

BORANG PENGESAHAN STATUS TESIS@

JUDUL: Extraction & Screening of Antioxidants in
Metroxylon Sagu

Ijazah: Sarjana Muda Sains Bioteknologi Dan Keperluan (Hons)

SESI PENGAJIAN: 2001

Saya NOR IDAYU BT. MHD TAHIR

(HURUF BESAR)

mengaku membenarkan tesis (LPS/Sarjana/Doktor Falsafah)* ini disimpan di Perpustakaan Universiti Malaysia Sabah dengan syarat-syarat kegunaan seperti berikut:

1. Tesis adalah hakmilik Universiti Malaysia Sabah.
2. Perpustakaan Universiti Malaysia Sabah dibenarkan membuat salinan untuk tujuan pengajian sahaja.
3. Perpustakaan dibenarkan membuat salinan tesis ini sebagai bahan pertukaran antara institusi pengajian tinggi.
4. **Sila tandakan (/)

SULIT

(Mengandungi maklumat yang berdarjah keselamatan atau kepentingan Malaysia seperti yang termaktub di dalam AKTA RAHSIA RASMI 1972)

TERHAD

(Mengandungi maklumat TERHAD yang telah ditentukan oleh organisasi/badan di mana penyelidikan dijalankan)

TIDAK TERHAD

Disahkan oleh

[Signature]
(TANDATANGAN PENULIS)

[Signature]
(TANDATANGAN PUSTAKAWAN)

Alamat Tetap: _____

PROF. DR. PERUMAL RAMASAMY

Nama Penyalia

Tarikh: 12 MAC 2003

Tarikh: _____

CATATAN: * Potong yang tidak berkenaan.

** Jika tesis ini SULIT atau TERHAD, sila lampirkan surat daripada pihak berkuasa/organisasi berkenaan dengan menyatakan sekali sebab dan tempoh tesis ini perlu dikelaskan sebagai SULIT dan TERHAD.

@ Tesis dimaksudkan sebagai tesis bagi Ijazah Doktor Falsafah dan Sarjana secara penyelidikan, atau disertasi bagi pengajian secara kerja kursus dan penyelidikan, atau Laporan Projek Sarjana Muda (LPSM).



DECLARATION

EXTRACTION AND SCREENING OF ANTIOXIDANTS IN *Metroxylon sagu*

NOOR IDAYU MHD TAHIR

**THIS THESIS IS SUBMITTED TO FULFILL THE REQUIREMENT IN
OBTAINING THE DEGREE OF SCIENCE BACHELOR WITH HONOURS**

**BIOTECHNOLOGY PROGRAMME
SCHOOL OF SCIENCE & TECHNOLOGY
UNIVERSITI MALAYSIA SABAH**

March 2004

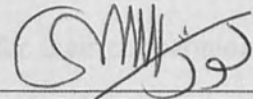


UMS
UNIVERSITI MALAYSIA SABAH

DECLARATION

I, hereby declare that this thesis is of my own work and effort, except the quotations, excerpts and facts, for which each source has been clearly stated.

March 12, 2004



NOOR IDAYU BT MHD TAHIR

HS2001-3040



ACKNOWLEDGEMENT

A special thank is extended to Professor Dr. Perumal Ramasamy JSM, who had provided a significant contribution by giving all the advices, guidance, support and knowledge needed throughout my work and learning processes. To Mr. Chong Tong Seng, Ms. Hew Chaw Sen, Mr. Foo Sek Hin, Ms. Ainol Azifah M. Faik, Ms. Nurulhusna Ab. Hamid and all the master students, for their support in completing this project, providing helpful advices, consultant and aid unreluctantly, day and night. To En. Taib b. Mawang of Kg. Kuala Balik, Beaufort who had kindly provided all the raw source of sago and its by-products, Jabatan Pertanian Beaufort and Miss Linda Hassan, for their cooperation and contribution. Not forgetting to all other parties who had contributed to the completion of this project. Last but not least, I dedicate this final project to my dearest family, and to all the lecturers and my fellow course mates; my Biotechnology family here in Universiti Malaysia Sabah.



VERIFICATION**VERIFIED BY**

Signature

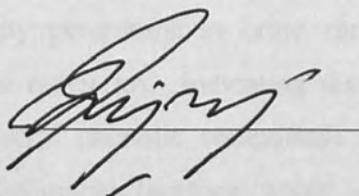
1. SUPERVISOR

(Prof. Dr. Perumal Ramasamy)



2. EXAMINER 1

(Dr. Charles S. Vairappan)



3. EXAMINER 2

(Dr. Zaleha Abd. Aziz)



4. DEAN

(Assoc. Prof. Dr. Amran Ahmed)



ABSTRACT

Antioxidants were extracted from sago pith, sago processing effluent and waste sago pith obtained from a small-scale sago processing site in Beaufort with methanol to be further studied with phytochemical and biological activity tests while hexane was used to extract vitamin A and its precursors. The extracts were submitted to four phytochemical tests which detected flavonoids, tannin and phenolic compounds in the extracts. The concentration of flavonoids was contained highest in sago pith, followed by its waste water and the waste pith. Tannin presence was observed highest in the processing waste water. Biological activities of the extracts were assessed through linoleic acid autoxidation test, peroxidation value determination test and brine shrimp lethality test. Sago pith showed the longest induction period of 5 days which indicates the highest antioxidant activity. The peroxidation value determination test showed that extract of waste water possess the highest capability of preserving the unsaturated fat from oxidizing, followed by sago pith extract and waste pith extract. Mortality percentage in brine shrimps incubated with different concentration of extracts was rather low, indicating that the extracts do not cause lethality in brine shrimps. Several phenolic compounds were observed after running silica gel 60 F₂₅₄ column chromatography (acetone: water: acetic acid, 75:20:5, v/v) with subsequent thin layer chromatography (n-propanol:water, 64:36, v/v). Vitamin C was determined at 264 nm by using a C18 reversed-phase column HPLC with a mobile phase of acetonitrile: water (75:25, v/v) in which the highest concentration was observed in waste water, followed by sago pith and waste pith. Carotene was detected at 450 nm using UV/Vis spectrophotometer where sago pith contain the highest concentration, followed by waste pith and waste water extract. Sago pith and its by-products (wastes) contain antioxidants and are non-toxic.

ABSTRAK

Bahan-bahan antioksidan diekstrak dari pokok sagu, sisa cecair dan sisa pepejal pemprosesan sagu yang diperolehi dari tapak pemprosesan sagu kecil-kecilan di Beaufort. Pengekstrakan dilakukan menggunakan metanol untuk analisis fitokimia, aktiviti biologi dan kromatografi. Heksana pula digunakan untuk mengekstrak vitamin A dan prekursoranya. Analisis fitokimia yang dijalankan ke atas ekstrak menunjukkan ia mengandungi flavonoid, tanin dan bahan-bahan fenolik. Kepekatan flavonoid adalah tertinggi di dalam ekstrak pokok sagu, diikuti dengan sisa cecair dan seterusnya sisa pepejal pemprosesan. Kandungan tanin tertinggi di dalam ekstrak sisa cecair pemprosesan. Aktiviti biologi ekstrak-ekstrak dikaji melalui ujian autoksidasi asid linoleik, ujian penentuan nilai peroksida dan ujian kemautan udang laut. Ekstrak pokok sagu memberikan tempoh induksi terpanjang iaitu 5 hari di mana aktiviti antioksidannya bertahan paling lama. Ujian penentuan nilai peroksida menunjukkan ekstrak sisa cecair mempunyai kebolehan tertinggi untuk mengelakkan pengoksidaan lemak tak tepu (asid linoleik), diikuti oleh ekstrak pokok sagu dan sisa pepejal. Peratus kemautan udang laut adalah rendah sama sekali di mana lebih tinggi kepekatan ekstrak yang digunakan, lebih tinggi peratus udang yang hidup. Kromatografi Lapisan Nipis (TLC) menggunakan gel silica 60 F₂₅₄ dengan fasa mobil n-propanol: air (64:36, v/v) selepas penulenan dengan kromatografi turus gel silica 60 F₂₅₄ dengan fasa mobil acetone: air: asid asetik (75:20:5, v/v) memperlihatkan kehadiran bahan-bahan fenolik dalam ekstrak-ekstrak. Vitamin C dapat dikesan pada 254 nm menggunakan Kromatografi Cecair Berkeupayaan Tinggi (HPLC) kolum C18 fasa berbalik dengan fasa mobil acetonitril: air (75:25, v/v) di mana kepekatan tertinggi terkandung dalam ekstrak sisa cecair pemprosesan, diikuti oleh ekstrak pokok sagu dan sisa pepejal. Karoten dikesan pada 450 nm menggunakan spektrofotometer UV/Vis di mana ekstrak pokok sagu mempunyai kepekatan tertinggi diikuti dengan sisa pepejal dan sisa cecair pemprosesan. Sagu dan bahan buangnya mempunyai bahan antioksidan dan tidak toksik.



CONTENTS

DECLARATION	ii
VERIFICATION	iii
ACKNOWLEDGEMENT	iv
ABSTRACT	v
ABSTRAK	vi
CONTENTS	vii
LIST OF TABLES	x
LIST OF FIGURES	xi
LIST OF PICTURES	xii
LIST OF SYMBOLS, UNITS AND ABBREVIATIONS	xiii
CHAPTER 1 INTRODUCTION	
1.1 Free Radicals and Antioxidants	1
1.2 <i>Metroxylon sagu</i>	4
1.3 Objectives of the Project	7
CHAPTER 2 LITERATURE REVIEW	
2.1 Sagu as a Source of Food	8
2.2 Antioxidant for the body	12
2.3 Antioxidant Studied in <i>Metroxylon sagu</i>	15
2.3.1 Vitamin A	15
2.3.2 Vitamin C	15
2.3.3 Phenolic Compounds	17
2.3.3 (a) Vitamin E	18
2.3.3 (b) Flavonoids	20
2.3.3 (c) Tannin	24
2.4 Measurements of Antioxidant System Component	26



CHAPTER 3 METHODOLOGY

3.1 Chemicals and Reagents	30
3.2 Crude Extract Preparation and Screening List	30
A) Flavonoid Screening (Wilstatter-Sianidin Test and Bate-Smith and Metcalf Test) Description	33
B) Tannin and Polyphenol Compound Screening Description	34
C) Linoleic Acid Autooxidation Test Description	34
D) Peroxidation Value Determination Test Description	35
E) Brine Shrimp Lethality Test Description	35
F) Column Chromatography Description	36
G) Thin Layer Chromatography Description	37
H) High Performance Liquid Chromatography Description	38
I) Carotene Spectrophotometry	39

CHAPTER 4 RESULTS AND DISCUSSIONS

4.1 Sample preparation and extraction	54
4.2 Screening Tests	
a) Flavonoid screening i) <i>Wilstatter-Sianidin Test</i>	41
ii) <i>Bate-Smith and Metcalf Test</i>	42
b) Tannin and polyphenol compound screening i) <i>Gelatin Test</i>	43
ii) <i>Polyphenol Test</i>	44
c) Linoleic Acid Autooxidation Test	44
d) Determination of Peroxidation Value	49
e) Brine Shrimp Lethality Test	51
f) Column Chromatography	53
g) Thin Layer Chromatography	53
h) High Performance Liquid Chromatography	56
i) Carotene Spectrophotometry	56



CHAPTER 5 CONCLUSIONS	58
REFERENCES	61
APPENDIX	
Appendix A	65
Appendix B	66
Appendix C	68
Appendix D	69
Appendix E	70
Appendix F	71
Appendix G	76
Table 3.1. Colour Changes and ΔE Included	33
Table 3.2. Surface and Thermal Characterization	34
Table 3.3. Composites Data in FTIR for WPMMA, C, and E Nanoparticles	35
Table 4.1. Weight Compositions of composites	39
Table 4.2. Results of Mechanical Analysis and Tensile Mechanical Data	41
Table 4.3. Comparison of Mechanical Properties of Test	43
Table 4.4. Absorbance of 200 nm and 400 nm wavelengths	46
Table 4.7. The Value of Absorbance Maximum of WPMMA	47
Table 4.8. Molecular Weight Characterization of WPMMA	49
Table 4.9. The mechanical properties of composites	49
Table 4.10. Molecular Weight of WPMMA in Different Compositions of WPMMA	51
Table 4.11. Percentage of WPMMA in WPMMA-MA and WPMMA	52
Table 4.12. Molecular Weight of WPMMA in WPMMA-MA and WPMMA	53
Table 4.13. Comparison Characterization	54



LIST OF TABLES

Table 1.1: Table of Palm Cultivation Area in Malaysia for the Year 1986	5
Table 2.1: Types of Antioxidant in Plant -Food Source	12
Table 2.2: Major Classes of Phytochemicals	13
Table 2.3: Functions of Ascorbic Acid	16
Table 2.4: Flavonoid Groups and Its Food Source	23
Table 2.5: Common Solvent System for Thin Layer Chromatography (TLC)	28
Table 3.1: Colour Changes and Its Indication	33
Table 3.2: Conclusion of Possible Observations	34
Table 3.3: Conditions Used to Run the HPLC for Vitamin A, C and E Screening	39
Table 4.1: Weight Composition of Samples	40
Table 4.2: Results of Wilstatter-Sianidin Bate-Smith and Metcalf Test	41
Table 4.3: Observation for Gelatin and Polyphenol Tests	43
Table 4.4: Absorbance at 500 nm versus Day of Incubation	44
Table 4.5: The Value of Antioxidant Activity Relative to BHT	47
Table 4.6: Volume of Sodium Thiosulphate Titrated	49
Table 4.7: Peroxidation Value (PV) of Samples	49
Table 4.8: Mortality Rate of Brine Shrimp in Different Concentration of Extracts	51
Table 4.9: Percentage of Brine Shrimp Survival after 24 and 48 hours	52
Table 4.10: Absorbance Reading for Carotene Detection at 450 nm	57
Table 4.11: Carotene Concentration	57



LIST OF FIGURES

Figure 2.1 (a, b, c, d and e): Synthetic Antioxidants Used as Food Additives	14
Figure 2.2: Ascorbic Acid Oxidation	17
Figure 2.3 (a): Structures of Tocopherols	19
Figure 2.3 (b): Structures of Tocotrienols	19
Figure 2.4: Vitamin C ‘Sacrificing’	20
Figure 2.5: Basic Structure of Flavonoids	21
Figure 2.6 (a): Chemical Structures of Flavonols and Flavones	22
Figure 2.6 (b): Chemical Structure of Flavanone	22
Figure 2.7: Structures of Typical Condensed and Hydrolysable Tannins	25
Figure 4.1: Graph of Absorbance at 500 nm versus Day Reading is Taken	46
Figure 4.2: Lipid Peroxidation Assay	47
Figure 4.3: General Structure of Hydroxycinnamic Acids	55



LISTS OF PICTURES

Picture 1.1: Spiny type of sago palm	4
Picture 1.2: Non-spiny type of sago palm	4
Picture 1.3 (a): A sago palm ready to be harvested	6
Picture 1.3 (b): Sago flour to be collected and dried	6
Picture 1.3 (c): Waste sago pith	6
Picture 1.3 (d): Sago processing effluent	6
Picture 2.1: The pith of sago palm	9
Picture 2.2: The pith inserted into the inlet of the macerating machine	9
Picture 2.3: Traditional mechanical cutter	10
Picture 2.4: Stepping platform	10
Picture 2.5: Sago flour sold in Beaufort and Kuala Penyu, Sabah	11
Picture 3.1: Solvent extraction of sago pith	31
Picture 3.2: Solvent extraction of waste sago pith	31
Picture 3.3: Sampling of sago processing effluent	31
Picture 3.4: The silica gel 60 F ₂₅₄ column prepared	37
Picture 4.1: Example of TLC plate chromatographed viewed under UV light	54

LIST OF SYMBOLS, UNITS AND ABBREVIATIONS

HO \cdot	hydroxyl radicals
O $_2^{\cdot-}$	superoxide ion
NO	nitric oxide
O \cdot	singlet oxygen
ROO \cdot	peroxyl radicals
HO $_2\cdot$	hydroperoxyl radical
DNA	deoxyribonucleic acid
RNA	ribonucleic acid
BHA	butylated hydroxyanisole
BHT	butylated hydroxytoluene
TBHQ	tertiary butylated hydroquinone
PG	propyl gallate
cm	centimetre
GSH	glutathione
Q $_{10}$	coenzyme Q
SOD	superoxide dismutase
LDL	low density lipoprotein
CH $_3$	methyl group
AMP	adenosine monophosphate
UV/Vis	Ultra Violet/Visible
RDA	Recommended Dietary Allowance
HPLC	High Performance Liquid Chromatography
PV	Peroxidation Value
TLC	Thin Layer Chromatography
H $_2$ O	water
NH $_3$	ammonia
g	gram
$^{\circ}$ C	degree Celcius
ml	mililitre
%	percentage
FeCl $_3$	ferum chloride (III)
nm	nanometre
N	Normality
ppm	part per million
DMSO	dimethyl sulphoxide
(w/v)	weight upon volume
HCl	hydrochloride acid
NaCl	natrium chloride
Abs $_{500}$	absorbance at 500 nm
Fe $^{2+}$	ferrous ion
Fe $^{3+}$	ferric ion
SCN $^-$	thiocyanate ion
Fe(SCN) $_6^{3-}$	ferum hexacyanoferat ion
Na $_2$ S $_2$ O $_3$	sodium thiosulphate
Abs $_{450}$	absorbance at 450 nm



CHAPTER 1

INTRODUCTION

1.1 Free radicals and antioxidant

Free radicals are atoms or group of atoms with odd electron number or unpaired electron that are produced after oxygen react with certain molecules (by-products of oxygen metabolism). When these reactive free radicals are formed, it will begin a chain reaction like a domino effect, resulting more destructive reaction. Free radicals promote oxidation that result in high energy and kills bacterial invaders by stealing electron from the nearest stable mode. However, danger comes when they react with main cellular component such as DNA or the cell membrane. Cell function will be disrupted or the cell can even die. Reactive radical species such as hydroxyl radicals ($\text{HO}\cdot$), superoxide ion ($\text{O}_2^{\cdot-}$), nitric oxide (NO), singlet oxygen (O^1), peroxy radicals ($\text{ROO}\cdot$) and hydroperoxyl radical ($\text{HO}_2\cdot$) need to be controlled and prevented from damaging the lipids, proteins and other complexes, for examples; unsaturated fatty acid, hyaluronic acid, DNA and RNA. These free radicals are also associated with diseases of ageing process.



To avoid the damaging effect of free radicals, human body gets protection from antioxidant defence system. Scientific research has shown that antioxidant has roles in protecting human from various diseases. There are plenty of antioxidant resources that can be consumed to decrease the action of free radicals against human such as carrots, tomatoes, yam, green vegetables, blueberries, ginkgo biloba, garlic, green tea and etcetera.

Antioxidant can be defined as substance that slow down or even stop oxidative damage on attacked molecules by free radicals. Oxidative damage is generally known as effects of free radicals reaction onto biological molecules. Antioxidant protects the body by removing the catalysts for example the enzyme in food oxidation or substituting the component or compound that are oxidize-prone or even repairing the damage made by free radicals.

Vitamin A, vitamin C, vitamin E, flavonoids, carotenoids, and tannins are some of the natural occurring antioxidant, just to name a few. While examples of synthetic antioxidant are butylated hydroxyanisole (BHA), butylated hydroxytoluene (BHT), tertiary butylated hydroquinone (TBHQ) and propyl gallate (PG) which are commercially used as food additives.

Vitamin A is fat-soluble vitamin which is found in animal product such as meat and milk. It can also be made by our body by its precursor, β -carotene. As an antioxidant, vitamin A protects cell membrane and fat tissue, helps to repair damage of the body by pollutants in air and increase body defence system.

Vitamin C was firstly renowned in 1970s when a scientist that won the Nobel Prize, Linus Pauling recommended a high daily dosage (8-10 gram) of vitamin C to avoid or to treat cold. As an antioxidant, vitamin C helps to fight against cardiovascular diseases by protecting the artery track from oxidative damage. There are also research that state vitamin C as a body protection from cigarettes smoke and other air pollutants.

Vitamin E is an effective chain breaking antioxidant in protecting poly-unsaturated lipid from harmed by free radicals. However, it interact better if taken along with other antioxidant especially vitamin C, selenium and β -carotene. Vitamin E can also increase cardiovascular health. It acts as vasodilator that expands the vein and prevents the hardening of arteries and blood clotting.

Bioflavonoids consist of 5000 substances that react as antioxidant. It can be found in fruits, vegetables, green tea, soy products, herbs and spices. Bioflavonoid activities are more effective if combined with vitamin C. this combination is used in allergy treatment, diabetes, hypertension and virus infection. One of the many examples in the flavonoid family is procyanidins that helps in regenerating other antioxidants that neutralize free radicals in the blood and body tissue.

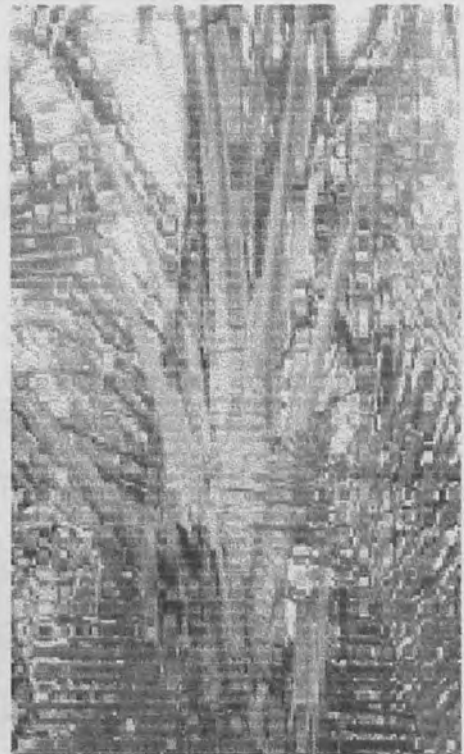


1.2 *Metroxylon sagu*

Sago palm is known as 'rumbia' plant amongst locals. It is said to be originated from Papua New Guinea. Sago palm are planted widely especially in Sarawak in swampy area near rice fields and settlement area. It has a few local names like 'mulong' for the Ibans and Malays, 'dulo' (for the non-spiny types) and 'balau' (for the spiny types) for the Kenyah people.



Picture 1.1: Spiny type of sago palm



Picture 1.2: Non-spiny type of sago palm

Sago palm has various usages. Its leaves are made into house thatches, hats, baskets and blowpipe darts. When the plant reaches twelve years old, the sticky sap is used as gum and starches. Ripe fruits and palm cabbage are eaten. There are also the

Rhynchophorus schach larvae (sago grubs) which live in the trunks of palms that have been cut down for 2-3 days. These grubs are edible and highly prized by the locals.

Sago palm grows in natural peat swamps with minimal drainage and more environmental friendly compared to other crops requiring complete drainage of peat basin (MARDI report in *The Planter* (78) 2002). Sago starch production is centered in the Oya and Mukah river area in Sibuan division, where the swamps extend as much as 32 km inland. In Peninsular Malaysia, sago palm planting is concentrated in Batu Pahat, Johor. Table 1.1 below shows the area of palm cultivation in Malaysia for the year 1986.

Table 1.1: Table of palm cultivation area in Malaysia for the year 1986.

(Source: Ministry of Agriculture)

Crop	Hectares under cultivation
African oil palm	1359671
Coconut palm	182776
Sago palm	647
Betel nut palm	445
Salac palm	41
Sugar palm	33

Pictures 1.3 (a), 1.3 (b), 1.3 (c) and 1.3 (d) shows a route of sago utilization into a product (sago flour) and in the same time producing a big amount of waste (solid and liquid).



Picture 1.3 (a):
A sago palm
ready to be
harvested



Picture 1.3 (b):
Sago flour to
be collected
and dried



Picture 1.3 (c): Waste sago pith



Picture 1.3 (d): Sago processing effluent



1.3 Objectives of the project

Extraction and screening of antioxidant in *Metroxylon sagu* are done to study the biochemical composition that are classified as antioxidants for examples; vitamin A, vitamin C, vitamin E, flavanoids, tannin, and other phenolic compounds qualitatively.

The main objectives of this research are;

- 1) Extracting antioxidant in *Metroxylon sagu* and its waste products.
- 2) Screening and determining the types of antioxidant in the extract of *Metroxylon sagu* and its waste products.
- 3) Studying the biological activity of antioxidant agent in the extract of *Metroxylon sagu* and its waste products.

There are two types of waste, the solid and the liquid waste. Both of these wastes are considered to possess no commercial value to the sago starch producer and farmers. In a biological point of view, these wastes can be utilized into beneficial and valuable materials and further studies on their biological properties should be carried out.



CHAPTER 2

LITERATURE REVIEW

2.1 Sago as a source of food

After ten to twelve years old, the sago palm reaches its optimum maturity and can be cut down for use. The trunk of the plant is the main part to be utilized and processed. The starch extraction rate for each section of sago trunk (90 cm) is about eighteen percent (Tan *et al.*, 2002). Sago is extracted by a crude method of crushing or macerating the pith, washing it thorough, allowing the starch to settle at the bottom and then collected, dried and stored for use.

The sago starch processing in Sabah is considered only a cottage industry where factories using modern technology imported from Europe that enables production of higher quality of sago starch are located in Sarawak. Picture 2.1 shows the trunk of a sago tree in which the pith is later processed.



Picture 2.1: The pith of sago palm

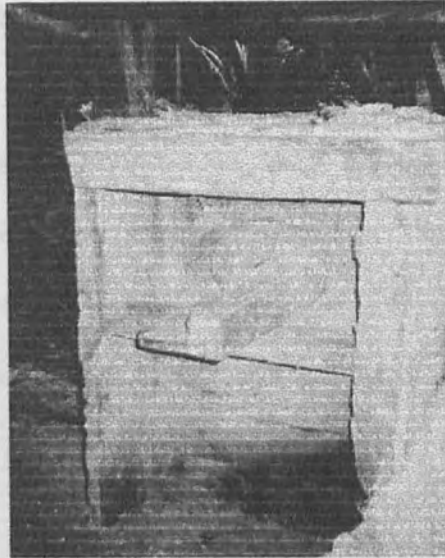
The photos below show a traditional technique used by villagers in Beaufort to process sago pith into sago flour.



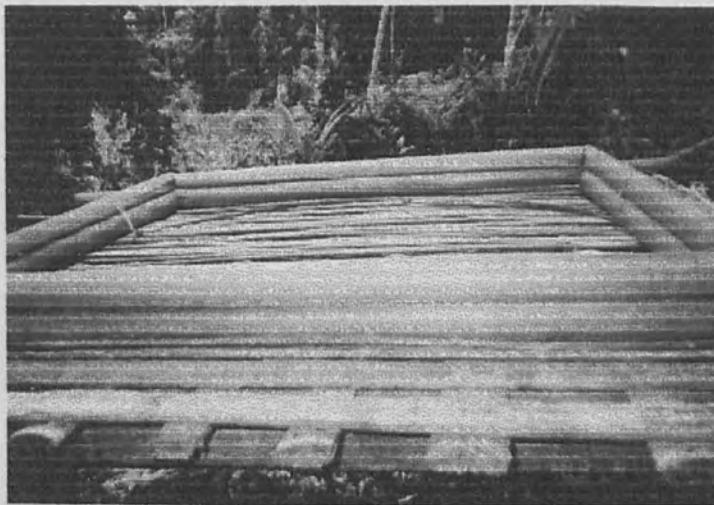
Picture 2.2: The pith is inserted into the inlet of the macerating machine.

There is another method used in small scale sago processing. The sago pith is chopped using a simple mechanical cutter shown in Picture 2.3 and then brought to a platform made of sago trunk (Picture 2.4) to be crushed to obtain the pith extract by stepping on

the blended pith. The liquid extract are collected under the platform, filtered and dried to produce starch flour.



Picture 2.3: Traditional mechanical cutter



Picture 2.4: Stepping platform

Starch and cellulose are the most commonly found carbohydrate, where carbohydrates constitute the greatest portion of food consumed in the world by man and

REFERENCES

- Arya, S. S., Natesan, V., Parihar, D. B. and Vijayaraghavan, R. K., 1979. Stability of Carotenoids in Dehydrated Carrots. *Journal of Food Technology*, **14**, 579-586.
- Bate-Smith, E. C. and Swain, T., 1962. In: Mason, H. S. and Florkin, A. M. (eds.) *Comparative Biochemistry*. **3**, 764. Academic Press, New York.
- Berger, K. G. and Hamilton, R. J., 1985. *Lipids and Oxygen: Is Rancidity Avoidable in Practice?* Blackie Academic & Professional, Glasgow.
- Bueno, M. P., 1997. Collaborative Study: Determination of Retinol and Carotene by High Performance Liquid Chromatography. *Food Chemistry*. **59** (1), 165-170. Elsevier Science Ltd.
- Chye, T. S., 1990. *Biokimia Tumbuhan Hijau*. Dewan Bahasa dan Pustaka, Kuala Lumpur.
- Day, A. J. and Williamson, G., 1999. Human Metabolism of Dietary Quercetin Glycosides. In: Gross, G. G. et al. (ed.), *Plant Polyphenols 2: Chemistry, Biology, Pharmacology, Ecology (Basic Life Science)*. **66**, 43-63. Kluwer Academic/ Plenum Publishers, New York.
- Donglin, Z., and Yasunori, H., 2003. Phenolic compounds, ascorbic acid, carotenoids and antioxidant properties of green, red and yellow bell peppers. *Food, Agriculture & Environment*. **1**(2), 22-27. WFL Publisher.
- Fasihuddin Ahmad and Hasmah Raji, 1993. *Kimia Hasil Semulajadi dan Tumbuhan Ubatan*. Dewan Bahasa dan Pustaka, Kuala Lumpur.
- Fasihuddin, B. A., Che Wan Zanariah and Che Wan Ngah, 1991. Phytochemical and Antibacterial Compound Screening in Medicinal Plants. In: *Medicinal Product from Tropical Rain Forests, Proceedings of Conference*. Forest Research Institute of Malaysia, Kuala Lumpur.
- Freedland, R. A. and Briggs S., 1980. *A Biochemical Approach to Nutrition (Outline Studies in Biology)*. Chapman and Hall Ltd., London.
- Fulcrand, H., 1999. Electrospray Contribution to Structural Analysis of Condensed Tannin Oligomers and Polymers. In: Gross, G. G. et al. (ed.), *Plant Polyphenols 2: Chemistry, Biology, Pharmacology, Ecology (Basic Life Science)* **66**, 223-245. Kluwer Academic/ Plenum Publishers, New York.



- Grace, S. C., Yamasaki, H. and Pryor, W. A., 1999. Spin Stabilizing Approach to Radical Characterization of Phenylpropanoid Antioxidants: An ESR Study of Chlorogenic Acid Oxidation in the Horseradish Peroxidase, Tyrosinase, and Ferrylmyoglobin Protein Radical Systems. In: Gross, G. G. et al. (ed.), *Plant Polyphenols 2: Chemistry, Biology, Pharmacology, Ecology (Basic Life Science)* **66**, 43-63. Kluwer Academic/ Plenum Publishers, New York.
- Gutteridge, J. M. C. and Halliwell, B., 1994. *Antioxidants in Nutrition, Health, and Disease*. Oxford University Press Inc., New York.
- Hagerman, A. E., Riedl K. M. and Rice R. E., 1999. Tannins as Biological Antioxidants. In: Gross, G. G. et al. (ed.), *Plant Polyphenols 2: Chemistry, Biology, Pharmacology, Ecology (Basic Life Science)* **66**, 495-505. Kluwer Academic/ Plenum Publishers, New York.
- Handelman, G. J. and Pryor W. A., 1999. Evaluation of Antioxidant Status in Humans. In: Papas, A. M. (ed.). *Antioxidant Status, Diet, Nutrition, and Health*. 37-57. CRC Press Inc., Boca Raton, Florida.
- Helle, J. C., Walton, C. R. and Huges, M. A., 2000. *The Flavonoids*. Chapman & Hall, London.
- Hendrich, S., Wang, G. J., Lin, H. K., Xu, X., Tew, B., Wang, H. and Murphy, P. A., 1999. Isoflavone Metabolism and Bioavailability. In: Papas, A. M. (ed.). *Antioxidant Status, Diet, Nutrition, and Health: 37-57*. CRC Press Inc., Boca Raton, Florida.
- Hernes, P. J. and Hedges, J. I., 1999. Geochemistry of Tannin: Methods and Applications. In: Gross, G. G. et al. (ed.), *Plant Polyphenols 2: Chemistry, Biology, Pharmacology, Ecology (Basic Life Science)* **66**, 43-63. Kluwer Academic/ Plenum Publishers, New York.
- How, S. E. 1989. *Penyelidikan Fitokimia dan Farmakologi Ke Atas Tumbuhan E. odoratum*. Disertasi Sarjana Muda Sains, Universiti Sains Malaysia.
- Ibanez, E. and Cifuentes, A., 2001. New Analytical Techniques in Food Science. In: Clydesdale, F. M. (ed.). *Critical Reviews in Food Science and Nutrition*. **4** (6). CRC Press LLC.
- Khachik, F., Beecher, G. R. and Smith, J. C., 1995. Lutein, lycopene, and their oxidative metabolites in chemoprevention of cancer. *Journal of Cell Biochemistry*. **22**, 236-246.
- Karlsson, J., 1997. *Antioxidants and Exercise*. Human Kinetics. Champaign, Illinois.



- Larson, R. A., 1997. *Naturally Occuring Antioxidants*. CRC Press LLC (Lewis Publishers). Boca Raton, Florida.
- Latiff, A., Zainuin, A., Petol, G. H., Ujang, S., Naemah, A. A., Rahmah, M. and Ikram, M. S., 1995. Phytochemical and Toxicity Screening of Plants from Fraser's Hill, Pahang. In: *Chemical Prospecting in the Malaysian Forest*. Pelanduk Publication (M) Sdn. Bhd., Kuala Lumpur.
- Lee, H. S., 2002. Inhibitory activity of *Cinnamomum cassia* bark-derived component against rat lens aldose reductase. *Journal of Pharmaceutical Science* (www.ualberta.ca/~csps) **5** (3), 226-230.
- Madulid D. A., 1991. Utilization of Philippine Palms. In: Johnson, D., (ed). *Palms for human needs in Asia (Palm Utilization and Conservation in India, Indonesia, Malaysia and the Philippines)*. WWF Project 3325, 251. A. A. Balkema, Rotterdam.
- Noguchi, N. and Niki, E., 1999. Chemistry of Active Species and Antioxidants. In: Papas, A. M. (ed.). *Antioxidant Status, Diet, Nutrition, and Health*: 37-57. CRC Press Inc., Boca Raton, Florida.
- Osawa, T. and Namiki, M., 1981. A Novel of Antioxidant Isolated from Leave Wax of Eucalyptus Leaves. *Agriculture Biology Chemistry* **45** (3), 735-739.
- Papas, A. M., 1999. Diet and Antioxidant Status. In: Papas, A. M. (ed.). *Antioxidant Status, Diet, Nutrition, and Health*: 89-106. CRC Press Inc., Boca Raton, Florida.
- Perez, R. M., Vargas, R., Martinez, F. J., Garcia, E. V. and Hernandez, B., 2003. Antioxidant Activity of Alkaloids From *Bocconia arborea*; A Study on Six Testing Methods. *Ars Pharmaceutica*. **44** (1), 5-21.
- Rohana, S., Vimala, S., Ilham, A. and Rashih, A. A., 2000. Antioxidant evaluation in Tongkat Ali Plant Parts. In: *International Conference on Traditional/ Complementary Medicine (Medicinal Herbs) Ensuring Safety and Efficiency Trough Research and Developme*, Kuala Lumpur.
- Ruddle, J., 1978. *Composition and Characteristics of Selected Palm Products*. <http://www.fao.org/docrep/X0451E/X0451e11.htm>
- Rumsey, S. C., Wang, Y. and Levine, M., 1999. Antioxidants and Diseases of the Eye. In: Papas, A. M. (ed.). *Antioxidant Status, Diet, Nutrition, and Health*: 37-57. CRC Press Inc., Boca Raton, Florida.
- Subramaniam, V., Mohd Ilham Adenan and Abdul Rashid Ahmad, 1999. Antioxidant in Malaysian Ulam. *Proceeding the 14th Annual Seminar of Natural Product Research*, Malaysia.



- Su, J. D., Osawa, T., Kawakishi S. and Namiki, M., (1988). Tannin Antioxidants from *Osbeckia Chinensis*. *Phytochemistry*. **27** (5), 1315-1319. Pergamon Press Publication.
- Takayuki, S., Yoshihide, H., Hideaki, H. and Toshihiko, O., 1994. Flavonoid with Strong Antioxidative Activity Isolated from Young Barley Leaves. In: Ho, C. T., Toshihiko, O., Huang, M. T. & Robert T. R. *Food Phytochemistry for Cancer Prevention II: Teas, Spices, and Herbs*: 154-161. American Journal Society, Washington.
- Tan, S. L, Mahmud, T. Y. T. and Khatijah I., 2002. The Starch Industry –Its Commercial Potential in *The Planter*. **78** (918), 485-494. Kuala Lumpur.
- Trevithick, J. R. and Mitton, K. P., 1999. Antioxidants and Diseases of the Eye. In: Papas, A. M. (ed.). *Antioxidant Status, Diet, Nutrition, and Health*: 37-57. CRC Press Inc., Boca Raton, Florida.
- Werner, I., Bacher, A. and Eisenreich W., 1999. Analysis of Gallic Acid Biosynthesis via Quantitative Prediction of Isotope Labeling Patterns. In: Gross, G. G. et al. (ed.), *Plant Polyphenols 2: Chemistry, Biology, Pharmacology, Ecology (Basic Life Science)* **66**, 43-63. Kluwer Academic/ Plenum Publishers, New York.
- Williams, S. R., 1999. *Essentials of Nutrition and Diet Therapy*. 7th Edition. Mosby Publication, Missouri.
- Williamson, G., Plumb, G. W. and Garcia-Conesa, M. T., 1999. Glycosylation, Esterification, and Polymerization of Flavonoids and Hydroxycinnamates: Effects on Antioxidant Properties. In: Gross, G. G. et al. (ed.), *Plant Polyphenols 2: Chemistry, Biology, Pharmacology, Ecology (Basic Life Science)* **66**, 43-63. Kluwer Academic/ Plenum Publishers, New York.
- Wu, H. C., Shiau, C. Y., Chen, H. M. and Chiou, T. K., 2003. *Journal of Food and Drug Analysis*, **11** (2), 148-153.
- Zamora, R., Hidalgo, F. J., and Tappel, A. L., 1991. Comparativeness of dietary β -carotene, vitamin E, selenium and coenzyme Q₁₀ in rat erythrocytes and plasma. *Journal of Nutrition*, **121**, 50-56.
- Zia-ur-Rehman, A. M. Salaria and Farzana Habib, 2003. Antioxidant Activity of Ginger Extract in Sunflower Oil. *Journal of the Science of Food and Agriculture*. **83**, 624-629. John Wiley & Sons, Ltd.

