half-way through normal economic life (Abdul Razak *et al.*, 2004). Palms infected with *G. boninense* produce yield 21% less than healthy palms at the same age (Nazzeb *et al.*, 2000). Singh (1990) found that heavily infected field yielded 26% less at 11 years after planting and 46% less at 15 years by which time incidence was 67%.

There is currently no effective cure for *G. boninense* infection in an existing stand. Field control of BSR by contact chemicals are not very successful (Soepena *et al.*, 2000) even with those *in vitro* screened effective against the fungus (Khairudin, 1990). Control by physical methods such as clean clearing and tree surgery had transient effects though there is testify that BSR can be eliminated if all the disease inoculum is removed before planting or replanting the crop. Preventive and ameliorative treatments such as biological control are commonly carried out in the oil palm plantation. Therefore, biological control can be a cure for the BSR disease and can be merely to arrest the disease spread by inoculation with a biological control agent that could guard from the plant inside out (endophytes).

Endophytes are microorganisms that live within the plants cells. The functions of endophytic microorganisms should be preferable to other biological control agents as they are colonizers internally and therefore more able to compete in vascular systems, depriving *Ganoderma* from both nutrients and space for its proliferation. Astute observations of the low incidence of disease due to pathogenic *Ganoderma* species in some natural stands, suggest that the disease is most likely kept under control by some biological means. Due to these observations, recent control measures to overcome the BSR incidence are now focused on the use of biological control agents. For example, saprophytes can be used to compete against *Ganoderma boninense* to reduce its opportunity for colonizing oil palm roots. Several promising antagonists such as *Trichoderma* (Shukla and Uniyal, 1989; Wijesekera *et al.*, 1996; Sariah, 2003) and *Penicillium* (Dharmaputra *et al.*, 1989) have shown *in vitro* antagonistic activity against *G. boninense*.

## 1.2 Justification of Study

Endophytes are organisms inhabiting plant organs that at some time in their life cycle can colonize the internal plant tissue without causing apparent harm to the host. Screening the potential endophytes in the oil palm roots and re-introducing them to the infected oil palm can theoretically lead to enhance suppression of BSR disease. The use of endophytic bacteria are preferred to other biological control agents as they are internal colonizers, with better ability to compete within the vascular system and limiting *G. boninense* for both nutrient and space during its proliferation. Besides that, the growth of the plant by endophytes can be stimulated through the promoting of nitrogen fixation, production of phytohormones and by enhancing available ability of minerals. Once the selection of the most potential endophytic bacteria, it can be inoculated into the roots of the oil palm seedlings for the manipulation of the indigenous bacterial communities in order to enhance the suppression of soil-borne pathogen and *Ganoderma* as well. Endophytes create least residue to the environment as compared to chemical control methods. Endophytes can be the important biological control agents which they are environmental friendly and there is no negative effect on both oil palm trees and the soil. Besides of suppressing of BSR disease incidence, the endophytes can promote the growth of the oil palm as well. The endophytes can be incorporated into the bio-fertilizers and it will be important and economical supplements for the oil palm trees.

## 1.3 Objectives

The objectives of this study are:

1. To isolate the potential endophytic bacteria in the roots of symptomless palms.
2. To evaluate the efficiency of endophytic bacteria in suppression of *G.boninense* *in vitro* (culture plate).
3. To evaluate the efficiency of endophytic bacteria in suppression of *G.boninense* *in planta* (nursery).
4. To determine the effects of endophytic bacteria on seedling growth of oil palm.

## 1.4 Hypotheses

The hypotheses for this study are:

H<sub>01</sub>: There is no significant effect of the endophytic bacteria on *Ganoderma boninense* *in vitro* and *in planta* and the growth of the oil palms seedlings.

H<sub>a1</sub>: There is significant effect of the endophytic bacteria on *Ganoderma boninense* *in vivo* and *in planta* and the growth of the oil palms seedlings.

H<sub>02</sub>: There are is significant effect of the endophytic bacteria on the growth of the oil palms seedlings after the application of treatments.

H<sub>a2</sub>: There is significant effect of the endophytic bacteria on the growth of the oil palms seedlings after the application of treatments.

## CHAPTER 2

### LITERATURE REVIEWS

#### 2.1 Oil Palm (*Elaeis guineensis* Jacq.)

*Elaeis guineensis* Jacq. which is commonly known as oil palm is the most important species in the genus *Elaeis* (family: *Palmae*). The second species is *Elaeis oleifera* Cortes which is found in South and Central America and is known as the American oil palm. Oil palm was the major commodity inside the Malaysian agriculture sector. Commercial cultivation of the oil palm started in 1917 and has expanded tremendously in recent years because it is suitable to be planted in most lands in Malaysia.

Malaysia is presently the world's leading exporter of oil palm having a 60% market share and oil palm was second only to soybean as the major source of vegetable oil. The total oil palm planted area in the country increased by 4.3% to 4.48 million hectares in 2008. The expansion in planted area occurred mainly in Sabah and Sarawak with a combined growth of 7% compared to 2% in Peninsular Malaysia. Sabah remained the largest oil palm planted state, accounting for 1.33 million hectares or 30% of the total planted area in the country. However, a soil fungus pathogen, *Ganoderma boninense*, which causes basal stem rot in oil palms trunk, ruins thousands of hectares of plantations in Southeast Asia every year. The disease causes infected palm trunks to fracture at the base, thus causing direct loss of oil palm trees.

#### 2.2 Basal Stem Rot (BSR) Disease

*Ganoderma boninense* is the causal agent of basal stem rot (BSR) disease which is the major disease in oil palm plantation in Malaysia (Singh, 1990). Previously, BSR was only found on older plants, but recently the disease was observed on five-year-old or

younger plants. According to Singh (1990), the BSR was found to infect the palms as young as 12-24 months after planting in the estate and 4-5 years after planting had been reported was the most increased incidence of the infection. Beside the very old palms, palms between 7-15 years old are also infected. The BSR disease is presently in most oil palm plantation particularly in the coastal marine clay areas (Khairudin, 1990), peat, localized inland soils (Ariffin *et al.*, 1993; Rao, 1990) and replanted areas (Singh, 1990).

The *G. boninense* problems had been known for decades, but the search for solution was considerably hampered by a natural constraint. The disease does not cause external symptoms until it is too far advanced and at a stage when trees cannot respond to treatment anymore. Primary infection of palms by *Ganoderma* species has been considered to occur by contact of living palm roots with colonized debris within the soil (Idris *et al.*, 2002). Secondary spread of inoculums has been assumed to be by contact of living palm roots with each other (De Oliveira *et al.*, 2005).

### **2.2.1 Infection of *Ganoderma boninense***

Rees *et al.* (2009) reported a detailed demonstration of the reproducible infection of intact roots of oil palm with *G. boninense*. Infection showed penetration, followed by rapid longitudinal progression of hyphae and colonisation of the lower stem (bole) of oil palm. In newly colonised tissue, *G. boninense* behaved as a hemibiotroph with numerous, wide, intracellular hyphae occupying entire host cells that possessed intact cell walls and contained discernible cytoplasm and organelles. In the bole this phase coincided with a complete depletion of previously abundant starch grains in advance of invasion. Subsequently, in the roots and colonised stem bases, widespread necrotrophic, enzymatic attack of all layers of the host cell walls occurred. Hyphae were intra, intercellular and intramural and associated with host cell wall degradation, which was often at a distance from hyphae, resulting in cavities within cell walls.

A third developmental stage was the formation of an extensive, melanized, tough mycelium, or pseudo-sclerotium, which surrounded roots and comprised many very thick-walled cells encasing more typical thin-walled hyphae. Macroscopic observation of and isolation from the bole of randomly felled, commercial palms

provided confirmatory evidence that multiple infections originated in the roots before spreading into the base of long-established palms (Rees *et al.*, 2009).

### **2.2.2 Control of basal stem rot disease**

Attempts to control this disease in the fields with fungicide have been made by various workers, but the results are inconclusive, though some systematic fungicides seem to be promising. The methods of fungicide application include soil drenching, trunk injection, or combination of these two methods (Erwinsyah, 2008). However, the results of fungicidal control of this disease in the field have been inconclusive (Idris *et al.*, 2002). This phenomenon is probably due to the fact that *Ganoderma* has various resting stages such as melanised mycelium, basidiospores and pseudosclerotia (Susanto *et al.*, 2005) that are more resistant to fungicides. On the other hand, once external symptoms appear the infection is already too severe, and a biocontrol agent cannot control the pathogen during this stage.

### **2.3 Endophytic Bacteria**

Endophytes are the microorganisms that live within or at least during part of their life cycle inside a plant. Cultivable endophytic communities can be isolated after surface sterilization of plant materials. They can generally be found within the cells, in the intercellular spaces and in the vascular system (Aravind *et al.*, 2009). The endophytes can also be found in various tissues such as tubers, fruit, stems, seeds and ovules. Zaiton (2006) discovered that among 1,323 microbial endophytes isolated from 3,600 oil palm roots, 65.23% were bacteria, 32.73% fungi and 2.04% actinomycetes. As cited in the extensive review of Kobayashi and Palumbo (2000), both gram-positive and gram-negative bacterial endophytes have been isolated from several tissue types in numerous plant species. However, most of the endophytes isolated from xylem vessels (about 78 to 84% of the population) were Gram-negative (Gardner *et al.*, 1982). Ranges of 93.04% to 100% of the isolated endophytic bacteria from oil palm roots were Gram negative and only 6.95% are Gram positive (Zaiton, 2006; Rahamath *et al.*, 2010). There are different mechanisms by which the endophytes enter and then colonize the plants such as the entry through stomata, lenticels, wounds including broken trichomes, the emerge of lateral roots areas, germinating radicle, undifferentiated meristematic root tissue, the penetration of the junction or cracks

between root hair and adjacent epidermal cells and the enzymatic processes involving degradation of cell wall bound polysaccharides (Quadt-Hallmann *et al.*, 1997).

The common bacterial species of endophytes reside in the plants are Fluorescent Pseudomonads, *Bacillus* spp., *Herbaspirillum* spp., *Serratia marcescens* and *Streptomyces* spp. (McLnroy *et al.*, 1995). Some of the endopyhtic bacteria from genera *Pseudomonas*, *Burkholderia* and *Serratia* are mostly found in healthy roots from symptomless palms. Study proved that in most plant the population densities of the endophytes are the greatest in the plant roots with the densities ranging from  $10^4$  to  $10^6$  colony forming unit (cfu) per g fresh weight in cotton and sweet corn roots (McLnroy *et al.*, 1995).

The population density of endophytes is highly variable by which it is mostly depending on the bacterial species, the host genotype as well as the host development stage, inoculum density and the environmental condition (Pillay *et al.*, 1997; Tan *et al.*, 2003). In oil palm, the population of endophytes varies due to the different palms age. Endophytes are more in mature (more than 11 years) followed by middle-aged (6-10 years) and young (1-5 years) palms (Zaiton, 2006). There is less significant different in the endophytes population in different types of soil such as coastal and peat soils. However, the inland soils there are high abundance of the endophytes with the genera of *Pseudomonas* and *Burkholderia* (Zaiton, 2006).

### **2.3.1 Fluorescent pseudomonads**

Fluorescent pseudomonads are non-enteric, Gram-negative, aerobic, straight, or slightly curved rods, which are non-fermenting and mobile belonging to  $\gamma$  - proteobacteria (Galli *et al.*, 1992). They are pervasive bacteria which are inhabitants of soil, water and phyllosphere commonly but predominant in plant rhizosphere due to the exudation of organic acids, sugars and amino acids (Lugtenberg and Dekkers 1999). Fluorescent pseudomonads are the most promising group of plant growth-promoting rhizobacteria (PGPR) involved in biological control of plant disease. Saprophytic fluorescent pseudomonads are typical inhabitants of agricultural field soils and plant rhizosphere and are involved in several interactions with plants (Schroth *et al.*, 1992). They are capable of utilizing many plant exudates as nutrient (Lugtenberg *et al.*, 1999) and are known to possess important traits in bacterial fitness such as the



## REFERENCES

- Abd-Alla, M. H. 1998. Growth and siderophore production *in vitro* of *Bradyrhizobium* (Lupin) strain under iron limitation. *Eur. J. Soil. Biol.* **34**:99-104
- Abdul Razak, J., Ahmad, H., Ramdhan, K., Idris, A.S., Rahim, A.S., Aminul, R. and Fauzi, I. 2004. Mechanical Trunk Injection for Control of *Ganoderma*. *MPOB Information Series* TT No.215
- Abdullah, F., Ilias, G.N.M., Nelson, M., Nur Ain Izzati, M. Z. and Umi Kalsom, Y. 2003. Disease Assessment and The Efficacy of Trichoderma as a Biocontrol Agent of Basal Stem Rot of Oil Palm. *Research Bulletin Science Putra* **11**:31-33
- Acea, M. L. and Alexander, M. 1988. Growth and Survival of Bacteria Introduced into Carbon Amended Soil. *Soil Biol. Biochem.* **20**:703-709
- Anjaiah, V., Koedam, N., Nowak-Thompson, B., Loper, J. E., Hofte, M., Tambong, J. T. and Cornelis, P. 1998. Involvement of Phenazines and Anthranilate in the Antagonism of *Pseudomonas aeruginosa* PNA1 and Tn5 Derivatives toward *Fusarium* spp. and *Pythium* spp. *Mol. Plant Microbe Interact* **11**:847-854
- Aravind, R., Dinu, A., Santosh, J. E., Kumar, A. and Ramana, K. V. 2009. Isolation and Evaluation of Endophytic Bacteria against Plant Parasitic Nematodes Infesting Black Pepper (*Piper nigrum* L.). *Indian Journal of Nematology* **39**: 211-217
- Ariffin, D., Idris, A. S. and Khairudin, H. 1993. Confirmation of *Ganoderma* Infected Palm by Drilling Technique. *Palm Oil Research Institute of Malaysia* 735-738
- Arima, K., Imanaka, H., Kousaka, M., Fukuda, A. and Tamura, G. 1964. Pyrrolnitrin, a New Antibiotic Substance Produced by *Pseudomonas*. *Agric. Biol. Chem.* **28**:575-576
- Ayyadurai, N., Ravindra Naik, P. and Sakthivel, N. 2007. Functional Characterization of Antagonistic Fluorescent *Pseudomonads* Associated with Rhizospheric Soil of Rice (*Oryza sativa* L.). *J Microbial Biotechnol* **17**:919-927
- Ayyadurai, N., Ravindra Naik, P., Sreehari Rao, M., Sunish Kumar, R., Samrat, S. K., Manohar, M. and Sakthivel, N. 2006. Isolation and Characterization of a Novel Banana Rhizosphere Bacterium as Fungal Antagonist and Microbial Adjuvant in Micropropagation of Banana. *J. Appl Microbial* **100**:926-937
- Azadeh, B. F., Sariah, M. and Wong, M. Y. 2010. Characterization of *Burkholderia cepacia* genomovar I as a Potential Biocontrol Agent of *Ganoderma boninense* in Oil Palm. *African Journal of Biotech.* **9**:3542-3548
- Aziz, A., Martin-Tanguy, J. and Larher, F. 1997. Plasticity of Polyamine Metabolism Associated with High Osmotic Stress in Rape Leaf Discs and with Ethylene Treatment. *Plant Growth Regul* **21**:153-163
- Bahme, J. B. and Schroth, M. N. 1987. Spatial-temporal Colonization Patterns of a Rhizobacterium on Underground Organs of Potato. *Phytopathology* **77**:1093-1100

- Baligh, M., Conway, K. E. and Delgado, M. A. 1991. Development of Bioassay System to Compare Quantities of Ammonia Produced by Strains Of *Pseudomonas* spp. Potential Biocontrol Agents Soil-Borne Fungi. *J. Plant Pathol* **10**:1228
- Baligh, M., Delgado, M. A. and Conway, K. E. 1999. Evaluation of *Burkholderia cepacia* Strains: Root colonization of *Catharanthus roseus* and *in-vitro* Inhibition of Selected Soil-Borne Fungal Pathogens. *Proc. of Oklahoma Acad. Sci.*, **79**: 19–27
- Barea, J., Navarro, M. and Montoya, E. 1976. Production of Plant Growth Regulators by Rhizosphere Phosphate-Solubilizing Bacteria. *J. Appl. Bacteriol* **40**:129-134
- Bartnicki-Garcia, S. and Lippman, E. 1973. Fungal Cell Wall Composition. In Laskin, A. L. and Lechevaluer, H. L.(Eds.). *Handbook of Microbiology*. Cleveland: Chemical Rubber
- Basiron, Y. 2007. Palm Oil Production through Sustainable Plantations. *European J. Lipid Sci. Technol.* **109**: 289-295
- Bennett, I., Broom, N. J. P., Cassels, R., Elder, J. S., Masson, N. D. and O'Hanlon, P. J. 1999. Synthesis and Antibacterial Properties of  $\beta$ -Diketone Acrylate Bioisosteres of Pseudomonic Acid A. *Bioorg. Med. Chem. Lett.* **9**:1847-1852
- Berg, G., Krechel, A., Ditz, M., Sikora, R. A., Ulrich, A. and Hallmann, J. 2005. Endophytic and Ectophytic Potato-associated Bacterial Communities Differ in Structure and Antagonistic Function against Plant Pathogenic Fungi. *Microbiol.Ecol.* **51**: 215-229
- Bevivino, A., Sarrocco, S., Dalmastrri, C., Tabacchioni, S., Cantale, C. and Chiarini, L. 1998. Characterization of a Free-Living Maize-Rhizosphere Population of *Burkholderia cepacia* : Effect of Seed Treatment on Disease Suppression and Growth Promotion of Maize. *FEMS microbial. Ecol.* **40**:115-118
- Bitter, W., Marugg, J. D., De Weger, L. A., Tommassen, J. and Weisbeek, P. J. 1991. The Ferric Pseudobactin Receptor *PupA* of *Pseudomonas putida* WCS358: Homology to *TorB* Dependent *Escherichia coli* Receptors and Specificity of the Protein. *Mol. Microbial* **5**:647-655
- Budzikiewicz, H. 1993. Secondary Metabolites from Fluorescent Pseudomonads. *FEMS microbial Ecol.* **104**:209-228
- Burd, G. I., Dixon, D. G. and Glick, B. R. 1998. A plant Growth Promoting Bacterium that Decrease Nickel Toxicity in Seedlings. *Appl. Environ. Microbiol.* **64**:3663-3668
- Buysens, S., Heungens, K., Poppe, J. and Hofte, M. 1996. Involvement of Pyochelin and Pyoverdine in Suppression of *Pythium*-Induced Damping Off of Tomato by *Pseudomonas aeruginosa* 7NSK2. *Appl. Environ. Microbiol* **62**:865-871
- Campbell, R., Renwick, A. and Coe, S. K. A. M. 1986. Antagonism and Siderophore Production by Biocontrol Agents, Plant Growth Promoting Organism and the General Rhizosphere Population. In Swinburne, T. R. (Ed.). *Iron, Siderophores and Plant Disease*. New York: Plenum

- Castric, P. 1994. Influence of Oxygen on the *Pseudomonas aeruginosa* Hydrogen Cyanide Synthase. *Curr. Microbiol* **29**:19-21
- Castric, P. A. 1981. The Metabolism of Hydrogen Cyanide by Bacteria. In Vennesland, B., Conn, E. E., Knowles, C. J., Westley, J. and Wissing, F. (Eds.). *Cyanide in Biology*. London:Academic
- Chet, I. 1987. *Trichoderma* Applications, Mode of Action and Potential as a Biocontrol Agent of Soilborne Plant Pathogenic Fungi. In Chet, I. (Ed.). *Innovative Approaches to Plant Diseases*. New York: Wiley
- Chiarini, L., Bevivino, A., Dalmastrri, C., Tacchioni, S. and Visca, P. 2006. *Burkholderia cepacia* Complex Species: Health Hazards and Biotechnological Potential. *Trends Microbiological*. **14**:277-286
- Chin-A-Woeng, T. F. C., Bloemberg, G. V., Van der Bij, A. J., Van der Drift, K. M. G. M., Schripse-ma, J., Kroon, B., Scheffer, R. J., Keel, C., Bakker, P. A. H. M., Tichy, H., de Bruijn, F. J., Thomas-Oates, J. E. and Lugtenberg, B. J. J. 1998. Biocontrol by Phenazine-1-carboxamide Producing *Pseudomonas chlororaphis* PCL 1391 of Tomato Root Rot Caused by *Fusarium oxysporum* f. sp. *radicis-lycopersici*. *Mol. Plant Microbe Interact* **11**:1069-1077
- Chong, K. P., Rossall S. and Markus, A. 2009. In Vitro Synergy Effect of Syringic Acid, Caffeic Acid and 4-hydroxybenzoic Acid against *Ganoderma boninense*. *International Journal of Engineering and Technology* **1(4)**:282-284
- Coombs, J. T., Michelsen, P. P. and Franco, C. M. M. 2004. Evaluation of Endophytic Actinobacteria as Antagonists of *Gaeumannomyces graminis* var. *Bio.Control* **29**:359-366
- Cox, C. D., Rinehart, K. L., Moore, M. L. and Cook, J. C. 1981. Pyochelin: Novel Structure of an Iron-chelating Growth Promoter for *Pseudomonas aeruginosa*. *Proc. Natl. Acad. Sci. USA* **78**:4256-4260
- De Freitas, J. R. and Germida, J. J. 1991. *Pseudomonas cepacia* and *Pseudomonas putida* as Winter Wheat Inoculants for Biocontrol of *Rhizoctania solani*. *Can J. Microbiol*. **37**:780-784
- De Oliveira, F. G. R., Candian, M. and Lucchette, F. F. 2005. A Technical Note on the Relationship between Ultrasonic Velocity and Moisture Content of Brazilian Hardwood (*Goupia glabra*). *Building and Environment* **40**: 297-300
- De Weger, L. A., Van Boxtel, R., van der Burg, B., Gruters, R. A., Geels, F. P., Schippers, B. and Lugtenberg, B. 1986. Siderophores and Outer Membrane Protein of Antagonistic, Plant Growth Stimulating Root-Colonizing *Pseudomonas* spp. *J. Bacteriol* **165**:585-594
- Demange, P., Wendenbaum, S., Linget, C., Bateman, A., MacLeod, J., Dell, A., Albrecht, A. M. and Abdallah, M. A. 1987. Bacterial Siderosphores: Structure and Physicochemical Properties of Pyoverdines and Related Compounds. In Winkelmann, G., van der Helm, D. and Neilands, J. B. (Eds.). *Iron Transport in Microbes, Plant and Animals*. Weinheim:VCH

- De Weger, L. A., Dekkers, L. C., van der Bij, A. J. and Lugtenberg, B. J. J. 1994. Use of Phosphate-Reporter Bacteria to Study Phosphate Limitation in the Rhizosphere and In Bulk Soil. *Mol Plant Microbe Interact* **7**:32-38
- Dekkers, L. C., Phoelich, C. C., Fits, L. V. D. and Lugtenberg, J. J. 1998. A Site-specific Recombinase is Required for Competitive Root Colonization by *Pseudomonas fluorescens* WCS365. *Proc. Natl. Acad. Sci. USA* **95**: 7051-7056
- Dharmaputra, O. S., Tjitrosomo, H. S. and Abadi, A. L. 1989. Antagonistic Effect of Four Fungal Isolates to *Ganoderma boninense*, The Causal Agent of Basal Stem Rot of Oil Palm. *J. Biotrop.* **3**: 41-49
- Dinesh, K. M. 2011. *Bacteria in Agrobiolgy: Plant Growth Response*. Gurukul Kangri University. New York: Springer
- Elad, Y and Baker, R. 1985. Influence of Trace Amounts of Cations and Siderophore-Producing Pseudomonads on Chlamyospore Germination of *Fusarium oxysporum*. *Ecol. Epidemiol.* **75**:1047-1052
- Elad, Y and Chet, I. 1987. Possible Role of Competition for Nutrient in Biocontrol of *Pythium* Damping Off by Bacteria. *Phytopathology* **77**:190-195
- Ellander, R. P., Mabe, J. A., Hamill, R. H. and Gorman, M. 1968. Metabolism of Tryptophan by *Pseudomonas aureofaciens* VI. Production of Pyrrolnitrin by Selected *Pseudomonas* species. *Appl. Microbiol.* **16**:753-758
- Ellis, R. J., Timms-Wilson, T. M. and Bailey, M. J. 2000. Identification of Conserved Traits in Fluorescent Pseudomonads with Antifungal Activity. *Environ. Microbiol* **2**:274-284
- El-Sayed, A. K., Hothersall, J., Cooper, S. M., Stephens, E., Simpson, T. J. and Thomas, C. M. 2003. Characterization of the Mupirocin Biosynthesis Gene Cluster from *Pseudomonas fluorescens* NCIMB 10586. *Chem. Biol.* **10**:410-430
- Erwinsyah. 2008 . Improvement of Oil Palm Wood Properties Using Bioresin. In Dinesh, K (Eds). *Bacteria in Agrobiolgy: Plant Growth Response*. New York: Springer
- Flaishman, W., Eyal, Z., Voisard, C. and Haas, D. 1990. Suppression of *Septoria tritici* by Phenazine or Siderophore Deficient Mutants of *Pseudomonas*. *Curr. Microbiol* **20**:121-124
- Foster, R. C. 1988. Microenvironment of Soil Microorganisms. *Biol Fert Soils* **6**:189-203
- Fridlender, M., Inbar, J. and Chet, I. 1993. Biological Control of Soilborne Plant Pathogens by a  $\beta$ -1,3 Glucanase-producing *Pseudomonas cepacia*. *Soil Biol. Biochem.* **25**:1121-1221
- Funaki, M., Tsuchiya, F., Maeda, K. and Kamiya, T. 1958. Cyanomycin, a New Antibiotic. *J. Antibiot. Ser. A.* **11**:143-149
- Galli, E., Barbieri, P. and Bestetti, G. 1992. Potential of *Pseudomonas* in the Degradation of Methylbenzenes. In Galli, E., Silver, S. and Withhold, B. (Eds.). *Pseudomonas: Molecular Biology and Biotechnology*. Washington: American Society for Microbiology

- Gamble, T. N., Betlach, M. R. and Tiedje, J. M. 1977. Numerically Dominant Denitrifying Bacteria from World Soils. *Appl. Environ. Microbiol* **33**:926-939
- Garcia de Salamone, I. E., Hynes, R. K. and Nelson, L. M. 2001. Cytokinin Production by Plant Growth Promoting Rhizobacteria and Selected Mutants. *Can. J. Microbiol.* **47**:404-411
- Gardner, J. M., Feldman, A. W. and Zablotowicz, R. M. 1982. Identity and Behavior of Xylem –Residing Bacteria in Rough Lemon Roots of Florida Citrus Trees. *Applied and Environmental Microbiology* **43**: 1335-1342
- Ghiglione, J. F., Gourbiere, F., Potier, P., Philippot, L. and Lensi, R. 2000. Role of Respiratory Nitrate Reductase in Ability of *Pseudomonas fluorescens* YT101 to Colonize the Rhizosphere of Maize. *Appl. Environ. Microbiol* **66**:4012-4016
- Glick, B. R. 1995. The Enhancement of Plant Growth by Free Living Bacteria. *Can. J. Microbiol* **41**:109-117
- Glick, B. R., Patten, C. L., Holguin, G. and Penrose, D. M. 1999. *Biochemical and Genetic Mechanisms Used by Plant Growth Promoting Bacteria*. London: Imperial College Press
- Glick, B. R., Penrose, D. M. and Li, J. 1998. A Model for the Lowering of Plant Ethylene Concentrations by Plant Growth Promoting Bacteria. *J. Theor. Bio.* **190**:63-68
- Grichko, V. P. and Glick, B. R. 2001. Amelioration of Flooding Stress by ACC Deaminase Containing Plant Growth Promoting Bacteria. *Plant Physiol Biochem.* **39**:11-17
- Grifoni, A., Bazzicalupo, M., Di Serio, C., Farcelli, S. and Fani, R. 1995. Identification of *Azospirillum* strains by Restriction Fragment Length Polymorphism of 16s rDNA and the Histidine Operon. *FEMS Microbiol. Lett.* **127**:85-91
- Gurusiddaiah, S., Weller, D. M., Sarkar, A. and Cook, R. J. 1986. Characterization of an Antibiotic Produced by a Strain of *Pseudomonas fluorescens* Inhibitory to *Gaeumannomyces graminis* var. *tritici* and *Pythium* spp. *Antimicrob. Agents Chemother* **29**:488-495
- Hallmann, J., Quadt-Hallmann, A., Mahaffee and Kloepper, J. W. 1997. Bacterial Endophytes in Agricultural Crops. *Canadian Journal of Microbiology* **43**: 8895-914
- Hasegawa, S., Meguro, A., Shimizu, M., Nishimura, T. and Kunoh, H. 2006. Endophytic Actinomycetes and Their Interactions with Host Plants. *Actinomycetologica.* **20**:72-81
- Hattori, T. 1988. Soil Aggregates as Microhabitants of Microorganism. *Rep. Inst. Agric. Res. Tohoku Univ.* **37**:23-36
- Hurek, T., Handley, L. L., Reinhold, H. B. and Piche, Y. 2002. Azoarcus Grass Endophytes Contribute Fixed Nitrogen to the Plant in an Unculturable State. *Plant-Microbe Interact* **15**: 233-242
- Idris, A. S., Ismail, S., Arifin, D. and Ahmad, H. 2002. Control of *Ganoderma* Infected Palm–Development of Pressure Injection and Field Applications. *MPOB Information Series TT No. 131*.

- Ilias, G.N.M. 2000. Trichoderma and Its Efficacy as a Bio-Control Agent of Basal Stem Rot of Oil Palm (*Elaeis guineensis* Jacq.). Ph.D. Thesis of Universiti Putra Malaysia, Selangor, Malaysia.
- Inbar, J. and Chet, I. 1991. Evidence that Chitinase Produced by *Aeromonas Caviae* is Involved in the Biological Control of Soil-Borne Plant Pathogens by this Bacteria. *Soil Biol. Biochem.* **23**:973-978
- Iniguez, A. L., Dong, Y. and Triplett, E. W. 2004. Nitrogen Fixation in Wheat Provided by *Klebsiella pneumoniae*. *Plant-Microbe Interact* **17**: 1078-1085
- Jackson, M. B. 1993. Are Plant Hormones Involved in Root to Shoot Communication? *Adv. Bot. Res.* **19**:103-187
- Jha, B. K., Pragash, M. G., Cletus, J., Raman, G. and Sakthivel, N. 2009. Simultaneous Phosphate Solubilization Potential and Antifungal Activity of New Fluorescent Pseudomonad Strains, *Pseudomonas cepacia*. *Plant Dis.* **25**:573-581
- Jinantana, J. and Sariah, M. 1997. Antagonistic Effect of Malaysian Isolates of *Trichoderma harzianum* and *Gliocladium Viren* on *Sclerotium rolfsii*. *Pertanika: J. Trop. Agric. Sci.* **20**: 35-41
- Katznelson, H. and Bose, B. 1959. Metabolic Activity and Phosphate-dissolving Capability of Bacterial Isolates from Wheat Roots, Rhizosphere and Non-rhizosphere Soil. *Can. J. Microbial.* **5**:79-85
- Keel, C., Schnider, U., Maurhofer, M., Voisard, C., Laville, J., Burger, P., Wirthner, P., Haas, D. and Defago, G. 1992. Suppression of Root Disease by *Pseudomonas fluorescens* CHA0: Importance of the Bacterial Secondary Metabolite, 2,4-diacetylphloroglucinol. *Mol. Plant Microbe Interact* **5**:4-13
- Khairudin, H. 1990. Results of Four Trials on *Ganoderma* Basal Stem Rot of Oil Palm in Goldenn Hope Estates. *PORIM* 113-131
- Kirner, S., Hammer, P. E., Hill, D. S., Altmann, A., Fischer, I., Weislo, L. J., Lanahan, M., van Pee, K. H. and Ligon, J. M. 1998. Functions Encoded by Pyrrolnitrin Biosynthetic Genes from *Pseudomonas fluorescens*. *J. Bacteriol* **180**:1939-1943
- Kobayashi, D. Y. and Palumbo, J. D. 2000. Bacterial Endophytes and Their Effects on Plants and Uses in Agriculture. In Dines, K. M. (ed.). *Bacteria in Agrobiolgy: Plant Growth Response*. New York: Springer
- Kraus, J. and Loper, J. E. 1995. Characterization of a Genomic Region Required for Production of the Antibiotic Pyoluteorin by the Biological Control Agent *Pseudomonas fluorescens* Pf-5. *Appl Environ Microbial* **61**:849-854
- Kuznetsov, V., Radyukina, N. and Shevyakova, N. 2006. Polyamines and Stress: Biology Role, Metabolism and Regulation. *Russ J. Plant Physiol.* **53**:583-604
- Lambrecht, M., Okon, Y., Vande B. A. and Vanderleyden, J. 2000. Indole-3-acetic-acid: A Reciprocal Signaling Molecule in Bacteria-plant Interaction. *Trends Microbiol.* **8**: 298-300
- Lamont, I. L. and Martin, L. W. 2003. Identification and Characterization of Novel Pyoverdine Synthesis Genes in *Pseudomonas aeruginosa*. *Microbiology* **149**:833-842

- Leisinger, T. and Margraff, R. 1979. Secondary Metabolites of Fluorescent Pseudomonads. *Microbial Rev.* **43**:422-442
- Lewis, T. A., Cortese, M. S., Sebat, J. L., Green, T. L., Lee, C. H. and Crawford, R. L. 2000. A *Pseudomonas stutzeri* Gene Cluster Encoding Biosynthesis of the CC<sub>14</sub>-Dechlorination Agent Pyridine-2,6-bis(Thiocarboxylic Acid). *Environ. Microbial.* **2**:407-416
- Lim, H. S., Kim, Y. S. and Kim, S. D. 1991. *Pseudomonas stutzeri* YPL-1 Genetic Transformation and Antifungal Mechanisms against *Fusarium solani*, an Agent of Plant Root Rot. *Appl. Environ. Microbial* **57**:510-516
- Liu, K., Fu, H., Bei, Q. and Luan, S. 2000. Inward Potassium Channel in Guard Cell as a Target for Polyamine Regulation of Stomatal Movements. *Plant Physiol* **124**:1315-1325
- Loper, J. E. 1988. Role of Fluorescent Siderophore Production in Biological Control of *Pythium ultimum* by a *Pseudomonas fluorescens* Strain. *Phytopathology* **78**:166-172
- Loria, R., Bukhalid, R. A., Fry, B. A. and King, R. R. 1997. Plant Pathogenicity in the Genus *Streptomyces*. *Plant Dis.* **81**:836-846
- Lorito, M., Woo, S. L., Ambrosio, M. D., Harman, G. E., Hayes, C. K., Kubicek, C. P. and Scala, F. 1996. Synergistic Interaction between Cell Wall Degrading Enzymes and Membrane Affecting Compounds. *Mol. Plant Microbe Interact* **9**:206-213
- Lugtenberg B. J. J., Kravchenko, L. V. and Simon, M. 1999. Tomato Seed and Root Exudate Sugars: Competition, Utilization by *Pseudomonas* Biocontrol Strains and Role in Rhizosphere Colonization. *Environ Microbial* **1**: 439-446
- Lugtenberg, B. J. J. and Dekkers, L. C. 1999. What Makes *Pseudomonas* Bacteria Rhizosphere Competent? *Environ Microbial* **1**:9-13
- Mansfield, J. W. 1983. Antimicrobial Compounds. In Callow, J. A. (Ed.). *Biochemical Plant Pathology*. UK: Wiley
- Marek-Kozaczuk, M., Derylo, M. and Skorupska, A. 1996. Tn5 Insertion Mutants of *Pseudomonas* Sp. 267 Defective in Siderophore Production and Their Effect on Clover (*Trifolium pretense*) Nodulated with *Rhizobium leguminosarum* var. *Trifolii*, *Plant soil* **179**:269-274
- Maurhofer, M., Hase, C., Meuwly, P., Mettraux, J. P. and Defago, G. 1994. Induction of Systemic Resistance of Tobacco Mosaic Virus by the Root Colonizing *Pseudomonas Fluorescens* Strain CHAO: Influence of the *Gaca* Gene of Pyoverdine Production. *Phytopathology* **84**:139-146
- Maurhofer, M., Keel, C., Schnider, U., Viosard, C., Haas, D. and Defago, G. 1992. Influence of Enhanced Antibiotic Production in *Pseudomonas Fluorescens* Strain CHAO on Its Disease Suppressive Capacity. *Phytopathology* **82**: 190-195
- Mazzola, M., Cook, R. J., Thomashow, L. S., Weller, D. M. and Pierson, L. S. 1992. Contribution of Phenazines Antibiotic Biosynthesis to the Ecological Competence of Fluorescent Pseudomonads in Soil Habitats. *Appl. Environ. Microbial* **58**:2616-2624

- McInroy, J. A. and Kloepper, J. W. 1995. Survey of Indigenous Bacterial Endophytes from Cotton and Sweet Corn. *Plant and soil* **173**: 337-342
- Mcloughlin, T. I., Quinn, J. P., Bettermann, A. and Bookland, R. 1992. *Pseudomonas cepacia* Suppression of Sunflower Wilt Fungus and Role of Antifungal Compounds in Controlling the Disease. *Appl. Environ. Microbiol.* **58**:1760-1763
- Mercado-Blanco, J., Van der Drift, K. M. G. M., Olsson, P., Thomas Oates, J. E., van Loon, L. C. and Bakker, P. A. H. M. 2001. Analysis of the PmsCEAB Gene Cluster Involved in Biosynthesis of Salicylic Acid and the Siderophore Pseudomonice in the Biocontrol Strain *Pseudomonas fluorescens* WCS374. *J. Bacteriol* **183**:1909-1920
- Milus, E. A. and Rothrock, C. S. 1997. Efficacy of Bacterial Seed Treatments for Controlling Pythium Root Rot of Winter Wheat. *Plant Dis.* **81**:180-184
- Mohd. Zainudin, N. A. I. and Faridah, A. 2008. Disease Suppression in *Ganoderma*-infected Oil Palm Seedlings Treated with *Trichoderma harzianum*. *Plant Protect. Sci* **44**:101-107
- Mossialos, D., Meyer, J. M., Budzikiewicz, H., Wolff, U., Koedam, N. and Baysee, C. 2000. Quinolobactin, a new Siderophore of *Pseudomonas Fluorescens* ATCC 17400 Whose Production is Repressed by the Cognate Pyoverdine. *Appl. Environ. Microbiol.* **64**:487-492
- Najmie, M. M. K., Khalid, K. and Jusoh, M. A. 2011. Density and Ultrasonic Characterization of Oil Palm Trunk Infected by *Ganoderma Boniniense* Disease. *Measurement Science Review* **11(5)**:160-164
- Nazeeb, M., Barakabah, S. S. and Loong, S. G. 2000. Potential of High Intensity Oil Palm Plantings in Diseased Environments. *The planter* **76**: 699-710
- Nielsen, M. N., Sorensen, J., Fels, J. and Pedersen, H. C. 1998. Secondary Metabolite and Endochitinase Dependent Antagonism toward Plant-Pathogenic Microfungi of *Pseudomonas Fluorescens* Isolates from Sugar Beet Rhizosphere. *Appl. Environ. Microbiol* **64**:3563-3569
- Niemi, K., Haggman, H. and Sarjala, T. 2001. Effects of Exogenous Diamines on the Interaction between Ectomycorrhizal Fungi and Adventitious Root Formation in Scots Pine *In Vitro*. *Tree Physiol* **22**:373-38
- Nimnoi, P., Pongsilp, N. and Lumyong, S. 2010. Endophytic Actinomycetes Isolated from *Aquilaria Crassna* Pierre Ex Lec and Screening of Plant Growth Promoters Production. *World J. Microbiol Biotechnol.* **26**:193-203
- Nowak, J. and Shulaev, V. 2003. Priming for Transplant Stress Resistance in *In Vitro* Propagation. *Biol.-Plant* **39**:107-124
- Nowak-Thompson, B., Gould, S. J. and Loper, J. E. 1997. Production of 2,4-Diacetylphloroglucinol by the Biocontrol Agent *Pseudomonas fluorescens* Pf 5. *Can. J. Microbiol* **40**:1064-1066



- O' Sullivan, D. J. and O' Gara, F. 1992. Traits of *Pseudomonas* spp. Involved in Suppression of Plant Root Pathogen. *Microbial Rev.* **56**:662-676
- Ogbebor, N. O., Adekunle, A. T., Eghafona, N. O. and Ogboghodo, A. I. 2010. *Ganoderma pseudoferreum*: Biological Control Possibilities with Microorganisms Isolated from Soils of Rubber Plantation in Nigeria. *African Journal of General Agriculture* **6**:4
- Okon, Y., Heytler, P. and Hardy, W. 1983. N<sub>2</sub> Fixation by *Azospirillum brasilense* and Its Incorporation into Host *Setaria italica*. *Appl. Environ. Microbiol* **46**:694-697
- Onofre-Lemus, J., Hernández-Lucas, I., Girard, L. and Caballero Mellado, J. 2009. ACC (1-Aminocyclopropane-1-Carboxylate) Deaminase Activity, a Widespread Trait in *Burkholderia* Species, and Its Growth-Promoting Effect on Tomato Plants. *Appl Environ Microbiol* **70**:6581-6590
- Palleroni, N. J. 1975. General Properties and Taxonomy of the Genus *Pseudomonas*. In Clarke, P. H., Richmond, M. H. (Eds.). *Genetics and Biochemistry of Pseudomonas*. USA: Wiley, Baltimore
- Palleroni, N. J. and Doudoroff, M. 1972. Some Properties and Taxonomic Subdivisions of the Genus *Pseudomonas*, *Annu. Rev. Phytopathol* **10**:73-100
- Palleroni, N., Kunisawa, R., Contopoulou, R. and Doudoroff, M. 1973. Nucleic Acid Homologies in the Genus *Pseudomonas*. *Int. J. Syst. Bacteriol* **23**:333-339
- Parmeela and Johri, B. N. 2004. Phylogenetic Analysis of Bacterial Endophtes Showing Antagonism against *Rhizoctonia solani*. *Current Science* **87**:687-692
- Patten, C. L and Glick, B. R. 2003. Role of *Pseudomonas putida* Indoleacetic Acid in Development of the Host Plant Root System. *Appl. Environ Microbiol* **68**:3795-3801
- Penrose, D. M. and Glick, B. R. 2003. Methods for Isolating and Characterizing ACC Deaminase Containing Plant Growth Promoting Rhizobacteria. *Physiol. Plant* **118**:10-15
- Perrig, D., Boiero, L., Masciarelli, O., Penna, C., Cassan, F. and Luna, V. 2007. Plant Growth Promoting Compounds Produced by Two Agronomically Important Strains of *Azospirillum brasilense* and Their Implications for Inoculant Formulation. *Appl. Microbial Biotechnol.* **75**:1143-1150
- Persello, C. F., Nussaume, L. and Robaglia, C. 2003. Tales from the Underground: Molecular Plant-Rhizobacteria Interactions. *J. Plant Cell Environ.* **26**: 189-199
- Pfender, W. F., Kraus, J. and Loper, J. E. 1993. A Genomic Region from *Pseudomonas fluorescens* Pf-5 Required for Pyrrolnitrin Production and Inhibition of *Pyrenophora tritici-repentis* In Wheat Straw. *Phytopathology* **83**:1223-1228
- Picard, C., Di Cello, F., Ventura, M., Fani, R. and Guckert, A. 2000. Frequency and Biodiversity of 2,4-Diacetylphoroglucinol-producing Bacteria Isolated from the Maize Rhizosphere at Different Stages of Plant Growth. *Appl. Environ. Microbiol* **66**: 948-955

- Pierson, L. S. and Thomashow, L. S. 1992. Cloning of Heterologous Expression of Phenazine Biosynthesis Locus from *Pseudomonas aureofaciens* 30-84. *Mol. Plant Microbe Interact* **53**:330-339
- Pieterse, C. M. J., Van Pelt, J. A., Van Wees, J., Ton, S. C. M., Leon-Kloosterziel, K. M., Keurentjies, J. J. B., Verhagen, B. W. M., van Knoester, M., Bakker, P. A. H. M. and Van Loon, L. C. 2001. Rhizobacteria-mediated Induced Systemic Resistance Triggering, Signaling and Expression. *Eur. J. Plant Pathol.* **107**:51-61
- Pillay, V. K. and Nowak, J. 1997. Inoculum Density, Temperature and Genotype Effects on *In Vitro* Growth Promotion and Epiphytic and Endophytic Colonization of Tomato (*Lycopersicon esculentum* L.) Seedling Inoculated with a Pseudomonad Bacterium. *Can. J. Microbiol* **43**: 354-361
- Pirttilä, A. M., Joensuu, P., Pospiech, H., Jalonen, J. and Hohtola, A. 2004. Bud Endophytes of Scots Pine Produce Adenine Derivatives and Other Compounds That Affect Morphology and Mitigate Browning of Callus Cultures. *Physiol. Plantarum* **121**:305-312
- Poppe, K., Taraz, K. and Budzikiewicz, H. 1987. Pyoverdine Type Siderophores from *Pseudomonas fluorescens*. *Tetrahedron* **43**:2261-2272
- Potgieter, H. and Alexander, M. 1966. Susceptibility and Resistance of Several Fungi to Microbial Lysis. *J. Bacteriol* **91**:3204-3208
- Quadt, H. A., Benhamou, N. and Kloepper, J. W. 1997. Bacterial Endophytes in Cotton: Mechanisms of Entering the Plant. *Canada Journal of Microbiol* **43**: 577-582
- Rahamath, B., Siti, N. F., Khairulmazmi and Idris, A. 2010. Control of *Ganoderma boninense*: A Causal Agent of Basal Stem Rot Disease in Oil Palm with Endophyte Bacteria *in vitro*. *International Journal of Agriculture & Biology* **12**: 833-839
- Ramamoorthy, V., Viswanathan, R., Raguchander, T., Prakasam, V. and Smaiyappan, R. 2001. Induction of Systemic Resistance by Plant Growth-promoting Rhizobacteria in Crop Plants against Pests and Disease. *Crop Prot.* **20**:1-11
- Rao, A. K. 1990. Basal Sem Rot (*Ganoderma*) in Oil Palm Smallholdings-IADP Johore Barat Experience. *Palm Oil Research Institute of Malaysia* 113-131
- Ravindra Naik, P. and Sakthivel, N. 2006. Functional Characterization of a Novel Hydrocarbonoclastic *Pseudomonas* Sp. Strain PUP6 with Plant-Growth-Promoting Traits and Antifungal Potential. *Res Microbiol* **157**: 538-546
- Ravindra Naik, P., Raman, G., Badri Narayanan, K. and Sakthivel, N. 2008. Assessment of Genetic and Functional Diversity of Phosphate Solubilizing Fluorescent Pseudomonads Isolated from Rhizospheric Soil. *BMC Microbiol* **8**:230
- Razak, A. J., Ahmad, H., Ramdhan, K., Idris, A. S., Rahim, A. S., Aminul, R. and Fauzi, I. 2004. Mechanical Trunk Injection for Control of *Ganoderma*. *MPOB information series* TT No.215
- Rees, R. W., Flood, J., Hasan, Y., Potter, U. and Cooper, R. M. (2009). Basal Stem Rot of Oil Palm (*Elaeis guineensis*); Mode of Root Infection and Lower Stem Invasion by *Ganoderma boninense*. *Plant Pathology* **58**:982-989

- Richardson, A. E., Hadobas, P. A., Hayes, J. E., O' Hara, J. E. and Simpson, R. J. 2001. Utilization of Phosphorus by Pasture Plants Supplied with Myo-Inositol Hexaphosphate is Enhanced by the Presence of Soil Microorganism. *Plant Soil* **229**:47-56
- Rosenblueth, M. and Martinez, R. E. 2004. Rhizobium Etli Maize Population and Their Competitiveness for Root Colonization. *Arch.Microbiol* **181**:337-344
- Rovira, A. D. 1965. Interactions between Plant Roots and Soil Microorganism. *Annu. Rev.Microbial*.**19**:241-266
- Ryu, C. M., Farag, M. A., Hu, C. H., Reddy, M. S., Wei, H. X., Pare, P. W. and Kloepper, J. W. 2003. Bacterial Volatiles Promote Growth in Arabidopsis. *Proc. Natl. Acad. Sci. U.S.A* **100**:4927-4932
- Sakthivel, N. and Gnanamanickam, S. S. 1987. Evaluation of *Pseudomonas fluorescens* for Suppression of Sheath-Rot Disease and for Enhancement of Grain Yields in Rice (*Oryza sativa*). *Appl. Environ. Microbiol.* **53**:2056-2059
- Salisbury, F. B. 1994. The Role of Plant Hormones Plant Environment Interactions. In Wilkinson, R.E. (Ed.). *Plant Environment Interaction*. New York: Dekker
- Sambrook, J., Fritsch, E. F. and Maniatis, T. 1989. *Molecular Cloning a Laboratory Manual, 2<sup>nd</sup>*. New York: Spring Harbor Laboratory Press.
- Sands, D. C. and Rovira, A. D. 1971. *Pseudomonas fluorescens* biotype G, the Dominant Fluorescent Pseudomonads in South Australian Soils and Wheat Rhizosphere. *J. Appl. Bacteriol* **34**:262-275
- Sariah, M., 2003. The Potential of Biological Management of Basal Stem Rot of Oil Palm Seedlings by Calcium Nitrate. *The Planter* **73**: 359–36
- Schena, L., Nigro, F., Pentimone, I. A. and Ligorio, A. I. 2003. Control of Postharvest Rots of Sweet Cherries and Table Grapes with Endophytic Isolates of *Aureobasidium pullulans*. *J. Postharv. Biol.Technol.* **30**: 209–220
- Scher, F. M. and Baker, R. 1982. Effects of *Pseudomonas putida* and a Synthetic Iron Chelator on Induction of Soil Suppressiveness to *Fusarium* Wilt Pathogen. *Phytopathology* **72**: 1567-1573
- Scherff, R. H. 1973. Control of Bacterial Blight of Soybean by *Bdellovibrio bacteriovorus*. *Phytopathology* **63**:400-402
- Schroth, M. N. and Hancock, J. G. 1981. Selected Topics in Biological Control. *Annu. Rev. Microbial* **35**:453-476
- Schroth, M. N., Hidebrand, D. C. and Panopoulos, N. 1992. Phytopathogenic Pseudomonads and Related Plant-Associated Pseudomonads. In Balows, A., Truper, H. G., Dworkin, M., Harder, W. and Schleifer, K. H. (Eds.). *The Prokaryotes : A Handbook on the Biology of Bacteria: Ecophysiology, Isolation, Identification, Applications*. New York: Springer
- Sesstisch, A., Howison, J. G., Perret, X., Antoun, H. and Martinez, R. E. 2002. Advances in Rhizobium Research. *Crit. Rev. Plant Sci.* **21**: 323-378

- Sessitsch, A., Coenye, T., Sturz, A. V., Vandamme, P., Ait Barka, E., Faure, D., Reiter, B., Glick, B. R., Wang-Prusk, G. and Nowak, J. 2005. *Burkholderia phytofirmans* sp. nov., a Novel Plant-associated Bacterium with Plant Beneficial Properties. *Int J Syst Evol Microbiol* **55**:1187–1192
- Sevilla, M., Burris, R. H., Gunapala, N. and Kennedy, C. 2001. Comparison of Benefit to Sugarcane Plant Growth and  $^{15}\text{N}_2$  Incorporation Following Inoculation of Sterile Plants with *Acetobacter diazotrophicus* Wild Type and Nif-mutants Strains. *Plant-Microbe Interact* **14**: 358-366
- Shah, S., Li, J., Moffatt, B. A. and Glick, B. R. 1998. Isolation and Characterization of ACC Deaminase Genes from Two Different Plant Growth-Promoting Rhizobacteria. *Can. J. Microbiol* **44**:833-843
- Shapira, R., Oedentch, A., Chet, I. and Eppenheim, A. B. 1989. Control of Plant Disease by Chitinase Expressed from Cloned DNA in *Escherichia coli*. *Phytopathology* **79**:1246:1249
- Shenker, M., Ghirlando, R., Oliver, I., Helmann, M., Hadar, Y. and Chen. Y. 1995. Chemical Structure and Biological Activity of Rhizoferrin-A Siderophore Produced by *Rhizopus Arrhizus*. *Soil Sci. Soc. Am. J.* **59**:837-843
- Shukla, A. N. and Uniyal, K. 1989. Antagonistic Interactions of *Ganoderma lucidium* (Leyss.) Karst. against Some Soil Microorganisms. *Curr. Sci.* **58**: 265–267
- Siddiqui, Z. A. 2006. PGPR: Prospective Biocontrol Agents of Plant Pathogens. In Siddiqui, Z. A. (Ed.). *PGPR: Biocontrol and Biofertilization*. Netherlands: Springer
- Singh, G. 1990. *Ganoderma*-The Scourge of Oil Palm in the Coastal Areas. *Palm Oil Research Institute of Malaysia* 7-35
- Smilanick, J.L., Denis-Arrue, R., Bosch, I. R., Gonzalez, A. R., Henson, D. and Janisiewicz, W. J. 1993. Control of Postharvest Brown Rot of Nectarines and Peaches by *Pseudomonas* species. *Crop Prot.* **12**:513-520
- Sneath, P. H. A., Stevens, M. and Sackin, M. J. 1981. Numerical Taxonomy of *Pseudomonas* Based on Published Records of Substrate Utilization. *Antonie Van Leeuwenhoek* **47**:423-448
- Soepena, H., Purba, R. Y. and Pawirosukarto, S. 2000. A Control Strategy for Basal Stem Rot (*Ganoderma*) on Oil Palm. *CAB International* 83-88
- Stanier, R. Y., Palleroni, N. J. and Doudoroff, M. 1966. The Aerobic Pseudomonads: A Taxonomic Study. *J. Gen. Microbiol* **43**:159-217
- Stephens, P. M., O' Sullivan, M and O' Gara, F. 1987. Influence of Bacteriophages on the Colonization of Strains of *Pseudomonas Fluorescens* in the Rhizosphere of Sugarbeet. *Appl. Environ. Microbiol.* **53**:1164-1167
- Stevens, A. M., Dolan, K. M. and Greenberg, E. P. 1994. Synergistic Binding of the *Vibrio Fischeri Luxr* Transcriptional Activator Domain and RNA Polymerase to Lux Promoter Region. *Proc. Natl. Acad. Sci. USA* **91**:12619-12623
- Stewart, V. 1988. Nitrate Respiration in Relation to Facultative Metabolism in Enterobacteria. *Microbiol Rev.* **52**:190-232

- Streit, W. R., Joseph, C. M. and Phillips, D. A. 1996. Biotin and Other Water-Soluble Vitamins are Key Growth Factors of Alfalfa Root Colonization By *Rhizobium meliloti* 1021. *Mol. Plant Microbe Interact* **5**:330-338
- Sturz, A. V., Chirstie, B. R. and Nowak, J. 2000. Bacterial Endophytes: Potential Role in Developing Sustainable Systems of Crop Production. *Crit. Rev. Plant Sci.* **19**:1-30
- Sunish Kumar, R., Ayyadurai, N., Pandiaraja, P., Reddy, A. V., Venkateswarlu, Y., Prakash, O. and Sakthivel, N. 2005. Characterization of Antifungal Metabolite Produced by a New Strain *Pseudomonas aeruginosa* Pupa3 that Exhibits Broad-Spectrum Antifungal Activity and Biofertilizing Traits. *J. Appl. Microbiol* **98**:145-154
- Susanto A., Sudharto P. S., Purba R. Y. 2005. Enhancing Biological Control of Basal Stem Rot Disease (*Ganoderma boninense*) in Oil Palm Plantations. *Mycopathologia* **159**: 153-157
- Tan, Z., Hurek, T. and Reinhold, H. B. 2003. Effect of N-fertilization, Plant Genotype and Environmental Condition on nifH Gene Pools in Roots of Rice. *Environ. Microbiol* **5**: 1009-1015
- Teoh, C. H. 2002. The Palm Oil Industry in Malaysia: From Seed to Frying Pan. Plantation Agriculture, WWF Malaysia
- Thomashow, L. S., Weller, D. M., Bonsall, R. F. and Pierson, L. S. 1990. Production of the Antibiotic Phenazine 1-Carboxylic Acid by Fluorescent *Pseudomonas* Species in the Rhizosphere of Wheat. *Appl. Environ. Microbiol* **56**:908-912
- Thuler, D., Floh, E., Handro, W. and Barbosa, H. 2003. Plant Growth Regulators and Amino Acids Released by *Azospirillum* Sp. *Lett. Appl. Microbiol.* **37**:174-178
- Tiwari, R. P., Hoondal, G. S. and Tewari, R. 2009. *Laboratory Techniques in Microbiology and Biotechnology*. Departments of Microbiology and Biotechnology. New Delhi: Bharat Shushan Abhlshek Publications
- Tombolini, R., vander Gaag, D. J., Gerhardson, B. and Jansson, J. K. 1999. Colonization Pattern of the Biocontrol Strain *Pseudomonas chlororaphis* MA342 on Barley Seeds Visualized by Using Green Fluorescent Protein. *Appl. Environ. Microbiol.* **65**:3674-3680
- Turner, J. M. and Messenger, A. J. 1986. Occurrence, Biochemistry and Physiology of Phenazine Pigment Production. *Adv. Microb. Physiol.* **27**:211-275
- Van Etten, H. D. and Kistler, H. C. 1984. Microbial Enzyme Regulation and Its Importance for Pathogenicity. In Kosuge, T and Nester, E. W. (Eds.). *Plant-Microbe Interactios*. New York: Macmillan
- Van Loon, L. C., Bakker, P. A. H. M. and Pieterse, C. M. J. 1998. Systemic Resistance Induced by Rhizosphere Bacteria. *Annu. Rev. Phytopathol.* **36**:453-483
- Van Peer, R. and Schipper, B. 1988. Plant Growth Response to Bacterization with Selected *Pseudomonas* spp. Strains and Rhizosphere. *Can. J. Microbiol* **35**:456-463

- Van Peer, R., Niemann, G. J. and Schippers, B. 1991. Induced Resistance and Phytoalexin Accumulation in Biological Control of *Fusarium* Wilt of Carnation by *Pseudomonas* sp. Strain. *Phytopathology* **81**:728-732
- Verma, S. C., Singh, A., Chowdhury, S. P. and Tripathi, A. K. 2004. Endophytic Colonization Ability of Two Deep-water Rice Endophytes, *Pantoea* sp. and *Ochrobactrum* sp. Using Green Fluorescent Protein Reporter. *Bio-technol. Lett.* **26**:425-429
- Voisard, C., Keel, C., Haas, D. and Defago, G. 1981. Cyanide Production in *Pseudomonas fluorescens* Helps Suppress Black Root Rot of Tobacco under Gnotobiotic Conditions. *EMBO J.* **8**:351-358
- Weller, D. M. 1988. Biological Control of Soilborne Plant Pathogens in the Rhizosphere with Bacteria. *Annu. Rev. Phytopathol.* **26**:379-407
- Wijesekera, H. T. R., Wijesundera, R. L. C. and Rajapakse, C. N. K. 1996. Hyphal Interactions between *Trichoderma viridae* and *Ganoderma boninense* Pat. Cause of Coconut Root and Bole Rot. *Sri Lanka J. Nat. Sci.* **24**: 217-219
- Zahnder, G. W., Murphy, J. F., Sikora, E. J. and Kloepper, J. W. 2001. Application of Rhizobacteria for Induced Resistance. *Eur. J. Plant Pathol.* **107**:39-50
- Zaiton binti Sapak. 2006. Bacterial Endophytis from Oil Palm (*Elaeis Guineensis*) and Their Anganonic Activity against *Ganoderma Boninense*. Degree of Master of Science. Universiti Putra Malaysia
- Zaiton, S., Sariah, M. and Zainal, A.M.A. 2008. Effect of Endophytic Bacteria on Growth and Suppression of *Ganoderma* Infection in Oil Palm. *International Journal of Agriculture and Biology.***10**:127-132