EFFECT OF CHICKEN MANURE BIOCHAR AND WATER STRESS ON ROOT CHARACTERISTICS AND YIELD OF PURSLANE (Portulaca oleracea L.)

SHUADINATY YUSOFF

DISSERTATION SUBMITTED IN PARTIAL FULFILMENT OF THE REQUIRMENTS FOR THE DEGREE OF BACHELOR OF AGRICULTURE SCIENCE WITH HONOURS

CROP PRODUCTION PROGRAMME FACULTY OF SUSTAINABLE AGRICULTURE UNIVERSITI MALAYSIA SABAH 2017



١.

UNIVERSITI MALAYSIA SABAH

| BORANG P | ENGESAHAN TESIS |
|--|---|
| JUDUL: EFFECT OF CHICKEN MANURE BLOCH | AR AND WATER STRESS ON ROOT CHARACTERISTICS |
| AND YIELD OF PURSLANE (Portulaca oler | |
| | |
| UAZAH: IJAZAH SARJANA MUDA SAINS PERTA | NIAN DENGAN KEPUJIAN |
| SAYA : SHUADINATY YUGOFF | SESI PENGAJIAN : _2016/2017 |
| (HURUF BESAR) | |
| Mengaku membenarkan tesis *{LPSM/ Sarjana/Dokto Sabah dengan syarat-syarat kegunaan seperti berikut: | r Falsafah) ini disimpan di Perpustakaan Universiti Malaysia - |
| 1. Tesis adalah hak milik Universiti Malaysia Saba | h. |
| | arkan membuat salinan untuk tujuan pengajian sahaja. |
| | esis ini sebagai bahan pertukaran antara Institusi pengajian |
| tinggi. 4. Sila tandakan (/) UNIVERSIT | NISTRALAN INALAYSIA SACTO |
| | nat yang berdarjah keselamatan atau kepentingan Malaysia b di AKTA RAHSIA RASMI 1972) |
| TERHAD (Mengandungi maklur mana penyelidikan dij | nat TERHAD yang telah ditentukan oleh organisasi/badan di alankan) |
| | |
| | Disahkan oleh: NURULAIN BINTI ISMAIL |
| | - PUSTAKAWAN KANAN |
| STATE_ | CAMERSITI MALAYSIA SABAH |
| (TANDATANGAN PENULIS) | (TANDATANGAN PUSTAKAWAN) |
| Alamat Tetap: <u>679, LORONG</u> | |
| MERUNTUM 2, TAMAN PANTAL LOKKAWI, 89580 PAPAR, SABAH | |
| CUREAWI, DISSO THINK SHOW | DR. MOHAMADU BOYIE JALLOH |
| | (NAMA PENYELIA) |
| TARIKH: 13/01/204 | TARIKH: 13/01/2017 |
| menyatakan sekali sebab dan tempoh tesis ini perlu | t daripada pihak berkuasa/organisasi berkenaan dengan dikelaskan sebagai SULIT dan TERHAD. Falsafah dan Sarjana Secara Penyelidikan atau disertai |
| bagi pengajian secara kerja kursus dan Laporan Proje | |
| | |

DECLARATION

I hereby declare that this dissertation is based on my original work except for citations and quotations which have been duly acknowledged. I also declare that no part of this dissertation has been previously or concurrently submitted for a degree at this or any other university.

.

SHUADINATY & SOFF BR13110166 13th JANUARY 2017



1. DR. MOHAMADU BOYIE JALLOH SUPERVISOR



ACKNOWLEDGEMENT

I would like to express my deepest appreciation to Dr. Mohamadu Boyie Jalloh, for his guidance, advice and supervision throughout the progress of my Final Year Project. Despite his busy schedule, there was always time made for me and other students under his supervision. He put his faith in me and urged me to do better.

I am highly indebted and thoroughly grateful to Mr. Rohizan Basir, Mrs. Nurul Syakina Marli, Mr. Razalie, Mrs. Ahjia Jekan (FSA laboratory assistants) as well as Mr. Frederick Florentius @ Firdaus, Mr. Yohanes and Mrs. Sitti Aminah from the Farm Administration Office, who gave permission to use all required apparatus and the necessary materials from the field until the laboratory phase. I am also grateful to Mrs. Anika Bulangou @ Flora for helping me shape up my laboratory skills.

Special thanks goes to my housemates, juniors and alumni of FSA, who helped me in the earlier progress of my project which I could not have it alone. Last but not least, I would like thank my parents, Mr. Hj. Yusoff and Mrs. Rose. Their investment in my life and knowing how proud they are means more than they know. I am so thankful to have these people in life whom I can trust in hard times and celebrate accomplishments with.



ABSTRACT

Purslane (*Portulaca oleracea* L.), aside from being known as a weed, is a healthy food that has been highly beneficial to humans as a nutritious vegetable and folk medicine for centuries, and recognized by the WHO as one of the most used medicinal plants. In this study the root characteristics and yield of purslane with biochar application and water stress was analyzed. With 45 polybags used during the study, biochar was added to the planting medium two weeks before planting at rates of 0 t/ha, 10 t/ha and 20 t/ha. One-week old purslane seedlings were exposed to drought conditions by giving different watering intervals of 1 day, 2 days and 3 days. After harvest, the fresh and dry weight of purslane was taken, while the root characteristics, such as total root length, root surface area and root/shoot ratio were taken by WinRHIZO software, as well as soil pH, total nitrogen and total available phosphorus as the soil chemical properties. For plant growth parameters, Treatment B10 (10t/ha biochar rate) showed the highest value of plant fresh weight (105.78 g) and plant dry matter (9.12 g). While Treatment 1D (1 day watering interval) showed the highest value of plant fresh weight of 115.85 g and plant dry matter of 10.30 g. As for root characteristics, Treatment B10 resulted in the highest value of root surface area, total root length and root/shoot ratio with 676.30 cm², 2941 cm and 0.19, respectively. For root surface area, Treatment 1D resulted in the highest value with 743.18 cm², Treatment 3D (3 days watering interval) showed the highest total root length (3399 cm) and Treatment 2D resulted in the highest root/shoot ratio (0.20). Soil chemical properties such as pH and total nitrogen were greatly affected by water stress compared to biochar rate, except for total available phosphorus. However, the results showed that there was no significant interaction between the biochar rate and watering intervals. This study concludes that, biochar had no significant effects on both plant growth and root characteristics due to the short period of application before planting, therefore less nutrients were released into the soil.



KESAN BIOCHAR DAN KETEGASAN AIR TERHADAP CIRI-CIRI AKAR DAN HASIL TANAMAN PURSLANE

ABSTRAK

Tanaman purslane (Portulaca oleracea L.), selain dikenali sebagai rumpai, ia adalah makanan yang sihat yang memberi banyak manfaat kepada manusia sebagai sayursayuran dan ubat-ubatan selama berabad, dan diiktiraf oleh WHO sebagai salah satu tumbuhan ubatan yang paling banyak digunakan. Dalam kajian ini, ciri-ciri akar dan hasil tanaman purslane dengan aplikasi biochar dan kuantiti air dianalisis. Dengan menggunakan 45 pasu semasa kajian, biochar telah ditambah kepada medium penanaman dua minggu sebelum menanam pada kadar 0 t/ha, 10 t/ha dan 20 t/ha. Anak pokok purslane berumur satu minggu telah diberi tekanan kemarau dengan memberikan hari penyiraman berbeza pada selang 1 hari, selang 2 hari dan selang 3 hari. Selepas menuai, berat basah dan kering telah diambil, manakala ciri-ciri akar seperti jumlah panjang akar, kawasan permukaan akar dan nisbah akar/pucuk telah diambil menggunakan perisian WinRHIZO, serta pH tanah, jumlah nitrogen dan jumlah fosforus sebagai sifat-sifat kimia tanah. Bagi parameter pertumbuhan pokok, Rawatan B10 (10t/ha biochar) menunjukkan nilai tertinggi bagi berat segar (105.78 g) dan berat kering (9.12 g). Manakala Rawatan 1D (selang 1 hari) menunjukkan nilai tertinggi bagi berat segar sebanyak 115.85 g serta berat kering sebanyak 10.30 g. Bagi ciri-ciri akar, Rawatan B10 mencatatkan nilai tertinggi bagi kawasan permukaan akar, jumlah panjang akar dan nisbah akar/pucuk yang masing-masing bernilai 676.30 cm², 2941 cm dan 0.19. Bagi kawasan permukaan akar, Rawatan 1D mencatatkan nilai tertinggi sebanyak 743.18 cm², Rawatan 3D (selang 3 hari) menunjukkan nilai tertinggi bagi jumlah panjang akar (3399 cm) dan Rawatan 2D mencatatkan nilai nisbah akar/pucuk tertinggi (0.20). Ciri-ciri kimia tanah seperti pH dan jumlah nitrogen telah dipengaruhi oleh ketegasan air, kecuali jumlah fosforus. Walau bagaimanapun, keputusan menunjukkan bahawa tidak ada interaksi yang bererti diantara kadar biochar dan selang penyiraman. Kajian ini menyimpulkan bahawa, biochar tidak mempunyai kesan yang besar ke atas kedua-dua pertumbuhan tumbuhan dan ciri-ciri akar kerana tempoh aplikasi adalah singkat sebelum menanam, oleh itu tidak banyak nutrien telah dibebaskan ke dalam tanah.



TABLE OF CONTENTS

| Contents DECLARATION VERIFICATION ACKNOWLEDGEMENT ABSTRACT ABSTRAK LIST OF TABLES LIST OF FIGURES LIST OF ABBREVIATIONS | Page ii iv v v vi ix x xi |
|---|---|
| CHAPTER 1 INTRODUCTION | |
| 1.9Background1.10Justification1.11Objectives1.12Hypotheses | 1 2 3 3 |
| CHAPTER 2 LITERATURE REVIEW | |
| 2.1 Purslane 2.1.1 Morphology 2.1.2 Physiology 2.1.3 Nutritional Values | 4 5 6 7 |
| 2.2 Water and Agriculture 2.2.1 Effect of Water Deficit on Plants 2.2.2 Effect of Water Deficit on Root Cl 2.2.3 Effect on Excessive Water on Plants | haracteristics 10 hts 11 |
| 2.3 Biochar and Their Properties 2.3.1 Types of Biochar | 11 |
| 2.4 Effect of Biochar on Soil Properties 2.4.1 Soil Water Retention 2.4.2 Soil pH 2.4.3 Soil Fertility 2.5 Effect of Biochar on Plant | 12 12 14 15 16 |
| 2.5.1 Plant Growth 2.5.2 Root Growth | 16 16 |



CHAPTER 3 METHODOLOGY

| 3.1 | Location | and Du | uration of Study | 18 |
|-----|-----------|-----------|--|----|
| 3.2 | Materials | ; | | 18 |
| 3.3 | Methods | | | 18 |
| | 3.3.1 E | Experim | nental Design and Treatments | 19 |
| | 3.3.2 P | Planting | Medium Preparation and Sowing of Seeds | 19 |
| | 3.3.3 H | larvest | ing | 19 |
| | 3.3.4 P | Paramet | ters | 20 |
| | 3. | .3.4.1 | Plant Yield | 20 |
| | 3. | 3.4.2 | Root Characteristics | 20 |
| | 3. | .3.4.3 | Soil Chemical Properties | 20 |
| | 3.3.5 S | Statistic | al Analysis | 20 |
| | | | | |

CHAPTER 4 RESULTS AND DISCUSSION

| 4.1 | Yield c | of Purslane | 21 |
|------|---------|-----------------------------|----|
| | 4.1.1 | Plant Fresh Weight | 21 |
| | | Plant Dry Matter | 23 |
| 4.2 | | Characteristics of Purslane | 25 |
| | | Root Surface Area | 25 |
| | 4.2.2 | Root Total Length | 27 |
| | 3.3.3 | Root/Shoot Ratio | 29 |
| 4.3 | Soil Ch | nemical Properties | 30 |
| | 4.3.1 | рН | 31 |
| | | Soil Total Nitrogen | 32 |
| | 4.3.3 | Total Available Phosphorus | 33 |
| CHA | PTER 5 | CONCLUSION | 35 |
| REF | ERENCES | S | 36 |
| APPI | ENDIX A | | 42 |
| APPI | ENDIX B | 8 | 43 |
| | | | |
| | | | 48 |



LIST OF TABLES

| Table | | Page |
|-------|---|------|
| 2.1 | Other names of Portulaca oleracea by language and country | 5 |
| 2.2 | Composition of nutrients per 100 g of raw purslane | 8 |
| 2.3 | Soil pH and plant growth | 14 |
| 3.1 | Treatments of the experiment | 19 |
| 4.12 | Chemical properties of the soil before planting | 30 |
| 4.13 | Mean soil pH for various treatments | 31 |
| 4.14 | Mean soil pH for various treatments | 31 |
| 4.15 | Mean Soil Total Nitrogen for various treatments | 32 |
| 4.16 | Mean Soil Total Nitrogen for various treatments | 32 |
| 4.17 | Mean total available phosphorus for various treatments | 33 |
| 4.18 | Mean total available phosphorus for various treatments | 33 |

.



LIST OF FIGURES

| Figure | | Page |
|--------|--|------|
| 2.1 | Purslane (Portulaca oleracea) without its flower | 6 |
| 3.1 | Layout of the experiment | 19 |
| 4.1 | Mean plant fresh weight for the various treatments | 21 |
| 4.2 | Mean plant fresh weight for the various treatments | 22 |
| 4.3 | Mean plant dry matter for the various treatments | 23 |
| 4.4 | Mean plant dry matter for the various treatments | 24 |
| 4.5 | Mean root surface area for the various treatments | 25 |
| 4.6 | Mean root surface area for the various treatments | 26 |
| 4.7 | Mean total root length for the various treatments | 27 |
| 4.8 | Mean total root length for the various treatments | 28 |
| 4.9 | Mean root/shoot ratio for the various treatments | 29 |
| 4.10 | Mean root/shoot ratio for the various treatments | 29 |



.

LIST OF SYMBOLS, UNITS AND ABBREVIATIONS

| ANOVA Ca | Analysis of Variance Calcium Centre for Agriculture and Bioscience International |
|-------------|--|
| CABI CAM | Crassulacean Acid Metabolism |
| CEC | Cation Exchange Capacity |
| CHN | Carbon Hydrogen Nitrogen |
| CRD | Completely Randomized Design |
| DAS | Days After Sowing |
| DOM | Dissolved Organic Matter |
| FSA | Faculty of Sustainable Agriculture |
| К | Potassium |
| LOI | Loss-on-ignition |
| Mg | Magnesium |
| N | Nitrogen |
| OB | Olive tree pruning biochar |
| Р | Phosphorus |
| РВ | Pine woodchip biochar |
| PEPC | Phosphoenolpyruvate carboxylase |
| RHB | Rice husk biochar |
| UMS | Universiti Malaysia Sabah |
| WB | Wood biochar |



CHAPTER 1

INTRODUCTION

1.1 Background

The genus Portulaca, which belongs to the Portulacaceae and sub family Portulacarioidae, has more than 120 species grown worldwide. *Portulaca oleracea*, or sometimes referred to as "pigweed", is widely considered as a weed due to its invasive characteristics. Purslane is not only cultivated, it also grows wild in fields and disturbed soils, as well as in open grassland and bushland, up to an altitude of 2,400 meters above sea level (Grubben and Denton, 2004). Purslane is a very good source of alpha-linolenic acid is an omega-3 fatty acid which plays an important role in human growth and development and in preventing diseases (Kamal Uddin *et al.*, 2014). Hence, many Asian and European regions grow purslane as a staple leafy vegetable. Other than human consumption, purslane has long been used for animal feed in Thailand (Siriamornpun and Suttajit, 2010). Seeds take 7 to 10 days to germinate at temperatures ranging between 20 and 32 degrees Celsius. Harvesting it can start 3-4 weeks after sowing, and 2-3 cuts at 2-3 week intervals are possible in commercial production (Grubben and Denton, 2004).

Water stress is the main abiotic stress that is perceived as a limiting factor in crop production worldwide compared to other environmental stresses. Water stress is a result of water deficit, such as a drought or high soil salinity. Every year, water stress on arable plants in different parts of the world disrupts agricultural activities and food supply (Ismail and Hasegawa, 2012). Water is an integral part of the plant body that plays a key role in growth initiation, maintenance of the developmental process of plant life and hence has a critical function in crop production (Aslam *et al.*, 2013). Scarcity of water results in developmental and morphological changes such as growth inhibition and reproductive failure, reduction of water content and turgor, stomatal



UNIVERSITI MALAYSIA SABAI

closure, limits gaseous exchange, reduction in transpiration and reduction in carbon assimilation rates. Therefore, drought stress affects crop yield at all stages of growth. The roots of plants are important for anchor and the uptake of nutrients and water by plant for various physiological, biochemical and other plant processes. Usually the growth of plant roots is influenced by genetic characteristics of the plant and various environmental factors such as soil type and properties, and climate. Soil conditions or health can be improved by amending the soil with either chemical or organic materials. However, the use of organic amendments such as biochar is more favoured as a sustainable agronomic practice.

Biochar is a term used to describe a carbon-rich product which is added to the soil, produced through a process called pyrolysis. Biochar can be created from wood, plant materials and manure (International Biochar Initiative, 2015). It has potential as an important tool for improving food security and diversity of agricultural land in areas with depleted soils, scarce organic resources and inadequate water supply and limited chemical fertilizers. Biochar also improves the quality and quantity of water and increases the retention of soil nutrients and agricultural chemicals for use by plants and crops. In the soil, biochar provides a good habitat for soil microbes such as bacteria that help in recycling of nutrients so that nutrients can be absorbed by plants.

Justification

Purslane has been highly beneficial to humans as a nutritious vegetable and folk medicine, for centuries, despite also being known as an invasive weed. Scientific analysis of its chemical components has shown that this common weed has uncommon nutritional value, making it one of the potentially important foods for the future (Kamal Uddin *et al.*, 2013). The usefulness of biochar for soil improvement and the environment has been recognized since Terra Preta or "black earth" was discovered and it is now used worldwide. According to Zheng *et al.*, (2010) adding biochar to fertile soils can reduce the need to use of chemical fertilizers.

Most farmers in Malaysia still do not know the other potentials of purslane because the uses are limited to folk medicines and as a herb and decorative plant. Hence, this study was done to find out how the purslane plant, including the roots, responds to biochar amendment and water stress. This study can contribute to increasing the



UNIVERSITI MALAYSIA SABAH



production and economic value of purslane since it has potential to be widely used in agriculture, such as living mulch and animal feed. This study can also help towards the effective utilization of biochar in agriculture. Also, biochar can be used to help combat climate change by holding carbon to soil and displacing fossil fuel use (International Biochar Initiative, 2015).

1.2 Objectives

The objectives of this study were:

- i. To evaluate the effects of biochar and water stress on the root characteristics of purslane.
- ii. To evaluate the effect of biochar and water stress on yield of purslane.

1.3 Hypotheses

- H_0 : The use of biochar and drought stress has no significant effect on root characteristics of purslane.
- H_A: The use of biochar and drought stress has significant effect on root characteristics of purslane.
- H_0 : The use of biochar and drought stress has no significant effect on yield of purslane.
- H_A: The use of biochar and drought stress has significant effect on yield of purslane.



CHAPTER 2

LITERATURE REVIEW

2.1 Purslane

Purslane, or its botanical name *Portulaca oleracea L*. is a succulent annual plant which belongs to the Portulacaceae family has worldwide distribution. The seeds of purslane are able to remain viable in soil for up to 40 years. Aside from being a noxious weed, purslane is also known as healthy food that contains a whole range of health-boosting nutrients, including higher omega-3 fatty acids, antioxidant vitamins and minerals, making it listed in the World Health Organization (WHO) as one of the most used medicinal plants and given the term 'Global Panacea' (Samy *et al.*, 2005). Many cultures embrace this plant as food including Europe, the Middle East, Asia and Mexico. Purslane is likely native to North Africa, the Middle East and the Indian subcontinent. It has been grown for more than 4,000 years as food and traditional medicine in China and parts of India.

Purslane is now naturalized in most parts of the world, both tropical and temperate region. It can be found in flower beds, cultivated fields, and roadsides or other disturbed or waste places. Also, there are some studies carried out to evaluate the potential of purslane as living mulch in broccoli production. Worldwide, purslane is also known by several names as shown in Table 2.1.



| Name | Language/Country | |
|-------------|---------------------|--|
| Purslane | English | |
| Krokot | Indonesia, Malaysia | |
| Портулак | Russia | |
| Persleyn | Philippines | |
| Mă chĭ xiàn | Chinese | |
| Genda pala | Sri Lanka | |
| Lunia | Hindi | |
| Pourpier | France | |
| Suberi-hiyu | Japan | |
| | | |

Table 2.1: Other names of *Portulaca oleracea* by language and country

Other names of *Portulaca oleracea* according to language and country Source: Anonymous, 2005

2.1.2 Morphology

According to Grubben *et al.*, (2004), purslane is an annual herb which has a stem with many branches; glabrous; up to 35 cm long; often red or brown in colour, with short internodes. The leaf are alternate and simple with a hairy ocrea up to 3mm long, often silvery-white in colour, irregularly fringed and has short petiole. The leaf blade is very small, linear to obovate-elliptical, 1-2 cm x 2-5 mm in size, rather thick and leathery and has dark green colour turning red. While, the inflorescence has a congested raceme with short branches and has 1-5 flowered clusters. The flowers are bisexual with perianth of 2 mm long, greenish with 4 lanceolate-elliptical lobes of 1.5 mm long, the outer pair keeled, white to pale pink coloured; have 5-8 stamens and ovary is superior with 1-celled styles, free and 3 mm long. The fruit has a trigonous nut up to 2 mm long, shiny black and smooth textured. The roots are rather thick with many fibrous secondary roots. For efficient absorption, the root system may reach 153 cm in diameter. In addition, it produces adventitious roots that emerge only from the cut or broken surface of stem fragments (Mitich 1997).





Figure 2.1 Purslane (Portulaca oleracea) without its flower

2.1.3 Physiology

Portulaca oleracea is mostly an annual, but it may be perennial in the tropics. It grows rapidly in open, disturbed soils. Purslane produces flowers, fruits and seeds within 6 weeks of germination. It has a wide tolerance of photoperiod, light intensity, temperature, soil type and moisture. This species is self-compatible which means that it is able to breed by its own pollen (CABI, 2015).

Most plants have evolved three photosynthetic pathways known as C3, C4, or Crassulacean Acid Metabolism (CAM). Purslane has the ability to adapt to a broad range of environmental conditions due, in large part, to its unique photosynthetic metabolism. Under sufficient water supply, purslane utilizes C4 photosynthetic metabolism. However, purslane will shift to a CAM-like photosynthetic metabolism under water stressed conditions, with nocturnal acid accumulation in the leaves and reduced CO_2 uptake. Phosphoenolpyruvate carboxylase (PEPC) catalyzes the initial step



UNIVERSITI MALAYSIA



in photosynthetic fixation of atmospheric CO_2 in C4 and CAM plants. Regulation of PEPC is primarily via phosphorylation by PEPC kinase (PEPCK). For well-watered purslane, PEPC and PEPCK transcript abundance are indicative of C4 metabolism, but in water stressed purslane, PEPCK transcripts accumulate at night, suggesting a shift in the phosphorylation pattern of PEPC to CAM-like metabolism (Proctor, 2013).

This allows the leaves to trap carbon dioxide at night and convert it to malic acid. Then during the daytime, the malic acid is converted into glucose. Due to this characteristic, purslane leaves will taste different in the morning and in the evening, when it has 10 times more malic acid (Blair, 2014).

2.1.4 Nutritional values

Purslane is one of the leafy vegetables containing high levels of vitamin E and essential omega-3 acids called alpha-linoleic acid (ALA). Omega-3 fatty acids are considered essential fatty acids. They are necessary for human health, but cannot be produced inside the body and have to be obtained from foods (Ehrlich, 2015). These essential fatty acids play key roles in maintaining the heart and enriching brain health. Purslane is high in vitamins A and C which are known for their antioxidant powers. It is also low in calories and fat, but this weed does contain high amounts of dietary minerals such as iron, magnesium, calcium, potassium and manganese (Melgren, 2010). The nutritional values per 100 g purslane are shown in Table 2.2.



| Composition | Raw pu | rslane |
|------------------------------|----------|--------|
| - | (100 | g) |
| Energy | 20 kcal | |
| Carbohydrates | 3.39 g | |
| Fat | 0.36 g | |
| Protein | 2.03 g | |
| Vitamins | | |
| Vitamin A | 1320 IU | |
| Thiamine (B ₁) | 0.047 mg | 4% |
| Riboflavin (B ₂) | 0.112 mg | 9% |
| Niacin (B ₃) | 0.48 mg | 3% |
| Vitamin B ₆ | 0.073 mg | 6% |
| Folate (B9) | 12 µg | 3% |
| Vitamin C | 21 mg | 25% |
| Vitamin E | 12.2 mg | 81% |
| Minerals | | |
| Calcium | 65 mg | 7% |
| Iron | 1.99 mg | 15% |
| Magnesium | 68 mg | 19% |
| - | (19%) | |
| Manganese | 0.303 mg | 14% |
| Phosphorus | 44 mg | 6% |
| Potassium | 494 mg | 11% |
| Zinc | 0.17 mg | 2% |
| Water constituent | 92.86 g | |

Table 2.2 Composition of nutrients per 100 g of raw purslane

Source: USDA, 2006

2.2 Water and Agriculture

Today, agriculture accounts for around 70% of water used in the world. Water resource is derived primarily from rain and is drained via ditches, streams and stored in dams, ponds, swamps and lakes. Water is a major constituent of living organisms, and comprises about 80 - 90% of fresh weight of herbaceous plants and over 50% of



UNIVERSITI MALAYSIA SABAH

woody plants (Rashidi and Seyfi, 2007). The functions of water in plants include the transport of nutrients and organic compounds. The material is transported in the form of nutrients from the soil and organic materials from photosynthesis and other cells; is the basis for biochemical reactions such as photosynthesis; is a buffer in the regulation of plant body heat; and the main constituent of protoplasm (Anonymous, 2003). In agriculture, water is needed for irrigation and watering. For example, a crop that requires irrigation is wetland rice where without water stagnation, it will not survive hence, affecting rice production. Also, water is essential for livestock to determine the performance of animals. For example, a laying hen that gets insufficient water will be affected in terms of laying performance, because water is vital in egg formation. Likewise, a dairy cow that has insufficient water will produce less milk, because water is the highest constituent in fresh cow milk (Akinbobola, 2015).

2.2.1 Effect of Water Deficit on Plants

Drought is defined as an extended period of unusually dry weather when there is not enough rain (Rodgers, 2013). It is a major threat which limits crop production as the growth and development of plants are severely affected by water stress. To date, the effects of drought stress on plants have been studied in many plants, including the effects on crop yield. Drought stress affects crop yield by reducing grain yield and all yield components, but also affects seed quality such as germination and vigour tests (Ahmad *et al.*, 2011). According to Tahar *et al.*, (2010) the different treatments of water stress led to a decline in the shoot and root dry weights, as well as total dry weight per plant.

Plant growth under drought is affected by altered physiological processes such as photosynthesis, transpiration and hormonal regulation. Photosynthesis is a biochemical process of nutrient establishment, such as carbohydrates, from water and carbon dioxide with the help of light energy. Photosynthesis will only occur in the presence of chlorophyll, a pigment that serves as a catcher and converts sunlight energy into chemical energy bound in carbohydrate molecules. The amount of water, sunlight and carbon dioxide available to the plant directly influences the amount of food a plant can produce for itself. In recent research using *Solanum scabrum* Mill., the leaf photosynthetic rate was tremendously reduced by decrease in stomatal





conductance under stress conditions, resulting in decreased leaf, stem and root growth (Assaha *et al.*, 2016).

Transpiration is the process by which moisture is carried through plants from roots to small pores on the underside of leaves, where it changes to vapour and is released to the atmosphere (Anonymous, 2016). Transpiration serves as transport of minerals, maintaining optimum turgidity and eliminates a large amount of heat from the leaves. Minerals absorbed into the roots move up the plant through the xylem. It also helps in the sorption of minerals from the soil and transports it through the system. Under conditions of water deficit, the stomatal pores of many mesophytes will close, therefore reducing transpiration significantly.

Hormones play a special role in plant reaction to water stress conditions. Abscisic acid (ABA) is the main hormone correlated to water stress. Plants under drought conditions substantially increase in the levels of ABA in shoots and roots (Nan *et al.*, 2002; Wang *et al.*, 2008; Leach *et al.*, 2011).

2.2.2 Effect of Water Deficit on Root Characteristics

As a plant organ, roots function as absorber of water, mineral salts and oxygen from the soil and are also an anchor for the establishment of the plant. Because the root system greatly affects the absorption of nutrients and water that plants need for photosynthesis, thus it increased plant productivity (Zulkifli, 2016).

Besides the reduction in dry weight, drought stress treatment showed a notable effect on the root/shoot ratio. The increase in root/shoot ratio was a result of a greater reduction in aboveground biomass rather than an increase in root biomass, because under drought stress the absolute root dry weight was not greater than that under well-watered treatment (Xu *et al.*, 2015). A study on root physiological traits and root biomass of *Reaumuria soongorica* by Shan *et al.*, (2015) showed that the seedlings retained higher root activity in serious drought stress by enhancing root respiration. Aslam *et al.*, (2013) observed that drought resistant maize had the maximum root fresh weight, compared to less drought resistant varieties.



2.2.3 Effect on Excessive Water on Plants

Too much water can bring damaging consequences. It can impact crop growth in both the short and long term. Waterlogging in crop plants results in oxygen deprivation as the excess water does not react chemically with the plant. Plants need oxygen for cell division, growth and the uptake and transport of nutrients. Since oxygen diffuses through undisturbed water much more slowly than in a well-drained soil, oxygen requirements rapidly exceed that which is available in the event of excess water (Ransom, 2013). Excess water promotes pathogen growth especially fungi. Fungi grow best in warm (70 to 90° F) conditions under limited light and lots of moisture. Large populations of fungi will attack plants stressed by excessive water. Bacterial and viral disease can also take advantage of these plants as a host to spread diseases. On the bottom parts of the plant, the wet roots succumb to root rot mainly caused by pathogen attack (Sprague, 2015). In terms of root growth, waterlogging impedes the oxidative breakdown of ethylene which influences root growth and functions such as root extension, while carbon dioxide in the soil can severely damage roots of certain species (Jackson, 2013).

2.3 Biochar and Their Properties

In recent years, health and quality of the soil have become a major concern in the agricultural field with increased efforts to reduce the use of chemicals which bring adverse impacts to the environment. Thus, many studies have been carried out on biochar to evaluate its potential as a soil amendment. The term biochar was first coined by Peter Read, referring to any charcoal that is used for amending soil (Read, 2009). Biochar comes from the combustion of organic materials from agricultural residues, such as grass, wood, rice husk, palm oil or animal manures, and is produced through burning under less oxygen, a process known as pyrolysis. However, this process differs from the formation of charcoal, because biochar is produced for the purpose of applying it to the soil.

Its history dates back 2,000 years to a civilization in the Amazon Basin where a carbon rich, highly fertile soil known as "Terra Preta" or black earth was discovered and analyzed, revealing high concentrations of charcoal and organic matter, such as animal and plant residues (Biochar Now, 2016). Terra Preta soils contain high



UNIVERSITI MALAYSIA SABAH



concentrations of nitrogen, phosphorus, potassium, calcium, and stable organic matter (Glaser *et al.*, 2001). The formation of this rich soil comes from the "slash-and-char" process practiced by the ancient Amazonians. In this process, plant material or crop remains were cut, burned, and buried to smolder, which eventually produced char, now commonly referred to as "biochar" or "agrichar" (Anonymous, 2016). The addition of biochar into the soil shows various of benefits in terms of soil quality including improving the cation exchange capacity (CEC); reducing soil acidity; improving soil structure; improving water holding capacity; and improving the efficiency of fertilization.

2.3.1 Types of Biochar

Biochar can be produced from various sources of biomass such as rice husks, wood remains, nut shells, manure and crop residues. The commonly used biochar includes that from rice husk, wood, empty fruit bunch and livestock manure.

Uzoma *et al.*, (2010) reported that application of cow manure biochar to sandy soil is not only beneficial for crop growth but it also significantly improved the physicochemical properties of the coarse soil. A field experiment by Varela *et al.*, (2013) stated that wood biochar (WB) added to soil increased the plant weight of water spinach by increasing the root size and leaf width, while rice husk biochar (RHB) added to soil increased the plant weight of water spinach by increasing the stem size and leaf length.

2.4 Effect of Biochar on Soil Properties

The amendment of biochar to the soil results in changes to the physical properties, chemical properties and biological properties of the soil. However the changes in soil properties depend on the types of biochar, application rate and the soil type itself.

2.4.1 Soil Water Retention

Soil water is the primary component of the soil in relation to plant growth. If the soil moisture content is optimum for plant growth, plants can readily absorb soil water. Soil is able to hold water due to their colloidal properties and aggregation qualities. The water is held on the colloids surface and other particles and in the pores (Anonymous,



UNIVERSITI MALAYSIA SABAI

REFERENCES

- Abbasi, M. K. and Ahsan, A., A. 2015. Ameliorating Effects of Biochar Derived from Poultry Manure and White Clover Residues on Soil Nutrient Status and Plant growth Promotion- Greenhouse Experiments. PLoS One site. http://journals.plos.org/plosone/article?id=10.1371/journal.pone.0131592.
 March 2016. Verified on 20 March 2016
- Abdulrazzaq, H., Jol, H., Ahmed Husni, A., and Abu-Bakr, R. 2014. Biochar from Empty Fruit Bunches, Wood, and Rice Husks: Effects on Soil Physical Properties and Growth of Sweet Corn on Acidic Soil. *Journal of Agricultural Science* **7(1)**: 192-200
- Agbede, T. M., Ale, M. O., Alagha, S. A., Ojomo, A. O. and Oladitan T. O. 2010. Comparative Evaluation of Poultry Manure and NPK Fertlizer on Soil Physical and Chemical Properties, Leaf Nutrient Concentrations, Growth and Yield of Yam (*Diascorea rotundata* Poir) in Southwestern Nigeria. *Journal of soil science* 6(5): 540-546
- Abiven, S., Hund, A., Martinsen, V. and Cornelissen, G. 2015. Biochar Amendment Increases Maize Root Surface Areas and Branching: A Shovelomics Study in Zambia. *Plant Soil* **(395)**: 45–55
- Akinbobola, A. 2015. Water and Its Importance To Livestock. A Livestocking site. http://www.livestocking.com/water-and-its-importance-to-livestock-2. Access on 6 March 2016. Verified on 20 March 2016
- Ahmad, A. M., Nezar, H. S., and Mullen, R. E. 2011. Drought Stress Effect on Crop Pollination, Seed Set, Yield and Quality. *Sustainable Agriculture Reviews 6*, 6:193-213
- Anonymous. 2003. Functions of Water in Plants. A Pennsylvania State University site. http://www.personal.psu.edu/faculty/a/s/asm4/turfgrass/education/turgeon/les sons/lesson05/corefiles/links/effectsmoist/1.html. Access on 30 April 2016. Verified on 7 May 2016
- Anonymous. 2005. Purslane. Flowers of India site. http://www.flowersofindia.net/ catalog/slides/Purslane.html. Access on 30 April 2016. Verified on 2 May 2016
- Anonymous. 2015. Soil moisture. An Agri Info site. http://www.agriinfo.in/?page=topic id=274. Access on 30 April 2016. Verified on 2 May 2016
- Anonymous. 2016. Transpiration: The Water Cycle. A U.S Geological Survey site. http://water.usgs.gov/edu/watercycletranspiration.html. Access on 30 April 2016. Verified on 7 May 2016
- Aslam, M., Zamir, M. S. I., Afzal, I., Mubeen, M., and Shoaib, A. 2013. Drought Stress, Its Effect on Maize Production and Development of Drought Tolerance through Potassium Application. *Cercetări Agronomice în Moldova* **2(154)**: 99 – 114



- Assaha, D. V., Liu, L., Ueda, A., Nagaoka, T., and Saneoka H. 2016. Effects of Drought Stress on Growth, Solute Accumulation and Membrane Stability of Leafy Vegetable, Huckleberry (*Solanum scabrum* Mill.). *Journal of Environmental Biology* **37(1)**: 107-14
- Biochar Now. 2016. History. A Biochar now site. http://www.biocharnow.com/index.ph /biochar/history. Access on 6 March 2016. Verified on 20 March 2016
- Blair, K. 2014. *The Wild Wisdom of Weeds: 13 Essential Plants for Human Survival.* Vermont: Chelsea Green Publishing
- Brennan, A., Jiménez, E. M., Puschenreiter, M., Alburquerque, J. A. and Switzer, C. 2014. Effects of Biochar Amendment on Root Traits and Contaminant Availability of Maize Plants in a Copper and Arsenic Impacted Soil. *Plant Soil* 379: 351–360
- CABI. 2015. *Portulaca oleracea* (purslane). A Centre for Agriculture and Bioscience International site. http://www.cabi.org/isc/datasheet/43609. Access on 6 March 2016. Verified on 20 March 2016
- Chan, K. Y., Van Zwieten, I., Meszaros, I., Downie, A., and Joseph, S. 2008. Agronomic Values of Greenwaste Biochar as a Soil Amendment. *Soil Research* **45(8)**: 629-634
- Colorado State University Extension. 2015. Soil pH and Plant Growth. Colorado State University Extension site. http://www.ext.colostate.edu/mg/Gardennotes/222. html. Access on 6 March 2016. Verified on 20 March 2016
- Dobermann A, Fairhurst T. 2000. *Rice: Nutrient disorders & nutrient management. Handbook series.* Canada: Potash & Phosphate Institute (PPI), Potash & Phosphate Institute of Canada (PPIC) and International Rice Research Institute
- Ehrlich, S. D. 2015. Omega-3 fatty acids. A University of Maryland Medical Center site. http://umm.edu/health/medical/altmed/supplement/omega3-fatty-acids. Access on 6 March 2016. Verified on 20 March 2016
- Ejieji, C. J. and Adeniran, K. A. 2010. Effects of Water and Fertilizer Stress on the Yield, Fresh and Dry Matter Production of Grain Amaranth (*Amaranthus cruentus*). *Australian Journal of Agricultural Engineering* **1(1)**: 18-24
- Fageria N. K. and Moreira, A. 2012. The Role of Mineral Nutrition on Root Growth of Crop Plants. In Donald L. Sparks, editor: *Advances in Agronomy*, Vol. 110, Burlington: Academic Press
- Glaser, B., Haumaier, L., Guggenberger, G., and Zech, W. 2001. The 'Terra Preta' Phenomenon: a Model for Sustainable Agriculture in the Humid Tropics. *Naturwissenschaften* **88**: 37–41
- Gravel, V., Dorais, M. and Claudine, M. 2013. Organic Potted Plants Amended with Biochar: Its Effect on Growth and Pythium Colonization. *Canadian Journal of Plant Science* **93(6)**: 1217-1227



- Hajibabaee, M. Azizi, F. and Zargari, K. 2012. Effect of Drought Stress on Some Morphological, Physiological and Agronomic Traits in Various Foliage Corn Hybrids. *American-Eurasian Journalof Agricultural & Environmental Science* 12 (7): 890-896
- Howard, T. 2014. The Effect of Biochar on the Root Development of Corn and Soybeans in Minnesota Soil and Sand. International Biochar Initiative site. http://www.biochar-international.org/teachers/schools. Access on 30 April 2016. Verified on 7 May 2016
- Huang, B. 2016. Grass roots. Gorund Maintenance site. http://www.groundsmag.com/mag/grounds_maintenance_grass_roots_2/. Access on 30 October 2016. Verfified on 28 November 2016
- Igalavithana, A. D., Ok, Y. S., Usman, A. R. A., Al-Wabel, M. I., Oleszczuk, P., and Lee, S. S. 2016. *Agricultural and Environmental Applications of Biochar: Advances and Barriers.* Wisconsin: Soil Science Society of America, Inc.
- International Biochar Initiative. 2015. Environmental Benefits of Biochar. A International Biochar Initiative site. http://www.biochar-international.org/biocha r/benefits. Access on 30 April 2016. Verified on 7 May 2016
- Ismail, M. M. R. and Hasegawa, H. 2012. Water Stress. Rijeka: InTech
- Jackson, M. B. 2013. The Impact of Flooding Stress on Plants and Crops. A Plant Stress site. www.plantstress.com/articles/waterlogging_i/waterlog_i.htm. Access on 1 May 2016. Verified on 5 May 2016
- Jones, D. L., Edwards-Jones, G. and Murphy, D. V. 2011. Biochar Mediated Alterations in Herbicide Breakdown and Leaching in Soil. *Soil Biology Biochemistry* **43**: 804–813
- Jonhston, A. M. 2002. Optimizing Plant Nutrition to Minimize Crop Stress. AgriBriefs Agronomic News Items. Spring, n.2. http://www.ppippic.org/ppiweb/agbrief.nsf/5a4b8be72a35cd46852568d9001a1 8da/de1044fdc616a11e85256b9600602f0c!OpenDocument. Access on 30 October 2016. Verified on 8 November 2016
- Kamal Uddin, M., Abdul, S. J., M. Sabir Hossain, M. A. U. Nahar, M. E. Ali, and M. M. Rahman. 2014. Purslane Weed (*Portulaca oleracea*): A Prospective Plant Source of Nutrition, Omega-3 Fatty Acid, and Antioxidant Attributes. *The Scientific World Journal*. http://dx.doi.org/10.1155/2014/95101
- Leach, K. A., Hejlek, L. G., Hearne L. B., Ngyuen H. T., Sharp, R. E., and Davis, G. L. 2011. Primary Root Elongation Rate and Abscisic Acid Levels on Maize in Response to Water Stress. *Crop Science* **51(1)**: 157-172
- Lehmann, J. and Joseph, S., 2009. *Biochar for environmental management: an introduction.* London: Earthscan Publishers



UNIVERSITI MALAYSIA

- Melgren, S. 2010. Edible Weeds 101: The Health Benefits of Purslane. A Mother Earth Living site. http://www.motherearthliving.com/natural-health/edible-weeds-101 -health-benefits-of-purslane.aspx. Access on 1 May 2016. Verified on 5 May 2016
- McLauglin, H. 2010. How Biohar Helps the Soil. https://www.thebiocharrevolution.com/. Access on 30 April 2016. Verified on 3 May 2016
- McLean, E. O. 1982. Soil pH and lime requirement. In Page, A. L., R. H. Miller and D. R. Keeney (Eds.) *Methods of soil analysis. Part 2 Chemical and microbiological properties. (2nd Ed.*). Wisconsin: Soil Science Society of America
- Mitich, L. W. 1997. Common purslane (*Portulaca oleracea*). Weed Technology **11(2)**: 394-397
- Murphy, J. and Riley, J. P. 1962. A Modified Single Solution Method for the Determination of Phosphate in Natural Waters. *Analytica Chimica Acta* **27**: 31–36
- Nan, R., Carman, J. G. and Salisbury, F. B. 2002. Water Stress, CO₂ and Photoperiod Influence Hormone Levels in Whea. *Journal of Plant Physiology* **159(3)**: 307-312
- Nejad, T. S. 2011. Effect of Drought Stress on Shoot/Root Ratio. World Academy of Science, Engineering and Technology **57**: 539-541
- Oscar, F. R. J and Pauric, C. M. G. 2012. *The 2012 Drought Considerations in Soil Fertility*. Agronomy Facts 3. October 15, 2012
- Ouyang, L., Wang, F., Tang, J., Yu, L., and Zhang, R. 2013. Effects of Biochar Amendment on Soil Aggregates and Hydraulic Properties. *Journal of Soil Science and Plant Nutrition* **13(4)**: 991-1002
- Prendergast-Miller, M. T., Duvalla, M., and Sohia, S. P. 2014. Biochar–root Interactions are Mediated by Biochar Nutrient Content and Impacts on Soil Nutrient Availability. *European Journal of Soil Science* **65**: 173-185
- Proctor, C. 2013. *Biology and Control of Common Purslane (Portulaca oleracea L.)* Dissertation for Degree of Doctor of Philosophy. University of Nebraska
- Pudasaini K., Ashwath N., Walsh, K. and Bhattarai, T. 2012. Biochar Improves Plant Growth and Reduces Nutrient Leaching in Red Clay Loam and Sandy Loam. *Hydro nepal.* 86-90
- Ransom, J. 2013. Impacts Of Flooding/Waterlogging On Crop Development. A North Dakota State University site. https://www.ag.ndsu.edu/cpr/plant-science/impact s-of-flooding-waterlogging-on-crop-development-07-03-13. Access on 30 April 2016. Verified on 2 May 2016



- Rashidi, M. and Seyfi, K. 2007. Effect of Water Stress on Crop Yield and Yield Components of Cantaloupe. *International Journal of Agriculture & Biology* **9(2)**: 271-273
- Read, P. 2009. This Gift of Nature is the Best Way to Save Us From Climate Catastrophe. The Guardian site. http://www.theguardian.com/commentisfree/2009/mar/27/biochar. Access on 30 April 2016. Verified on 2 May 2016
- Rodgers, A. I. 2013. Drought. A National Geographic site. http://education.nationalgeo graphic.org/encyclopedia/drought/. Access on 30 April 2016. Verified on 7 May 2016
- Samy, J., Sugumaran, M. and Lee, K. L. W. 2005. *Herbs of Malaysia: An Introduction to the medicinal, culinary, aromatic and cosmetic use of herbs*. Kuala Lumpur: Times Edition
- Shan, L., Yang, C., Li, Y., Duan, Y., Geng, D., Li, Z., Rong, Z., Duan, G., and Zhigunov,
 A. V. 2015. Effects of Drought Stress on Root Physiological Traits and Root
 Biomass Allocation of *Reaumuria soongorica*. Acta Ecologica Sinica 35(2015): 155-159
- Siriamornpun, S. and Suttajit, M. 2010. Microchemical Components and Antioxidant Activity of Different Morphological Parts of Thai Wild Purslane (*Portulaca oleracea*). Weed Science **58(3)**: 182-188
- Smebye, A., Allinga, V., Vogtb, R. D., Gadmarb, T. C., Mulderc, J., Cornelissen, G., and Hale, S. E. 2015. Biochar Amendment to Soil Changes Dissolved Organic Matter Content and Composition. *Chemosphere* **142**: 100-105
- Sprague, D. 2015. The Impact of Too Much Water on plants. The Vindicator site. http://www.vindy.com/news/2015/aug/13/the-impact-of-too-much-water-onplants/?print. Access on 30 April 2016. Verified on 2 May 2016
- Stauffer, B., Carle N., and Spuhler, D. 2011. Soil Amendment. A Sustainable Sanitation and Water Management site. http://www.sswm.info/content/soil-amendment. Access on 30 April 2016. Verified on 7 May 2016
- Tahar, B., Abdellah A., Abdulkhaliq, A. A. S., Ali, M. A. 2010. Effect of Water Stress on Growth and Water Use Efficiency (WUE) of Some Wheat Cultivars (*Triticum durum*) grown in Saudi Arabia. *Journal of Taibah University for Science* 3: 39-48
- Ulyett, J., Sakrabani, R., Kibblewhite, M. and Hann, M. 2014. Impact of Biochar Addition on Water Retention, Nitrification and Carbon Dioxide Evolution from Two Sandy Loam Soils. *European Journal of Soil Science* **65**: 96–104
- USDA. 2006. USDA National Food and Nutrient Analysis Program Wave 10j. Beltsville: MD
- Uzoma, K. C., Inoue, M., Andry, H., Fujimaki, H., A. Zahoor and Nishihara, E. 2010. Effect of Cow Manure Biochar on Maize Productivity under Sandy Soil Condition. *Soil Use and Management* **27**: 205–212



- Varela, O. M., Eva, B. R., Huang, W. J., Chien, C. C., and Wang, Y. M. 2013. Agronomic Properties and Characterization of Rice Husk and Wood Biochars and Their Effect on the Growth of Water Spinach in a Field Test. *Journal of Soil Science and Plant Nutrition* **13(2)**: 251-266
- Vinall, K. and Watt, M. 2013. Using WINRhizo and Photoshop to Determine Root Length, Diameter and Branching. A Prometheus Wiki site. http://www.publish.csiro.au/prometheuswiki/tiki-pagehistory.php?page=Using WINRhizo and Photoshop to determine root length, diameter and branching&preview=14. Access on 30 April 2016. Verified on 2 May 2016
- Wang, C., Yang, A., Yin, H., and Zhang, J. 2008. Influence of Water Stress on Endogenous Hormone Contents and Cell Damage of Maize Seedlings. *Journal of Integrative Plant Biology* **50(4)**: 427-434
- Widowati, Utomo, W. H., Guritno, L. A. and Soehono, L. A. 2012. The Effect of Biochar on the Growth and N Fertilizer Requirement of Maize (*Zea mays* L.) in Green House Experiment. *Journal of Agricultural Science* **4(5)**: 255-262
- Xu, W., Cui, K., Xu, A., Nie, L., and Peng, S. 2015. Impact of Biochar on the Water Holding Capacity of Loamy Sand Soil. Drought Stress Condition Increases Root to Shoot Ratio via Alteration of Carbohydrate Partitioning and Enzymatic Activity in Rice Seedlings. *Acta Physiologiae Plantarum* **37(9)**: 2-11
- Yu, O. Y., Raichle, B., and Sink, S. 2013. Impact of Biochar on the Water Holding Capacity of Loamy Sand Soil. *International Journal of Energy and Environmental Engineering* **4(44)**: 1-9
- Zheng W., Sharma, B. K., and N. Rajagopalan. 2010. Using biochar as a Soil Amendment for Sustainable Agriculture. An Illinois Sustainable Technology Center site. http://istc.illinois.edu/research/biochar/soil_amendment/sustainabl e_agriculture.cfm. Access on 6 March 2016. Verified on 20 March 2016
- Zulkifli, E. 2016. Memahami Pola Hujan dan Pengaruh Efek Kekeringan Terhadap Produksi Kelapa Sawit. Impact of biochar on The Water Holding Capacity of Loamy Sand Soil. https://www.linkedin.com/pulse/memahami-pola-hujan-danpengaruh-efek-kekeringan-kelapa-eko-zulkifli. Access on 30 October 2016. Verfified on 28 November 2016

