EFFECT OF STORAGE TEMPERATURE AND HANDLING OF SABA BANANA BUNCHES AND HANDS TO ITS POSTHARVEST QUALITY

TING WEI SOON

DISSERTATION SUBMITTED IN PARTIAL FULFILMENT OF THE REQUIREMENTS FOR THE DEGREE OF BACHELOR OF AGRICULTURAL SCIENCE WITH HONOURS

HORTICULTURE AND LANDSCAPING PROGRAMME FACULTY OF SUSTAINABLE AGRICULTURE UNIVERSITI MALAYSIA SABAH 2017



UNIVERSITI MALAYSIA SABAH

	BORANG PENGESAHAN TES	IS
JUDUL: EFFECT	OF STORAGE TEMPERATURE AND HAW S AND HANDS TO ITS POSTHARI	DLING DE SAGA BANANA 1EST QUALITT
UAZAH:	OF BACHELOR OF AGRICULTURE S TICULTURE AND LAUDSCAPING)	CIENCE WITH HONOURS
	WEL SUDN SESI PENGAJIAN	= 2013-2017
Mengaku membenarka Sabah dengan syarat-sy	an tesis *(LPSM/Sarjana/Doktor Falsafah) ini dis yarat kegunaan seperti berikut:- ık milik Universiti Malaysia Sabah.	simpan di Perpustakaan Universiti Malaysia
•	Universiti Malaysia Sabah dibenarkan membuat s dibenarkan membuat salinan tesis ini sebagai b /)	ahan pertukaran antara institusi pengajian
SULIT	(Mengandungi maklumat yang berdarj seperti yang termaktub di AKTA RAHSI/	ah keselamatan atau kepentingan Malaysia A RASMI 1972)
TERHAI	D (Mengandungi maklumat TERHAD yang mana penyelidikan dijalankan)	g telah ditentukan oleh organisasi/badan di
TIDAK 1	TERHAD	Disahkan oleh: NURULAIN BINTI ISMAIL
(TANDATANGAI		(TANDATANGAN PUSTAKAWAN)
Alamat Tetap: 105-A KAMYUNG TIONG, KUALA TERENGGANU TERENGGANU	20(00)	
TARIKH:]3	1.1.2017	SCHOOL OF SUSTAINABLE AGRICULTION
menyatakan sekal *Tesis dimaksudka	ik berkenaan. T dan TERHAD, sila lampirkan surat daripada pihak b Ii sebab dan tempoh tesis ini perlu dikelaskan sebagai an sebagai tesis bagi Ijazah Doktor Falsafah dan Sarja cara kerja kursus dan Laporan Projek Sarjana Muda (L	i SULIT dan TERHAD. na Secara Penyelidikan atau disertai
		UN

DECLARATION

I hereby declare that this dissertation is based on my original work except for citations and quotations which have been duly acknowledged. I also declare that no part of this dissertation has been previously or concurrently submitted for a degree at this or any other university.

TING WEI SOON

BR13110188 29/11/2016



1. Dr. Jupikely James Silip SUPERVISOR

DR. JUPIKELY JAMES SILIP SENIOR LECTURER AACADEMIC ADVISOR SCHOOL OF SUSTAINABLE AGRICULTURE UNIVERSITI MALAYSIA SABAH



ACKNOWLEDGEMENT

I would like to express my gratitude and appreciation to my supervisor, Dr Jupikely James Silip for the guidance and advices along my work. I am greatly indebted for his persistant effort in guiding me and supervised me in doing my research efficiently and productively. All that I learnt from through Dr will be greatly beneficial for my future career.

I would also like to acknowledge with much appreciation the crucial role of the staff of laboratory, who gave permission to use all required materials and equipment to complete the data analysis of my research. Next, I would like to thanks Mdm. Nurul Syakina Binti Marli for aiding my laboratory work.

My gratefulness and thanks is directed to all the lectures, friends and staffs of FPL for their kindly help and support during my experimental work. I would like to extend special thanks to Teo Wei Ting and Chan Su Yi for their gracious help during the course of the research.

My sincere gratitude goes to my family for standing by me regardless of what happened, supporting me, believing in me and encouraging me throughout these years.



ABSTRACT

A study on effect of storage temperature and Saba banana bunches and hands handling to its postharvest quality was carried out at Faculty of Sustainable Agriculture, Universiti Malaysia Sabah, which was to compare different storage temperature such as cold room temperature (10±2°C), room temperature (24±3°C) and outside temperature (28±5°C). Every storage temperature had three whole bunches, three half top bunches, three half bottom bunches and three hands of bananas to be tested. After 2 weeks, bananas were withdrawn and tested for data analysis. Completely randomized design with three replicates for analysis was used in this study. Among the physico-chemical parameters such as total weight loss, peel and pulp colour, firmness, pulp to peel ratio, total soluble solid concentration, pH value, titratable acidity were tested in this study. There was no significant interaction between samples handling and storage temperature observed in the study as well as no significant difference on sample handling towards the postharvest quality of Saba banana. Cold room temperature recorded minimum (9.04%) weight loss in this 2 weeks storage, whereas outside temperature exhibited maximum (21.07%) weight loss. The highest firmness (3.36kg/f) was recorded in cold room temperature, whereas outside temperature had lowest firmness (1.12kg/f). For the pulp to peel ratio and total soluble solid concentration, room temperature recorded maximum value (2.33 and 23.2% Brix) respectively. Outside temperature recorded lowest pulp to peel ratio (0.87) and cold room temperature exhibited minimum soluble solid concentration (5.90% Brix). Lowest titratable acidity was shown by cold room temperature (0.14%) and room temperature recorded maximum (0.34%) titratable acidity. The best temperature was cold room storage temperature which resulted in the optimum postharvest quality of banana.



KESAN SUHU SIMPANAN DAN PENGENDALIAN TANDAN DAN TANGAN TERHADAP KUALITI LEPAS TUAI PISANG SABA

ABSTRAK

Satu kajian berkenaan kesan suhu simpanan dan pengendalian tandan dan tangan pisang Saba terhadap kualiti lepas tuai telah dijalankan di Fakulti Pertanian Lestari, Universiti Malaysia Sabah, untuk membandingkan suhu simpanan berlainan seperti dalam bilik sejuk (10±2°C), suhu bilik (24±3°C) dan suhu luar (28±5°C). Setiap suhu simpanan akan ada tiga tandan, tiga tandan atas, tiga tandan bawah dan tiga tangan pisang untuk dikaji. Selepas dua minggu, pisang dikeluarkan dan dikaji untuk mengumpulkan data. Rekabentuk Rawak Lengkap dengan tiga replikasi bagi agronomi analisis telah digunakan untuk kajian ini. Antara parameter fiziko-kimia seperti pengurangan berat, warna kulit dan daging, ketegangan, nisbah antara kulit dan daging, jumlah kepekatan pepejal larut, nilai pH, tertitrat keasidan telah diuji dalam kajian ini. Tidak ada interaksi antara sampel pengendalian dan suhu penyimpanan diperhatikan dalam kajian ini dan juga tiada perbezaan yang signifikan dalam pengendalian lepas tuai terhadap kualiti pisang Saba Suhu bilik sejuk mencatatkan minimum (9.04%) pengurangan berat selama penyimpanan 2 minggu, manakala suhu luar dipamerkan maksimum (21.07%) pengurangan berat. Ketegangan tertinggi (3.36kg/f) dicatatkan pada suhu bilik sejuk, manakala suhu luar mempunyai ketegangan paling rendah (1.12kg/f). Untuk nisbah antara daging dan kulit, dan jumlah kepekatan pepejal larut, suhu bilik mencatatkan nilai maksimum (2.33 dan 23.2% Brix) masing-masing. Suhu luar merekodkan nisbah minimum (0.87) dan suhu bilik sejuk mempamerkan kepekatan pepejal larut minimum (5.90% Brix). Keasidan tertitrat terendah telah ditunjukkan oleh suhu bilik sejuk (0.14%) dan suhu bilik merekodkan maksimum (0.34%) keasidan. Suhu terbaik untuk lepai tuai pisang yang paling optimum adalah suhu sejuk simpanan.



TABLE OF CONTENT

Content	Page
Content DECLARATION	i
VERIFICATION	ii
ACKNOWLEDGEMENT	iii
ABSTRACT	iv
ABSTRAK	v
TABLE OF CONTENT	vi
LIST OF TABLES	viii
ITST OF FIGURES	
LIST OF FIGURES LIST OF SYMBOLS, UNITS AND ABBREVIATIONS	
LIST OF FORMULAE	xi
CHAPTER 1 INTRODUCTION	
1.1 Background	1
1.2 Justification	2 3 3
1.3 Objective	3
1.4 Hypotheses	3
CHAPTER 2 LITERATURE REVIEW	_
2.1 Musa	4
2.2 Botanical Description of Banana	5
2.3 Different Stages of Maturation	7
2.4 Post-harvest Physiology	8
2 4 1 Diaments	8 9
2.4.2 Ethylene Production and Respiration	9 10
2 4 3 PhenoliC	10
2.4.4 Sweetness and Starch Content	10
2.5 Factors Affecting Postharvest Loss of Banana	11
2.5.1 Pre-harvest Factors	11
2.5.2 General Postharvest Factors	12
2.6 Factors Affecting Postharvest Quality 2.6.1 Temperature and Relative Humidity	12
2.6.2 Controlled Atmosphere (CA)	13
2.6.3 Modified Atmosphere (MA)	13
CHAPTER 3 METHODOLOGY	
3.1 Place, material and method	14
	14
3.2.1 Determination of Weight loss and Pulp to Peel Tatio	14
3.2.2 Determination of Peel and Pulp colour	15
3.2.3 Determination of Banana Pulp Firmness	15
3.2.4 Soluble Solid Concentration	16
3.2.5 pH value	16
3.2.6 Titratable Acidity (TA)	16
3.3 Statistical Analysis	17



CHAPTER 4 RESULTS	20
4.1 Colour Index of Banana Peel and Pulp	
4.2 Firmness of Banana	
4.3 Pulp to Peel ratio	21 22
4.4 Total soluble solid concentration (SSC)	22
4.5 pH value	23
4.6 Titratable Acidity	25
CHAPTER 5 DISCUSSION	
5.1 Colour Index of Banana Peel and Pulp	25
5.2 Firmness of Banana	26
5.3 Pulp to Peel ratio	26
5.4 Total soluble solid concentration (SSC)	26
5.5 pH value	27
5.6 Titratable Acidity	27
CHAPTER 6 CONCLUSION	28
6.1 Conclusion	28
6.2 Recommendations	
	29
REFERENCES APPENDICES	34



LIST OF TABLES

TABLE		PAGE
4.1	The average mean for each parameters under different storage temperature and samples handling	19



LIST OF FIGURES

FIGURE		PAGE
2.1	Banana stool-Reproduction stage	5
2.2	Hands of banana	6
3.1	Banana Peel Colour Index	15
4.1	Effect of storage temperature on the firmness of Saba banana after 2 weeks of harvesting.	20
4.2	Effect of storage temperature on the pulp to peel ratio of Saba banana after 2 weeks of harvesting.	21
4.3	Effect of storage temperature on total soluble solid concentration of Saba banana after 2 weeks of harvesting.	22
4.4	Effect of storage temperature on the pH value of Saba banana after 2 weeks of harvesting.	23
4.5	Effect of storage temperature on titratable acidity of Saba banana after 2 weeks of harvesting	24



LIST OF SYMBOLS, UNITS AND ABBREVIATIONS



LIST OF FORMULAE

Formula		Page
3.1	Weight Loss = $\frac{Initial \ weight - Final \ weight}{Initial \ weight} X \ 100\%$	14
3.2	Pulp to Peel ratio = $\frac{Pulp \ weight}{Peel \ weight} X \ 100\%$	15
3.3	%TA = <u>Titre (m) x N(NaOH) x <i>acid meg.factor</i> X 100ml</u> Sample volume (mL) x Sample weight (g)	17



CHAPTER 1

INTRODUCTION

1.0 Background

Banana (*Musa spp*) is the most important tropical fruit in worldwide. In Malaysia, it is ranked second in term of production with 530,000 metric tons of total production (FAO, 2003). Among all the variety of banana cultivars grown, Saba (Musa balbisiana) are gaining popularity in both domestic and international market. Saba is known as a cooking banana which is cooked before been eaten. There are more variety of cultivars under cooking banana such as Pisang Nangka, Pisang Raja and Pisang Relong. Saba banana is usually processed into everyone's favourite banana chips where 95% of the country's earning is derived from this product. (Molina and Roa, 2000).

Most of the part of saba banana can be used economically which is very cost efficient. The fruit itself can be eaten fresh when ripe but is usually eaten when cooked. Different kind of food products are derived from it such as chips, banana flour and flakes. Inflorescene of banana is consumed as vegetables. Wrapping materials used the leaves of banana to wrap the food products. Finely chopped pseudostem is cooked and used as feed for livestocks mainly hog, cattle and poultry.

Bananas and plantains will be harvested while still in mature green stage especially for a commercial purpose because they need to be ripened in controlled conditions when transported to destination markets. The ideal harvesting time for banana fruit is when the fruits still have sufficient green life to maximise the yield and sustain long enough for marketing purpose. Plantains however, have faster maturity time than bananas if harvested at the same time.



There are several criterions to be factored in harvesting banana fruit. One of them is the age of bunch after emergence from pseudostem. When bananas are growing rapidly during harvesting time, fruit size and finger richness are important measures of harvest maturity. Hands maturity index vary in a stem where hands at the proximal end of thr stem are more mature than those at the distal end.

Banana plant is known as a large perennial herb with around twelve leaves that are up to 9 feet long. The inflorescence (flower stalk) grows through pseudostem. After 60-90 days, the fruits reached their maturity when flowers first appear. The banana fruits are made up of bunch, hands and fingers where each bunch consists of few numbers of hands along and each hand consists of two rows of fruits which called as fingers.

1.1 Justification

Postharvest handling is a major problem in the fruit industry in Malaysia. Proper handling of banana at the post-harvest stage is not given proper attention. Rough handling often resulted in damage and losses to the fruits when the products reach the market. All these losses of fresh produce had become a major challenge in the postharvest sector (Atanda, 2011). Growers tend not to understand the effect of their harvesting and handling on the quality of produce when it reaches to the market. This would result in disagreement between farmers and traders where farmers have sent to market what they consider to be good quality products but traders see the produce had deteriorated badly by the time these arrive at the market (FAO, 2003).

Banana is a climacteric fruit with poor storage characteristics because it has high respiratory rate and ethylene production after harvest. This make the banana highly perishable and prone to postharvest losses (Turner, 2001). Most of the problems are mostly inclined towards harvesting immature fruits but there are also cases where bunches remained longer after maturity due to labour shortages. In both cases, postharvest losses will be inflicted to the farmer and also traders where banana suffer difficulties for even ripening (Adam, 2015).

Some factors lead to the loss of postharvest in Malaysia are inappropriate production practices where the farmers do not have sufficient knowledge. Poor agribusiness management like improper practices from farm to market would affect the banana quality. To minimise these losses, there must be a proper guideline on the



UNIVERSITI MALAYSIA SABAH

2

handling of banana during post-harvest stage (Hailu, 2013). A low quality produce from poor postharvest handling will result in reducing price and reputation of the production area. Improved harvesting and handling of produce will give better appearance and shelf life of the product.

1.2 Objective

The objective of this research is to study the effect of storage temperature and handling of Saba banana bunches and hands to its postharvest quality.

1.3 Hypotheses

Ho₁: There is no significant difference of the effect of storage temperatures on the postharvest quality of Saba banana

H_{A1}: There is significant difference of the effect of storage temperature on the postharvest quality of Saba banana

Ho₂: There is no significant difference of the effect of handling of Saba banana bunches and hands to its postharvest quality

H_{A2}: There is significant difference of the effect of handling of Saba banana bunches and hands to its postharvest quality

Ho₃: There is no significant interaction between storage temperatures and handling of Saba banana bunches and hands to its postharvest quality

H_A: There is significant interaction between storage temperature and handling of Saba banana bunches and hands to its postharvest quality



CHAPTER 2

LITERATURE REVIEW

2.1 Musa

The classification of banana is consisted of Musaceae family, genus Musa and species or hybrids in this genus. The name Musa is derived from Sanskrit. There are two type of bananas that are native from Southeast Asia. They are identified as Musa acuminate and Musa balbisiana (Robinson, 1996; Stover & Simmonds, 1987). Among the species, hybrids of diploid, triploid and tetraploid are found in Musa acuminata and between Musa acuminata and Musa balbisiana (Zhang *et al.*, 2005). For example, Musa cavendish is pure triploid acuminate (AAA group) and is a type of dessert banana but both Musa paradisiaca and Musa sapientum belong to AAB group (Stover & Simmonds, 1987b) and are characterized by higher starch concentration.

Banana is one of the most widely cultivated fruit in Malaysia. The whole banana plantation covered about 26,000 hectares with a total production of 530,000 metric tonnes (Chai *et al.*, 2014). About 50% of the bananas growing land is cultivated with Berangan banana and the popular cultivation of Cavendish types like Lady Finger banana, Silk banana, Raja banana, Awak banana and Saba banana. There is an excess of production and large quantities of fruits are lost during commerce, as a consequence of deficient postharvest handling. (Ovando-Martinez *et al.*, 2009). New economic strategies are considered for banana as a food ingredient (Maribel *et al.*, 2009).

Bananas are energy-rich in the form of starch and sugar (Molina *et al.*, 2002) and also contain high amounts of essential minerals, such as potassium, and various vitamins such as A, B, C, B1 and B2 (Noor Aziah and Choo, 2010). Banana are famous for its aroma and easy to peel and eat, besides rich in calcium as well as low in sodium content (Wall, 2006). A report by Sharrock and Lustry (2000) indicated that digestion





time of banana is less than apple, which is 105 min and 210min, respectively. These characteristics of banana allow it to be used in special diets where low fat, minerals, ease of digestion and vitamins content are required (Nakasone and Paul, 1998, Aurore *et al.*, 2009).

2.2 Botanical description of banana

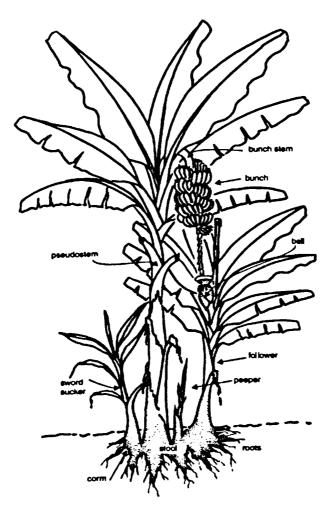


Figure 2.1 Banana stools- Reproduction Stage Source: (Agrilink, 1998)



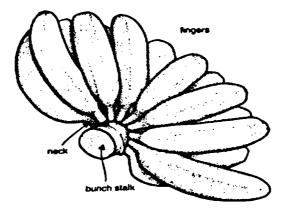


Figure 2.2 Hand of Bananas Source: (Agrilink, 1998)

The banana plant is believed can last up to 50 years due to its characteristics which are monocotyledonous, herbaceous and evergreen. Rhizome of the plant is responsible for bearing developing suckers and inflorescence that bear flowers which eventually bear the fruit. The plant itself also has an adventitious root system. According to figure 2.1, the banana flowers can be found on nodal structures which are arranged accordingly by a stout peduncle. At the nodal structures, each node comprised of two rows of flowers. Female flowers are on the basal node that develop into fruit and sometimes will grow in numbers between 5 to 16 nodes per stalk. In comparison, the male flowers can be found on the distal part. The flowers remain tightly closed on the node and these form the bell. Banana fruits will be developed from hermaphrodite flowers which formed by several nodes between the male and female nodes.

In order to increase the fruit size, the bell is cut when the distance between the last hand and the top bell is about 15 cm. This is to prevent the growth of meristem and direct the plant photosynthetic energy. (National Agriculture Research Institute Guyana, 2003). After harvesting the fruits, the aerial parts of the plant die and usually chopped down or naturally fallen down to the ground. New suckers will grow up from the base of the mother plant to replace the aerial parts that have died. The suckers eventually grow into a mature plant with a height of 3 metres or more depending on variety and conditions (Karamura *et al*, 1995).

Water is the most abundant element in the banana fruit especially within the pulp and peel of banana. A plantain fruit has lower water content compared to the pulp of a dessert banana. A fully ripe banana has 75% water of its pulp mass while the



UNIVERSITI MAL

plantain has 66% of its pulp water mass (Robinson *et al*, 1984). The water content increases at ripening but started to decrease from the peel externally due to transpiration. Degradation of the peel from continuous ripening process further reduces the water loss. The two main components for yield of bananas are fruit mass and cycle time. Fruit mass included hands per bunch (Figure 2.2), fingers per bunch, finger length and caliper. Cycle time refer to the interval between harvest time. Improving yield, therefore involves either an increase in fruit mass or a reduction in cycle time. These two components, as well as fruit quality are affected by the environment, cultural practises, biological and post-harvest factor

2.3 Different stages of maturation

Chlorophyll is produced within the peel of banana when banana starts to develop as a result of direct sunlight that required to grow the fruit. The green peel as the interior fruit absorb all the sunlight and begins to undergo a chemical change that helps in reaching the maturity of the fruit. During the process, the sweetness of the fruit is enhanced with the release of nutrients of the interior of the peel. After achieving maturity, the peel begins to lose chlorophyll content and changes from green to yellow. The peel will continue to deteriorate, leaving only a thin covering that can be peeled away manually from fruit with ease during this transformation. Since banana is a climacteric fruit and ripen quickly, it does not take a long time to go from a bitter green banana to a sweet yellow banana. According to the desired use, banana and plantain are consumed or locally processed at various stages of maturation which have been classified from stage 1, green skin, to stage 7, yellow skin with black spots (Emaga *et al.*, 2010).

Green banana is rich in starch and greater part of the starch found is the resistant starch type 2 (Faisant *et al.*, 1995). Around 70g/100g of starch (dry basis) is contained in its flour and around 12g/100 (dry basis) of fibre content in it. (Juarez-Garcia *et al.*, 2006). At different maturity stages, banana fruit exhibits significant difference (p < 0.01) on physio-chemical characteristics and fruit firmness. The soluble solids concentration increase from early stages until the end of maturity, while fruit firmness become softer during ripening, due to the action of enzymes involved in pectin degradation in the cell wall and middle lamella



Banana especially plantain is very abundant in starch content and becoming a primary replacement for the starch industry. (Chong and Noor Aziah, 2009). Ethylene acted as a catalyst for climacteric change and ripening process for consumption. The quality and appearance of banana are determined when consumed fresh with yellow-coloured peel, flavors, consistency of pulp and starch-sugar transition. The classification of ripening defined by various stages of colour index where stages 1 to 3 are not ideally to be consumed because the fruit is still green, hard and rich in starch. At stage 7 however, banana is overripe and muddy. (Aurore *et al.*, 2009).

2.4 Post-harvest Physiology

2.4.1 Pigments

Visual maturity index often look for colour changes in fruits during ripening process. The colour is usually determined by various pigments present in the peel and flesh (Rood, 1957). When fruits start to reach maturity, the fruit colour changed from green to yellow, red or other colours due to the presence of accessory pigments. (Romani and Jennings, 1971). Colour changes are either dependent or independent of ethylene action based on the types of pigments involved and the species of fruit. (Lelievre *et al.*, 1997). Chlorophyll 'a' and 'b' are the most dominate pigments in fruits and vegetables where these pigments can be found in all photosynthetic and storage tissues.

Chlorophyll degradation is mainly responsible for the colour change in fruits from green to yellow, which later exposed the presence of yellow carotenoids pigments. (Seymour *et al.*, 1993). Carotenoids are C₄₀ iso-propanoid compounds that contribute in various physiological processes in plants and other organisms (Fraser *et al.*, 2004). These carotenoids compounds are important in photosynthesis where they act as energy carriers and photo-oxidation protectors (Van Berg *et al.*, 2000). Some other functions of these compounds are as pollinator which mediated by insects in flowers and serve as indicators of maturity in fruits that make fruits attractive for human consumption (Arias *et al.*, 2000).

Carotenoids are free radical scavengers and act as a natural antioxidant. One of the most significant values of carotenoids is its essential role in human nutrition and prevention of diseases (Ye *et al.*, 2000). Since we cannot synthesize carotenoids, thus





we only consume fruits and vegetables to meet our nutritional requirements. The most studied carotenoids are β -carotene and lycopene. There are also some other carotenoids which received considerable attention like lutein and zeaxanthin due to their antioxidant properties (Volker *et al.*, 2002).

2.4.2 Ethylene Production and Respiration

The most important biological process is respiration accompanied by the release of ethylene in climacteric fruit like banana. As a growth regulating hormone, ethylene plays a significant role in the shelf life of banana by affecting the ripening rate (Pech *et al.*, 2008). This result into flesh softening, the development of colour and production of aroma compounds that attributed to the evolution of ethylene. Ethylene biosynthesis takes place through enzyme mediated metabolic pathway by using sulfur containing amino acid methionine as the biological precursor (Argueso *et al.*, 2007).

Excessive softening and firmness loss is correlated to ethylene emission in banana (Botondi *et al.*, 2003). The mechanism behind the ethylene emission is the conversion of structural carbohydrates into simple sugars. This conversion usually functions as an energy source for continued respiration. Respiration and evolution of ethylene speed up the ripening process, leading to accelerated membrane deterioration, nutrient depletion and excessive water losses. During ripening, respiration considerably increases over a short period (Eskin, 1991). If the storage atmosphere is not properly managed, rapid ripening leads to internal tissue breakdown with the production of volatile characteristics of over ripe fruit (Mathooko, 1995).

The respiration process involves oxygen and carbohydrates to make intermediate products and eventually carbon dioxide and water. This mechanism accompanied by enzymatic activities cause excessive textural softening leading to adverse effects on the nutritional and sensory quality of produce (Prasanna *et al.*, 2007). Certain enzymes like catalase and superoxide dismutase play a protective role against the destructive mechanism evolved during ripening which scavenging free radicals harmful to the tissues. Reactive oxygen species were involved in senescence and aging of fresh commodities (Yang and Ying, 2008). These enzymes avert the harmful effect of active oxygen species and hydroxyl groups by converting them into the water. The pace of action of different enzymes vary with the species and variety of fruits (Vicente, 2007).





2.4.3 Phenolic

Polyphenol content is highest during fruit growth and under normal ripening condition. The content started to decrease with the fruit ripening process. (Lakshminarayana *et al.*, 1970). These observations indicate a correlation between fruit ripening and loss of secondary metabolic substrates. Polyphenolic compounds in fruit and fruit products were linked to prevention of degenerative diseases due to their antioxidant activity. Fruit phenolic phytochemicals have the potential to inhibit oxidative damage to cellular DNA by their antioxidant activities, thus preventing mutagenesis and tumorigenesis (Kelly *et al.*, 2001). Epidemiological studies have shown that the chemicals also have the capability to reduce the risk of cardiovascular disease and stroke through inhibition of cellular oxidative damage.

The maturity of fruit and processing techniques applied prior to consumption affected the chemical composition of fruit. This also directly related to the health of consumers who consume the fruits. When the fruit ripen, all sugars, ascorbic acid, soluble solids and pectin will reach their highest levels. Acidity and phenolic acids will show the lowest levels (Bulk *et al.*, 1997). Recently, awareness about the role of antioxidants become higher in the promotion of health due to their function as chemoprotective agents. (Teissedre and Waterhouse, 2000). This awareness has contributed to the rise in consumer demand for fresh fruits. There were enough evidence to show between in vivo and in vitro studies to support the role of antioxidant polyphenolic compounds in the prevention of disease and thus promotion of health.

2.4.4 Sweetness and starch content

Another characteristic of banana ripening is an increase of sugar content (Blankenship *et al.*, 1993). Unripe banana tastes starchy and less sweet since most of the carbohydrate is present in the form of starch, which is a long chain of covalently bonded glucose molecules (Zhang *et al.*, 2005). The starch is converted to soluble solids as the banana fruit starts to ripen. When the banana is fully ripe, the starch content drops from 25% to less than 1% while soluble solids content increase significantly (Cordenunsi and Franco, 1995). The types of sugar present include glucose, fructose, and sucrose, among which more than 70% of the total sugar is sucrose (Marriott *et al.*, 1981). Thus, the banana tastes sweet when ripe. Besides, the sugar content is considered a feasible



UNIVERSITI MALAYSIA



parameter for evaluation of fruit ripening since it gains good correlation with optical properties and physical properties during ripening (Kader, 1999; Liew and Lau, 2012).

2.5 Factors affecting Postharvest Loss of Banana

2.5.1 Pre-harvest factors

Quality assessed after harvest is largely due to the result of conditions and treatments of fruit experience during growth and development and at harvest (Munasque *et al.*, 1990). Pre-harvest factors affecting the postharvest quality of banana, include harvest maturity, type of cultivar, climate, quality of soil, type of chemicals applied, and water status (Thompson 2003). Knowles *et al.* (2001) indicated that the soil type and its fertility affect the chemical composition of banana. Rabus and Streif (2000) also explained that the mineral content of banana, such as phosphorus, calcium and potassium can be used to test their postharvest quality. Water and nutrient uptake abilities are varied for rootstocks used in banana production and in resistance to pests and diseases, and thus have a profound effect on the postharvest life of the produce (Tomala *et al.*, 1999). Ferguson et al. (1999) also cited climate factors have a strong effect on the nutritional quality of fruits. Both light intensity and temperature are crucial factors in the postharvest quality of fruits. Trees that are constantly exposed to sunlight may have a different result in term of quality and postharvest characteristics from those growing on the shady side.

2.5.2 General postharvest factors

Before fruits are consumed, processed or cooked, they remained as living tissues. Therefore, by extending the shelf life of these living produce with controlling respiration technique will improve storability of the fresh produce (Daniel, 2007). Wounded tissues can be formed from minimally processed fruits which lead to browning discolouration and tissue softening. The intensity of the wound response is influenced by a few numbers of factors which included different species and variety, water vapour pressure, O2 and CO2 concentrations, and the presence of inhibitors (Brecht *et al.*,2004). During postharvest handlings like storage and packaging, banana undergo physiological changes such as degradation of chlorophyll, tissue softening, decrease in phenolic acid contents and breakdown of cell materials due to respiration (Sharma and Singh 2000).



- Abdullah, H., Lizada, M. C. C., Tan, S. C., Pantastico, E. B. and Tongdee, S. C. 1990. Storage of banana. In: Hassan, A., and Pantastico, E. B. 1990. Banana Fruit Development, Postharvest Physiology, Handling and Marketing in ASEAN. Pp. 44-64.
- Adam, B. 2015. Assessment of Banana Postharvest Handling Practices and Losses. Journal of Biology, Agriculture and Healthcare **5(17)**: 2224-3208. Retrieved from https://www.researchgate.net/publication/294573321_Assessment_of_Banana_Po stharvest_Handling_Practices_and_Losses_in_Ethiopia on September 20, 2016.
- Argueso C. T., Hansen M. and Kieber, J. J. 2007. Regulation of Ethylene Biosynthesis. J. *Plant Growth Regul.* **26**: 92–105. Retrieved from http://dx.doi.org/10.1007/s00344-007-0013-5 on November 2, 2016.
- Arias, R., Lee, T., Logendra, L. and Janes, H. 2000. Correlation of Lycopene Measured by HPLC with the L*, a*, b* Color Readings of Hydroponic Tomato and the Relation of Maturity with Color and Lycopene Content. Journal of Agricultural and Food Chemistry **48**:1697-1702.
- Atanda, S. A. 2011. The Concepts and Problems of Post-harvest Food Losses in Perishable Crops. *African Journal of Food Science* **5(11)**: 603-613. Retrieved from http://dx.doi.org/10.5897/AJFS on November 10, 2016.
- Aurore, G., Parfait, B. and Hirasmane, L. 2009. Bananas Raw Materials for Making Processed Food Products. *Trends in Food Science Technology* **20(2)**: 78-91
- Blankenship, S. M., Ellsworth, D. D. and Powell, R. L. 1993. A Ripening Index for Bananas Based on Starch Content. *Horticultural Technology* **3(3)**: 338-339. Retrieved from http://horttech.ashspublications.org/content/3/3/338.abstract on November 10, 2016.
- Botondi, R., DeSantis, D., Bellincontro, A., Vizovitis, K., Mencarelli, F. 2003. Influence of Ethylene Inhibition by 1 - methylcyclopropene on the Apricot Quality, Volatile Production, and Glycosidase Activity on Low- and High-aroma Varieties of Apricots J. Agr. Food Chem. **51**:1189–1200.
- Brecht, J.K., Felkey, K., Bartz, J.A., Schneider, K.R., Saltveit, M.E., Talcott, S.T. 2004. A Fresh-Cut Vegetables and Fruits. *Hort Rev* **30**:185–251.
- Bulk, R., Fadil, E., Babiker, E., Tinay, A. 1997. Changes in Chemical composition of types Guava Fruits during Development and Ripening. Food Chem. **59**, 395-399.
- Chai, M., Ho, Y.W., Liew, K.W. and Asif, J. M. 2004.Biotechnology and In Vitro Technique Mutagenesis for Banana Improvement. Downloaded from http://www.fao.org on 1/6/2013
- Chong, L. C. and Noor, A. A. A. 2010. Effects of Banana Flour and b-glucan On the Nutritional and Sensory Evaluation of Noodles. *Food Chemistry* **119**: 34-40
- Cordenunsi, R. and Franco, M. L. 1995. Starch Breakdown during Banana Ripening: Sucrose Synthase and Sucrose Phosphate Synthase. *J Agr. Food Chemistry* **43**: 347-351. Retrieved from http://dx.doi.org/10.1021/jf00050a016 on November 14, 2016.
- Dadzie, B. K. and Orchard, J. E. 1997. Routine Post-Harvest Screening of Banana/Plantain Hybrids: Criteria and Methods. International Plant Genetic Resources Institute. Retrieved from http://www.bioversityinternational.org/uploads/tx_news/Routine_postharvest_sc reening_of _banana_plantain_hybrids__Criteria_and_methods_235.pdf on November 21, 2016.



- Daniel, L. 2007. Innovations in the Development and Application of Edible Coatings for Fresh and Minimally Processed Fruits and Vegetables. Comprehensive Reviews in Food Science and Food Safety, 6: 60–75. doi:10.1111/j.1541-4337.2007.00018.x
- Dharmasena, D.A.N. and Rathnayake, H.C. 2004. Effect of Environmental Temperature Variation on Pysico-Chemical Properties of Tomato in a Sri Lankan Marketing Channel. J. Agric. Eng. 8, 40-48.
- DOA Philippines. 2014. Saba Banana Production Guide. Retrieved from http://cagayandeoro.da.gov.ph/wp-content/uploads/2013/04/SABA-BANANA-PRODUCTI ON-GUIDE.pdf on November 10, 2016.
- Elamin, O. M. A. 2014. Effect of Drying Temperature on Some Quality Attributes of Mango Slices. *In. J. Innov. Sci. Res.* **4(2)**: 91-99. Retrieved from file:///C:/Users/User/Downloads/IJISR -14-130-02.pdf on November 23, 2016.
- Emaga, T., Robert, C., Ronkart, S. N., Wathelet, B. and Paquot, M. 2008. Dietary Fibre Components and Pectin Chemical Features of Peels during Ripening in Banana and Plantain Varieties. *Bioresource Technology* **99**: 4346–4354. Retrieved from http://dx.doi.org/10.1016/j.biortech.2007.08.030 on November 15, 2016.
- Eskin, N. A. M. 1991. Quality and Preservation of Fruits. CRC Press, Boca Raton, FL. Pp. 212-313.
- Faisant, N., Buleon, A., Colonna, P. and Molis, C. 1995. Digestion of Raw Banana Starch in the Small Intestine of Healthy Humans: Structural Features of Resistant Starch. *Brit. J. Nutr.* **73**: 11–123
- FAO. 2013. Banana. Retrieved from http://www.fao.org/3/a-a0185e/a0185e0a.htm
- Ferguson, I., Volz, R., Woolf, A., and Cavalieri, R. P. 1999. Pre-harvest Factors that Affect Physiological Disorders of Fruit. *Postharvest Bio. Tech.* **15**: 255–257
- Fonsah, E. G., 2003. Integrated Quality Control Management Strategies in Banana Fruits Production, Packaging, and Marketing. *J. Food Distribution Res.* **34**: 99-105.
- Fraser, P. D. and Bramley, P. M. 2004. The Biosynthesis and Nutritional Uses of Carotenoids. Prog. Lipid Res. **43**: 228–265
- Galeazzi, M. A. M., Sgarbierri, V. C. and Constantinides, S. M. 1981. Isolation, Purification and Physico-chemical Characterization of Polyphenoloxidases (PPO) from a Dwarf Variety of Banana (*Musa cavendishii* L.). *J. Sci. Food and Agri.* 46: 150–155. Retrieved from http://dx.doi.org/10.1002/jsfa.1006 on November 9, 2016.
- Garder, D. 2012. Measurement of the Fruits and Vegetables Soluble Solids Contents. Retrieved from http://fruitandnuteducation.ucdavis.edu/files/162033.pdf on November 10, 2016.
- Hailu, M. 2013. Review on Postharvest Technology of Banana Fruit. *A. J. Biotech.* **12(7)**: 635-647. Retrieved from http://dx/doi.org/10.5897/AJBX12.020 on November 19, 2016.
- Jobling, J. 2000. Temperature Management is Essential for Maintaining Produce Quality. Sydney Postharvest laboratory Information Sheet. Melbourne, Australia. Retrieved from http:// www.postharvest.com.au. on September 21, 2016.
- Juarez -Garcia, Agama-Acevedo, E., Sáyago-Ayerdi, S. G., Rodríguez-Ambriz, S. L. and Bello-Pérez, L. A. 2006. Composition, Digestibility and Application in Breadmaking of Banana Flour. Plant Foods for Human Nutrition 61: 131-137.
- Kader, A. A. 2002. Quality Parameters of Fresh-cut fruit and Vegetable products. In: Lamikanra O, Editor. Fresh-cut fruits and vegetables. Boca Raton, Fla.: CRC Press LLC. Pp. 11–20.
- Kader, A. A. 1999. The Relationship of Fruit Maturity, Ripening, and Quality. International Symposium on Effect of Pre and Postharvest Factors on Storage of Fruit. Pp. 203-204.
- Kanellis, A. K., Solomos, T. and Mattoo, A. K. 1989. Changes in Sugars, Enzymic Activities and Acid Phosphatase Isoenzyme Profiles of Bananas Ripened in Air or



Stored in 2.5% O2 with and without Ethylene. *Plant Physiology* **90**: 251-258. Retrieved from http://dx.doi.org/10.1104/pp.90.1.251 on November 12, 2016.

- Karamura, E. B. and Karamura, D. A. 1995. *Banana Morphology*. Part 2: The Aerial Shoot. In: *Bananas and plantains*, S.R. Gowen, ed. London: Chapman and Hall. Pp. 190–205.
- Kelly, M., Xu, J., Alexander, K., and Loo, G. 2001. Disparate Effects of Similar Phenolic Phytochemicals as Inhibitors of Oxidative Damage to Cellular DNA. *Mutation Res.* **485**: 309-318.
- Knowles, L., Trimble, M.R., Knowles, N.R. 2001. Phosphorus Status Affects Postharvest Respiration, Membrane Permeability and Lipid Chemistry of European Seedless Cucumber Fruit (*Cucumis sativus* L.). *Postharvest Biol Technol* **21**:179–88.
- Lakshminarayana, S., Subhadva, N. V., and Subramanyam, W. 1970. Some Aspects of Developmental Physiology of the Mango Fruit. *J. Hort. Sci.* **45**: 133-142. Retrieved from http://dx.doi.org/10.1080/00221589.1970.11514339 on November 21, 2016.
- Lelievre, J. M., Latche, A., Jones, B., Bouzayen, M. and Peach, J. C. 1997. Ethylene and Fruit Ripening. Physiol Plant **101**: 727–739
- Madrid M. and Lopez-lee, F., 1998. Differences in Ripening Characteristics of Controlled Atmosphere or Air-stored Bananas. *Acta Horticulturae* 464.
- Maribel, O. V. 2009. Unripe Banana Flour as an Ingredient to Increase the Undigestible Carbohydrates of Pasta. *Food Chemistry* **113**: 121-126. Retrieved from http://dx.doi.org/10.1016/j.foodchem.2008.07.035 on November 20, 2016.
- Marriott, J., Robinson, M. and Karikari, S. K. 1981. Starch and Sugar Transformation during the Ripening of Plantains and Bananas. *Journal of the Science of Food and Agriculture* 32(10): 1021-1026. Retrieved from http://dx.doi.org/10.1002/jsfa.2740321011 on November 17, 2016.
- Mathooko, F. M. 1995. Studies on the Biochemical and Physiological Responses of Fruits and Vegetables to Treatment with Elevated Levels of Carbon Dioxide. Ph.D. Okayama University. Pp 1-166.
- McGuire, R. G. 1992. Reporting of Objective Color Measurements. *Hort Sci.* 27: 1254-1255.
- Medina-Suárez, R., Manning, K., Fletcher, J., Aked, J., Bird, C. R. and Seymour, G. B. 1997. Gene Expression in the Pulp of Ripening Bananas, Two-Dimension Sodium Dodecyl Sulfate-polyacrylamide Gel Electrophoresis of in vitro Translation Production and cDNA Cloning of 25 Different Ripening-related mRNAs. *Plant Physiology* 115: 453-461. Retrieved from http://dx.doi.org/10.1104/pp.115.2.453 on November 23, 2016.
- Molina, A. B. and Roa, V. N. 2000. Advancing Bananas and Plantains R and D in Asia and the Pacific. National Research, Development and Extension Agenda for Banana. Pp. 93.
- Molina, A. B., Eusebio, J. E., Roa, V. N., Vandebergh, I. and Maghuyop, M. A. G. 2003. Advancing Banana and Plantain R and D in Asia and the Pacific. Proceedings of the 2[™] Bapnet Steering Committee Meeting. Jakarta. Pp. 12.
- Nakasone, H. Y., Paull R. E., 1998. Major Tropical Fruits. CAB International, Wallingford, England. 445pp.
- Ovando-Martinez, M., S. Sáyago-Ayerdi, E. Agama-Acevedo, I. Goñi and L.A. BelloPérez. 2009. Unripe Banana Flour as an Ingredient to Increase the Undigestible Carbohydrates of Pasta. Food Chem. **113(1)**: 121–126
- Pariser, E. 1982. Post-harvest of Food Losses in Developing Countries. In: Nutrition Policy Implementation, Springer, Pp. 337-372.
- Pathak, N. and Sanwal, G. G. 1998. Multiple Forms of Polygalacturonase from Banana Fruits. *Phytochemistry* **48**: 249-255. Retrieved from http://dx.doi.org/ 10.1016/S0031-9422(98)00005-3 on November 22, 2016



- Pech, J.C., Bouzayen, M. and Latche, A. 2008. Climacteric Fruit Ripening: Ethylene-Dependent and Independent Regulation of Ripening Pathways in Melon Fruit. *Plant Sci.* **175**: 114–120
- Prasanna, V., Prabha, T. N., Tharanathan, R. N. 2007. Fruit Ripening Phenomena: An Overview. *Critical Reviews in Food Science and Nutrition* **47**: 1–19. Retrieved from http://dx.doi.org/10.1080/10408390600976841 on November 6, 2016.
- Rabus, C., and Streif, J. 2000. The Major Effect of Various Preharvest Treatment on the Development of Internal Browning Disorders in Braeburn apples. Acta Hort **518**: 151–7.
- Reid, M. S. 1992. Ethylene in Postharvest Technology. In: Kader A. A. *Postharvest technology of Horticultural crops*, 2nd edition, technical editor, University of California, division of Agriculture and Natural Resources, Publication 3311. Pp. 97-108.
- Robinson, J. 1996. Bananas and Plantains, Crop Production Science in Horticulture (CAB International, UK).
- Robinson, J. C., and Nel, D. 1984. Banana Bunch Covers Effective in Winter. Nelspruit, South Africa. *Information Bulletin Citrus and Subtropical Fruit Research Institute* **138**: 5-6
- Romani, R.J.; Jennings, W.G. 1971. Stone fruits. In: Hulme, A.C. (ed) The Biochemistry of Fruits and their Products Volume 2. New York, USA; Academic Press. pp. 411-436.
- Rood, P. 1957. Development and Evaluation of Objective Maturity Indices for California Freestone Peaches. Proc. Am. Soc. Hort. Sci. **70**: 104-1 12.
- Saltveit, M. E. 1999. Effect of Ethylene on Quality of Fresh Fruits and Vegetables. *Postharvest Biology and Technology* **15**: 276-292. Retrieved from http://dx.doi.org/10.1016/S0925-5214(98)00091-X on November 8, 2016.
- Seymour, G.B., Taylor, J.E., and Tucker, G.A. 1993. Biochemistry of the fruit ripening. Chapman and Hall Publishers, London, 454 pp.
- Shahjahan, M. S., Sheel, M. A., Zaman, M. A. and Sakur, M. A. 1994. Optimization of Harvesting Maturities for Major Mango Cultivars in Bangladesh. *Bangladesh J. Sci. Res.* **12**: 209-215.
- Sharma, R. M. and Singh, R. R. 2000. Harvesting, Postharvest Handling and Physiology of Fruits and Vegetables. In: Verma, L. R., Joshi, V. K. Editors. *Postharvest Technology of Fruits and Vegetables Vol. 1. Handling, processing, fermentation and waste management*. Tagore Garden, New Delhi: Indus Publishing Co. Pp 94–147.
- Sharrock, S. and Lustry, C. 2000. Nutritive Value of Banana, in INIBAP Annual Report, Montpellier, France. Pp. 28-31.
- Shewfelt, R. L. 2009. *Measuring Quality and Maturity in Postharvest Handling, A System Approach*. Academic press, London, Pp. 461-481.
- Shiga, T. M., Soares, C. A., Nascimento, J. R., Purgatto, E., Lajolo, F. M. and Cordenunsi, B. R. 2011. Ripening Associated Changes in the Amounts of Starch and Non-starch Polysaccharides and Their Contributions to Fruit Softening in Three Banana Cultivars. *Journal of Science Food and Agriculture* **91**: 1511-1516
- Soltani, M., Alimardani, R. and Omid, M. 2010. Prediction of Banana Fruit Quality during Ripening Stage using Capacitance Sensing System. *Australian Journal of Crop Science* **4(6)**: 443-447
- Stover, R. H. & Simmonds, N. W. 1987. *Banana.* 3rd edition. John Wiley & Sons, Inc. New York. Pp. 468.
- Tapre, A.R. 2012. Study of Advanced Maturity Stages of Banana. International Journal of Advanced Engineering Research and Studies E-ISSN2249–8974.



- Teissedre, P. and Waterhouse, A. 2000. Inhibition of Oxidation of Human Low-Density Lipoproteins by Phenolic Substances in Different Essential Oil Varieties. *Journal of Agricultural and Food Chemistry* **48**: 3801–3805. Retrieved from http://dx.doi.org/ 10.1021/jf990921x on November 14, 2016.
- Thompson, A. K. 1996. Postharvest Treatments. In: *Postharvest Technology of Fruit and Vegetables*. Cambridge, Mass.: Blackwell Science Ltd. Pp 95-128.
- Thompson AK. 2003. Preharvest Factors on Postharvest Life. In: Fruit and vegetables. Ames, Iowa: Blackwell Publishing Ltd. p 1–8.
- Tomala, K., Andziak, J., Kobusinski, K., Dziuban, Z., Sadowski, A. 1999. Influence of the Rootstocks on Fruit Maturity and Quality of 'Jonagold' apples. In: Proceedings of the International Seminar, *Apple Rootstocks for Intensive Orchards*, Warsaw Ursynow, Poland, 18–21 August 1999.
- Turner, D. W. 2001. Bananas and Plantains. Pp. 45-77. In: Mitra, S. K. Editor. *Postharvest Physiology and Storage of Tropical and Subtropical Fruits.* CABI Publishing, U.K.
- Ulrich, R. 1974. Organic Acids in Biochemistry of Fruits and Their Product. Hulme, A. C. Editor. Academic press, New York. Pp. 89-118.
- UNCTAD. 2008. Handbook of statistics: Market Information in the Commodities Areas. Retrieved from http://www.unctad.org/ on October 24, 2016.
- Van Den Berg, H., Faulks, R., Granado, H.F., Hirschberg, J., Olmedia, B., Sandmann, G., Southon, S. and Stahl, W. 2000. The potential for the Improvement of Carotenoid Levels in Foods and the Likely Systemic Effects. Journal of the Science of Food and Agriculture 80: 880-912.
- Vincente, A. R. 2007. The linkage Between Cell Wall Metabolism and Fruit Softening: Looking to the Future. *J Sci Food Agric* **87**: 1435–1448. Retrieved from http://dx.doi.org/10.1002/jsfa.2837 on November 21, 2016.
- Volker, B., Puspitasari-Nienaber, N. L., Ferruzzi, M. and Schwartz, S. J. 2002. Trolox Equivalent Antioxidant Capacity of Different Geometrical Isomers of μ-carotene, bcarotene, lycopene and zeaxanthin. Journal of Food and Agricultural Food Chemistry **50**: 221-226
- Wall, M. M. 2006. Ascorbic Acid, Vitamin A, and Mineral Composition of Banana (Musa sp.) Cultivars Grown in Hawaii. J. Food Comp. Anal. 19: 434-445. Retrieved from http://dx.doi.org/10.1016/j.jfca.2006.01.002 on November 24, 2016.
- Wills, R., McGlasson, B., Graham, D. and Joyce, D. 1998. *Postharvest, An Introduction to the Physiology and Handling of Fruit, Vegetables and Ornamentals.* 4th edition, CABI International.
- Yang, D. and Ying, T. 2007. Antioxidant Activities of Various Extracts of Banana. *Asia Pac J Clin Nutr.* **16**: 158-163
- Ye, X., Al-Babili, S., Klöti, A., Zhang, J., Lucca, P., Beyer, P. and Potrykus, I. 2000. Engineering the Provitamin A (β-carotene) Biosynthetic Pathway into (Carotenoid-Free) Rice Endosperm. Science **287**: 303-305. Retrieved from http://dx.doi.org/ 10.1126/science.287.5451.303 on November 19, 2016.
- Zhang, P., Whistler, Miller, R. L. and Hamaker, J. M. 2005. Banana Fruits Starch: Production, Physicochemical Properties, and Digestibility: A review. *Carbohydrate Polymers* **59(4)**: 443-458. Retrieved from http://dx.doi.org/ 10.1016/j.carbpol.2004.10.014 on November 19, 2016.

