Cloning, Expression and Functional Characterization of a Novel a-Humulene Synthase, Responsible for the Formation of Sesquiterpene in Agarwood Originating from Aquilaria malaccensis

ABSTRACT

This study describes the cloning, expression and functional characterization of a-humulene synthase, responsible for the formation of the key aromatic compound a-humulene in agarwood originating from Aguilaria malaccensis. The partial sesquiterpene synthase gene from the transcriptome data of A. malaccensis was utilized for full-length gene isolation via a 30 RACE PCR. The complete gene, denoted as AmDG2, has an open reading frame (ORF) of 1671 bp and encodes for a polypeptide of 556 amino acids. In silico analysis of the protein highlighted several conserved motifs typically found in terpene synthases such as Asp-rich substrate binding (DDxxD), metal-binding residues (NSE/DTE), and cytoplasmic ER retention (RxR) motifs at their respective sites. The AmDG2 was successfully expressed in the E. coli:pET-28a(+) expression vector whereby an expected band of about 64 kDa in size was detected in the SDS-PAGE gel. In vitro enzyme assay using substrate farnesyl pyrophosphate (FPP) revealed that AmDG2 gave rise to two sesquiterpenes: a-humulene (major) and β -caryophyllene (minor), affirming its identity as α -humulene synthese. On the other hand, protein modeling performed using AlphaFold2 suggested that AmDG2 consists entirely of a-helices with short connecting loops and turns. Meanwhile, molecular docking via AutoDock Vina (Version 1.5.7) predicted that Asp307 and Asp311 act as catalytic residues in the a-humulene synthase. To our knowledge, this is the first comprehensive report on the cloning, expression and functional characterization of a-humulene synthase from agarwood originating from A. malaccensis species. These findings reveal a deeper understanding of the structure and functional properties of the a-humulene synthase and could be utilized for metabolic engineering work in the future.