

**CHARACTERIZATION OF *Trichoderma*  
ISOLATED AND THEIR LIGNOCELLULOLYTIC  
ACTIVITIES**

**AZRIAH BINTI ASIS**



**UMS**  
**UNIVERSITI MALAYSIA SABAH**  
**THIS THESIS SUBMITTED IN FULFILLMENT FOR  
THE MASTER OF SCIENCE**

**BIOTECHNOLOGY RESEARCH INSTITUTE  
UNIVERSITI MALAYSIA SABAH  
2017**

# UNIVERSITI MALAYSIA SABAH

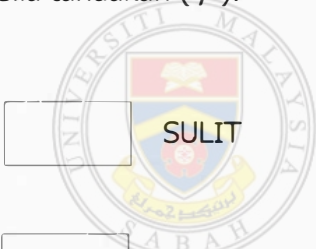
## BORANG PENGESAHAN STATUS TESIS

JUDUL: **CHARACTERIZATION OF *Trichoderma* ISOLATED AND THEIR LIGNOCELLULOLYITIC ACTIVITIES**

IJAZAH: **MASTER OF SCIENCE (BIOTECHNOLOGY)**

Saya, **AZRIAH BINTI ASIS**, Sesi **2013-2017**, mengaku membenarkan tesis sarjana ini disimpan di Perpustakaan Universiti Malaysia Sabah dengan syarat-syarat kegunaan seperti berikut:

1. Tesis ini adalah hak milik Universiti Malaysia Sabah.
2. Perpustakaan Universiti Malaysia Sabah dibenarkan membuat salinan untuk tujuan pengajian sahaja.
3. Perpustakaan dibenarkan membuat salinan tesis ini sebagai bahan pertukaran antara institusi pengajian tinggi.
4. Sila tandakan ( / ):



SULIT

(Mengandungi maklumat yang berdarjah keselamatan atau kepentingan Malaysia seperti yang termaktub di dalam AKTA RAHSIA 1972)

TERHAD

(Mengandungi maklumat TERHAD yang telah ditentukan oleh organisasi/badan di mana penyelidikan dijalankan)

TIDAK  
TERHAD

Disahkan oleh,  
**NURULAIN BINTI ISMAIL**  
LIBRARIAN  
UNIVERSITI MALAYSIA SABAH

**AZRIAH BINTI ASIS**  
**MZ1311002T**

(Tandatangan Pustakawan)

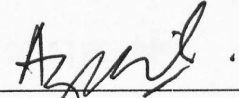
Tarikh: 10 August 2017

(Assoc. Prof. Dr. Shafiquzzaman Siddiquee)  
Penyelia

## DECLARATION

I hereby declare that the material in this thesis is my own except for quotations, excerpts, equations, summaries and references, which have been duly acknowledged.

10 August 2017



---

Azfiah Binti Asis  
MZ1311002T



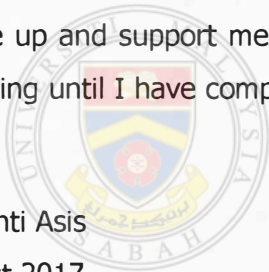
UMS  
UNIVERSITI MALAYSIA SABAH

## ACKNOWLEDGEMENT

I would like to take this great opportunity to thank everyone that helped in succession of my Master thesis completion. First of all, I would like to thank God for giving me strength and blessed all the work of my hand in doing this project. Moreover, the super great supervisor of me has been continuously teaching, guiding and advising me until I am in the correct 'path' of understanding and working on this project without giving up. He is Assoc. Prof. Dr. Shafiquzzaman Siddiquee. I would also like to really thank my friends, Shafawati, Afiza, Rolando, Elizabeth, Salahuddin, Tamar and others that never feel tired of listening to what I'm asking regarding this project and feel frank to share with me the knowledge. Besides, thank you also to lab assistants Mdm. Azian, Mr. Mony, Mr. Emran, Mr. Adam and others that always helped out in understanding the operation of the instruments and providing all my needs in the laboratory. Last but not least, I am thankful to be the daughter of my parents and part of my siblings. They always cheer me up and support me when I feel down or depressed. Thanking you all in participating until I have completed this work.

Azriah Binti Asis

10 August 2017



UMS  
UNIVERSITI MALAYSIA SABAH

## ABSTRACT

*Trichoderma* is a cosmopolitan fungus that prevalent in the soil and other diverse habitats. It has gained vast economical importance's because industrial enzymes production, antifungal, antibiotics, biocontrol agents and plant growth promoter. *Trichoderma* species produced the lignocellulolytic enzymes activities that assisted in the degradation of woody lignocellulose materials for industrial applications. Therefore, an accurately identification of *Trichoderma* isolates at the species level is highly desirable. In this study, the main aim to accurately identify of *Trichoderma* isolates at the species level based on the morphological characteristics, lignocellulolytic enzyme activities and multilocus gene sequencing based on the Internal Transcribed Spacers 1 and 2 (ITS1 and ITS2) regions of the rDNA, a partial sequence of the Translation Elongation Factor 1-alpha (*tef1*) and calmodulin (*cal*). A total of 53 isolates of *Trichoderma* were isolated from wet paddy field in Tuaran, Sabah, Malaysia. On the combination of morphological characteristics and multilocus gene sequencing analysis were positively identified three *Trichoderma* species, namely *T. asperellum*, *T. harzianum* and *T. reesei*. The phylogenetic relationships were constructed based on the Internal Transcribed Spacers 1 and 2 (ITS1 and ITS2) regions of the rDNA gene, a partial sequence of the Translation Elongation Factor 1-alpha (*tef1*) and calmodulin gene (*cal*) by using UPGMA method and found three sections such as *T. asperellum* in the "*Trichoderma*" section, *T. harzianum* in the "*Pachybasium*" section, and *T. reesei* in the "*Longibrachiatum*" section. Mycelial growth and biomass yield among three species isolates were examined on Potato Dextrose Agar and Potato Dextrose Broth, respectively, with different temperatures; 20, 25, 30, 35 and 40 °C. *T. asperellum* isolates were grown very well of mycelia growth and produced the highest biomass yield at 30 °C, followed by *T. harzianum* and *T. reesei*. The lignocellulolytic activities were assayed based on their ability to develop dark brown pigments, yellow halo zone, and clear white zone on tannic acid media (TAM) for lignin, Jensen Media (JM) for cellulose and modified Melin–Nokrans media (MMNM) for starch. The diameters of halo zones were measured for the analysis of their ability in degrading lignin, cellulose, and starch. The best seven *Trichoderma* isolates [S1(9)10<sup>-1</sup>(3), E3(6)10<sup>-1</sup>(2), W2(2)10<sup>-1</sup>(2), S3(1)10<sup>-1</sup>(1), N2(4)10<sup>-2</sup>(3), N2(2)10<sup>-1</sup>(2) dan S3(6)10<sup>-1</sup>(2)] were found the potential lignocellulolytic agents based on the diameter of dark brown pigments and halo zones formed. *Trichoderma* species are found to synthesize polyphenol oxidase, endoglucanases, and are able to hydrolyze starch to glucose in the three different media. Thus, the potential *Trichoderma* isolates can be further applied as biocontrol agents in controlling disease and increasing yield of agricultural crops.

## ABSTRAK

### **MORFOLOGI, LIGNOSELLULOLISIS DAN PENCIRIAN MOLEKUL PENCILAN TRICHODERMA DARIPADA SAWAH PADI, TUARAN**

*Trichoderma* adalah kulat kosmopolitan yang tersebar luas di dalam tanah dan pelbagai habitat lain. Ia mempunyai kepentingan dalam ekonomi kerana pengeluaran enzim untuk industri, anti-kulat, antibiotik, agen kawalan biologi dan membantu pertumbuhan tumbuhan. Spesies *Trichoderma* menghasilkan enzim lignosellulolisis yang membantu dalam degradasi lignoselulosa bahan ber kayu untuk diaplikasikan dalam industri. Oleh itu, pengenalan pastian *Trichoderma* yang tepat di peringkat spesies adalah sangat diperlukan. Dalam kajian ini, tujuan utama adalah untuk mengenal pasti dan mengasingkan *Trichoderma* dengan tepat di peringkat spesies berdasarkan ciri morfologi, aktiviti enzim lignocellulolisis dan multilokus jujukan gen berdasarkan Internal Transcribed Spacers 1 and 2 (ITS1 and ITS2) kelompok rDNA, sebahagian daripada jujukan Translation Elongation Factor 1-alpha (*tef1*) and calmodulin (*cal*). Sebanyak 53 pencilan *Trichoderma* telah diasingkan daripada sawah padi di Tuaran, Sabah, Malaysia. Berdasarkan gabungan ciri morfologi dan analisis multilokus jujukan gen, ia secara positif telah mengenal pasti tiga spesies *Trichoderma* iaitu *T. asperellum*, *T. harzianum* dan *T. reesei*. Hubungan filogenetik telah dibina berdasarkan disalin Internal Transcribed Spacers 1 and 2 (ITS1 and ITS2) kelompok rDNA, sebahagian daripada jujukan Translation Elongation Factor 1-alpha (*tef1*) and calmodulin (*cal*) gen dengan menggunakan kaedah UPGMA dan menunjukkan tiga bahagian iaitu *T. asperellum* dalam seksyen "Trichoderma", *T. harzianum* dalam seksyen "Pachybasium", dan *T. reesei* dalam seksyen "Longibrachiatum". Pertumbuhan miselium dan hasil biojisim antara tiga spesies telah diuji ke atas Potato Dextrose Agar dan Potato Dextrose Broth, masing-masing, dengan suhu yang berbeza; 20, 25, 30, 35 dan 40 °C. *T. asperellum* dapat tumbuh dengan dan menghasilkan pertumbuhan mycelia dan hasil biojisim yang paling tinggi pada 30 °C, diikuti oleh *T. harzianum* dan *T. reesei*. Aktiviti lignocellulolisis telah dicerakinkan berdasarkan keupayaan mereka untuk menghasilkan pigmen coklat gelap, halo zon kuning, dan zon putih yang jelas ke atas media asid tannic (TAM) untuk lignin, Jensen Media (JM) untuk selulosa dan Melin-Nokrans media (MMNM) untuk kanji. Diameter halo zon diukur untuk analisis keupayaan mereka dalam menguraikan lignin, selulosa, dan kanji. Tujuh pencilan *Trichoderma* [*S1(9)10<sup>1</sup>(3)*, *E3(6)10<sup>1</sup>(2)*, *W2(2)10<sup>1</sup>(2)*, *S3(1)10<sup>1</sup>(1)*, *N2(4)10<sup>2</sup>(3)*, *N2(2)10<sup>1</sup>(2)* dan *S3(6)10<sup>1</sup>(2)*] telah dikenalpasti mempunyai potensi sebagai ejen lignocellulolisis berdasarkan diameter coklat gelap dan zon halo yang terbentuk. Spesies *Trichoderma* mampu mensintesis poliphenol oxidase, endoglucanases, dan dapat menghidrolisis kanji kepada glukosa dalam tiga media yang berbeza. Oleh itu, pencilan *Trichoderma* yang berpotensi boleh diaplikasikan sebagai agen kawalan biologi dalam mengawal penyakit dan meningkatkan hasil tanaman pertanian.

# LIST OF CONTENTS

	Page
<b>TITLE</b>	
<b>DECLARATION</b>	ii
<b>CERTIFICATION</b>	iii
<b>ACKNOWLEDGEMENT</b>	iv
<b>ABSTRACT</b>	v
<b>ABSTRAK</b>	vi
<b>LIST OF CONTENTS</b>	vii
<b>LIST OF TABLES</b>	xi
<b>LIST OF FIGURES</b>	xii
<b>LIST OF SYMBOLS AND UNITS</b>	xvii
<b>LIST OF ABBREVIATIONS</b>	xviii
<b>LIST OF APPENDICES</b>	xx
<b>CHAPTER 1 INTRODUCTION</b>	
1.1 General Introduction	1
1.2 Problem Statement	3
1.3 Hypothesis	4
1.4 Research Objectives	4
<b>CHAPTER 2 LITERATURE REVIEW</b>	
2.1 <i>Trichoderma</i>	6
2.2 Biodiversity of <i>Trichoderma</i>	7
2.3 Identification of <i>Trichoderma</i> Species	9
2.3.1 Morphology Identification of <i>Trichoderma</i>	10
(a) Macroscopic Features	10
(b) Microscopic Features	11
2.3.2 Molecular Identification of <i>Trichoderma</i>	12
2.4 Molecular Marker	16

2.4.1	Internal Transcribed Spacers (ITS) of Ribosomal DNA (rDNA)	18
2.4.2	Translational Elongation Factor 1- $\alpha$ ( <i>tef1</i> ) Gene	19
2.4.3	Calmodulin ( <i>cal</i> ) Gene	20
2.5	Phylogenetic Analysis of <i>Trichoderma</i>	21
2.6	Applications and Significance of <i>Trichoderma</i>	21
2.6.1	Biocontrol Agents	21
2.6.2	Production of Important Enzymes	24
2.6.3	Producers of Secondary Metabolites (SMs), Antibacterial and Antibiotic Metabolites	26
2.6.4	Use of <i>Trichoderma</i> in Pollution Remediation	27
2.7	Lignocellulolytic Activity of <i>Trichoderma</i>	29
2.7.1	Microorganisms Producing Cellulose-Degrading Enzymes	33
2.7.2	Microorganisms Producing Hemicellulose-Degrading Enzymes	34
2.7.3	Microorganisms Producing Lignin-Degrading Enzymes	35
<b>CHAPTER 3 METHODOLOGY</b>		
3.1	Sample Collection	37
3.1.1	Soil Sampling	37
3.2	Isolation of the <i>Trichoderma</i> Species	38
3.3	Stock culture of <i>Trichoderma</i> Species	38
3.4	Morphological Characterization	39
3.4.1	Macroscopic Observations	39
3.4.2	Microscopic Observations	39
3.4.3	Effect of Different Temperatures on the Mycelial Growth of <i>Trichoderma</i> Species	40
3.4.4	Effect of Different Temperatures on the Biomass Yield of <i>Trichoderma</i> Species	41



3.5	Molecular Analysis	41
3.5.1	DNA Extraction	41
3.5.2	DNA Amplification	42
3.5.3	Visualization of PCR Products	44
3.5.4	Purification and Sequencing of PCR Products	44
3.5.5	Sequence Analysis and Phylogenetic Inference of ITS 1 and ITS 2 Regions of the rDNA, Elongation Factor 1- $\alpha$ ( <i>tef1</i> ), and Calmodulin ( <i>cal</i> ) Genes	45
3.6	Lignocellulolytic Analysis of <i>Trichoderma</i> Species	46
3.6.1	Enzymatic Degradation of Lignin	46
3.6.2	Enzymatic Degradation of Cellulose	46
3.6.3	Enzymatic Degradation of Starch	47
 <b>CHAPTER 4 RESULTS</b>		
4.1	Introduction	48
4.2	<i>Trichoderma</i> Isolates	48
4.3	Morphological Characterization of <i>Trichoderma</i> Isolates	51
4.3.1	Macroscopic Observation	51
4.3.2	Microscopic Characteristics	53
	(a) <i>Trichoderma asperellum</i>	55
	(b) <i>Trichoderma harzianum</i>	57
	(c) <i>Trichoderma reesei</i>	59
4.3.3	Effect of Temperature on Mycelial Growth of <i>Trichoderma</i> Species	61
4.3.4	Effect of Temperature on Biomass Yield of <i>Trichoderma</i> Species	64
4.4	Molecular Analysis	67
4.4.1	DNA Extraction	67
4.4.2	PCR Amplification and Purification	68
	(a) PCR Amplification of ITS1 and ITS2 Gene	68

	(b) PCR Amplification of Translational Elongation Factor 1- $\alpha$ Gene ( <i>tef1</i> )	68
	(c) PCR Amplification of Calmodulin ( <i>cal</i> ) Gene	69
4.4.3	<i>TrichO</i> KEY Analysis of ITS Regions	70
4.4.4	BLAST Analysis of ITS Regions, <i>tef1</i> and <i>cal</i> Genes	74
4.4.5	Phylogenetic Tree Analysis	75
	(a) Phylogenetic Tree of ITS Regions	76
	(b) Phylogenetic Tree of <i>tef1</i> Gene	78
	(c) Phylogenetic Tree of <i>cal</i> Gene	80
4.5	Lignocellulolytic Activities of <i>Trichoderma</i> Species	82
<b>CHAPTER 5</b>	<b>DISCUSSION</b>	
5.1	Species Identification	86
5.1.1	Morphological Observation	86
5.1.2	Effect of Temperature on Mycelial Growth of <i>Trichoderma</i> Species	89
5.1.3	Effect of Temperature on Biomass Yield of <i>Trichoderma</i> Species	91
5.1.4	Molecular Analysis of <i>Trichoderma</i> Species and Phylogeny	91
5.2	Lignocellulolytic Activities of <i>Trichoderma</i> Species	95
<b>CHAPTER 6</b>	<b>CONCLUSION</b>	98
<b>REFERENCES</b>		100
<b>APPENDICES</b>		123

## LIST OF TABLES

	Page
Table 2.1:	17
Identification of target nucleic loci, primer sets and resolution level within <i>Sclerotinia sclerotiorum</i> or among <i>S. sclerotiorum</i> and related species in the <i>Sclerotiniaceae</i>	
Table 2.2:	29
Bioremediation of various pollutants using <i>Trichoderma</i> spp.	
Table 2.3:	35
Important microorganisms producing hemicellulose-degrading enzymes	
Table 3.1:	43
Information for primers used and source	
Table 4.1:	49
Isolates collection number and origin of the samples used in this study	
Table 4.2:	53
Colony color variations among 53 <i>Trichoderma</i> isolates	
Table 4.3:	54
The representative <i>Trichoderma</i> isolates identified using an online interactive key, shapes and averaged sizes of conidia and phialide	
Table 4.4:	65
Effect of temperatures on dry weight biomass of <i>Trichoderma</i> spp.	
Table 4.5:	71
BLAST and <i>TrichO</i> KEY comparison analysis of ITS1 and ITS2 regions and its accession number	
Table 4.6:	74
BLAST analysis of ITS, <i>tef1</i> and <i>cal</i> gene and GenBank accession number	

## LIST OF FIGURES

		Page
Figure 2.1:	Preparation of the slide for fungal microscopic observation based on Riddle	11
Figure 2.2:	Microscopic characters of (a) <i>Trichodemra harzianum</i> and (b) <i>Trichoderma virens</i> . (i and iii) A conidiophores and phialides (Bar=10 $\mu\text{m}$ ) and (ii and iv) conidia (Bar = 10 $\mu\text{m}$ )	12
Figure 2.3:	Schematic representation of rDNA region with primers ITS1 and ITS2 localization (arrows)	18
Figure 2.4:	Schematic representations of <i>tef1</i> regions including intron and exon	20
Figure 2.5:	Diagrammatic structure and chemical composition of lignocellulose residues. The plant cell wall contains three major layers, namely, the middle lamella, the primary wall and the secondary wall.	31
Figure 2.6:	Structure of lignin and lignin precursors of H-, G-, and S- units in lignin	32
Figure 2.7:	Structure of cellulose formed from $\beta$ -1, 4-linked cellobiose units, with hydrogen bonding between parallel chains	32
Figure 3.1:	Random sampling of different identified sites in wet paddy field, Tuaran, Sabah	37
Figure 3.2:	(a) Preparation of slide culture and (b) placing of slide culture on the glass rod rest on the filter paper in the Petri dish	40
Figure 4.1:	Population number (CFU) of <i>Trichoderma</i> spp. versus non <i>Trichoderma</i> in Tuaran wet paddy field	50
Figure 4.2:	A representative culture of <i>Trichoderma</i> isolates after 6 days incubation on PDA at $28 \pm 2$ °C. A: $E1(4)10^{-1}(1)$ , B: $E1(4)10^{-1}(3)$ , C: $E3(3)10^{-1}(2)$ , D: $N2(1)10^{-2}(3)$ , E: $N2(6)10^{-1}(3)$ , F: $S1(9)10^{-1}(3)$ , G: $W2(4)10^{-1}(2)$ , G: $N1(5)10^{-1}(3)$ , H: $S3(4)10^{-1}(1)$	52
Figure 4.3:	A representative isolate [ $N3(3)10^{-2}(1)$ ] shows	56

morphological characteristics of *T. asperellum*. a: Front colony grown in PDA for 6 days; b: Reverse colony; c: Conidia; d-f. Conidiaphores. c-f were observed under light microscope with 400X magnification

- Figure 4.4: A representative isolate [W2(2)10<sup>-1</sup>(2)] shows morphological characteristics of *T. harzianum*. a: Front colony grown in PDA for 6 days; b: Reverse colony; c: Conidia; d-f: Conidiaphores. c-f were observed under light microscope with 400X magnification 58
- Figure 4.5: A representative isolate [S1(9)10<sup>-1</sup>(3)] shows morphological characteristics of *T. reesei*. a: Front colony grown in PDA for 6 days; b: Reverse colony; c: Conidia; d-f: Conidiaphores. c-f were observed under light microscope with 400X magnification 60
- Figure 4.6: Colony radius of *T. asperellum*, *T. harzianum* and *T. reesei* grown on PDA at 20 °C from day 1 to day 5. Values are the means of three replicates. Vertical bars indicate standard deviations of the mean 61
- Figure 4.7: Colony radius of *T. asperellum*, *T. harzianum* and *T. reesei* grown on PDA at 25 °C from day 1 to day 5. Values are the means of three replicates. Vertical bars indicate standard deviations of the mean 62
- Figure 4.8: Colony radius of *T. asperellum*, *T. harzianum* and *T. reesei* grown on PDA at 30 °C from day 1 to day 5. Values are the means of three replicates. Vertical bars indicate standard deviations of the mean 63
- Figure 4.9: Colony radius of *T. asperellum*, *T. harzianum* and *T. reesei* grown on PDA at 35 °C from day 1 to day 5. Values are the means of three replicates. Vertical bars indicate standard deviations of the mean 64
- Figure 4.10: Effect of temperature on dry-weight biomass yield of *T. asperellum*, *T. harzianum* and *T. reesei* grown on PDB. The biomass was weighted after 8 66

days of incubation prior to 24 hours of oven-dried at 60 °C. Values are the means of three replicates. Horizontal bars indicate standard errors of the mean

- Figure 4.11: Agarose gel electrophoresis of genomic DNA extracted from *Trichoderma* isolates. The white error indicates the genomic DNA extracted. Lane M: 100 bp DNA ladder, Lane 1: E1(4)10<sup>-1</sup>(3), Lane 2: N1(2)10<sup>-2</sup>(1), Lane 3: N2(2)10<sup>-1</sup>(1), Lane 4: N2(4)10<sup>-2</sup>(3), Lane 5: N2(6)10<sup>-1</sup>(3), Lane 6: N3(1)10<sup>-2</sup>(2); Lane 7: N3(2)10<sup>-2</sup>(2); Lane 8: N3(3)10<sup>-1</sup>(1); Lane 9: W1(6)10<sup>-1</sup>(3); Lane 10: W2(2)10<sup>-1</sup>(2); Lane 11: W2(4)10<sup>-1</sup>(2); Lane 12: S1(2)10<sup>-1</sup>(3); Lane 13: S1(7)10<sup>-1</sup>(3); Lane 14: S1(9)10<sup>-1</sup>(3); Lane 15: S2(3)10<sup>-1</sup>(1); Lane 16: S2(3)10<sup>-1</sup>(2) 67
- Figure 4.12: PCR amplification of ITS genes in the nine *Trichoderma* isolates. Lane C: Control, Lane M: 100 bp DNA ladder (First Base), Lane 1: N1(4)10<sup>-1</sup>(3), Lane 2: E3(7)10<sup>-1</sup>(2), Lane 3: E2(6)10<sup>-1</sup>(1), Lane 4: N1(3)10<sup>-2</sup>(1), Lane 5: N1(7)10<sup>-1</sup>(3), Lane 6: N1(8)10<sup>-1</sup>(3), Lane 7: E3(8)10<sup>-1</sup>(2), Lane 8: E3(2)10<sup>-2</sup>(1), Lane 9: E3(3)10<sup>-1</sup>(2) 68
- Figure 4.13: PCR amplification of *tef1* gene in the 15 *Trichoderma* isolates. C: Control; M: 100 bp DNA ladder (First Base); Lane 1: N1(1)10<sup>-2</sup>(2); Lane 2: N1(2)10<sup>-2</sup>(1); Lane 3: N1(3)10<sup>-2</sup>(1); Lane 4: N1(4)10<sup>-1</sup>(3); Lane 5: N1(7)10<sup>-1</sup>(3); Lane 6: N1(8)10<sup>-1</sup>(3); Lane 7: N2(1)10<sup>-2</sup>(3); Lane 8: N2(2)10<sup>-1</sup>(1); Lane 9: N2(4)10<sup>-2</sup>(3); Lane 10: N2(6)10<sup>-1</sup>(2); Lane 11: N3(1)10<sup>-2</sup>(2); Lane 12: N3(2)10<sup>-2</sup>(1); Lane 13: N3(3)10<sup>-2</sup>(1); Lane 14: N2(6)10<sup>-1</sup>(3); Lane 15: N3(3)10<sup>-2</sup>(3) 69
- Figure 4.14: PCR amplification of *cal* gene in the 9 *Trichoderma* isolates. C: Control; M: 100 bp DNA ladder (First Base); Lane 1: N1(3)10<sup>-2</sup>(1); Lane 2: N1(5)10<sup>-1</sup>(3); Lane 3: N2(6)10<sup>-1</sup>(3); Lane 4: N1(3)10<sup>-2</sup>(1); Lane 5: N3(3)10<sup>-2</sup>(3); Lane 6: S1(7)10<sup>-1</sup>(3); Lane 7: E3(6)10<sup>-1</sup>(2); Lane 8: S3(4)10<sup>-2</sup>(3); Lane 9: S3(2)10<sup>-2</sup>(2) 70
- Figure 4.15: Species identification results of E3(2)10<sup>-1</sup>(2) by (a) *TrichO*KEY and (b) BLAST search. The BLAST 73

result shows top five hits with 99% similarity to *T. asperellum*

- Figure 4.16: Phylogenetic relationship of 53 *Trichoderma* isolates inferred by UPGMA analysis of ITS1 and ITS2 regions. The numbers given above the selected branches indicate the bootstrap coefficients >50%. The bold letter indicates the respective sequences from GenBank, whereas all the isolates used in this study are given by the collection number without species identification. *Fusarium solani* (AM412643) act as the outgroup for this analysis 77
- Figure 4.17: Phylogenetic relationship of 53 *Trichoderma* isolates inferred by UPGMA analysis of *tef1* sequences. The numbers given above the selected branches indicate the bootstrap coefficients >50%. The bold letter indicates the respective sequences from GenBank, whereas all the isolates used in this study are given by the collection number without species identification. *Fusarium solani* (JF740784) act as the outgroup for this analysis 79
- Figure 4.18: Phylogenetic relationship of 53 *Trichoderma* isolates inferred by UPGMA analysis of *cal* sequences. The numbers given above the selected branches indicate the bootstrap coefficients >50%. The bold letter indicates the respective sequences from GenBank, whereas all the isolates used in this study are given by the collection number without species identification. *Fusarium solani* (HQ412319) act as the outgroup for this analysis 81
- Figure 4.19: A representative picture of *Trichoderma* isolates on their ability to degrade lignin producing dark brown zone. (a) W2(2)10<sup>-1</sup>(2)= 12.9 mm; (b) N1(5)10<sup>-1</sup>(3)= 13.03 mm; (c) S3(4)10<sup>-1</sup>(1)= 9.37 mm; (d) S1(2)10<sup>-1</sup>(3)= 5.82 mm 82
- Figure 4.20: A representative picture of *Trichoderma* isolates on their ability to degrade cellulose producing halo zone. (a) S1(9)10<sup>-1</sup>(3)= 14.42 mm; (b) S3(1)10<sup>-1</sup>(1)= 10.44 mm; (c) E3(6)10<sup>-1</sup>(2)= 8.93 mm; (d) N3(3)10<sup>-2</sup>(1)= 0 mm 83

Figure 4.21: A representative picture of *Trichoderma* isolates on their ability to degrade starch producing clear halo zone. (a)  $W2(2)10^{-1}(2)= 12.22$  mm; (b)  $W2(4)10^{-1}(2)= 18.0$  mm; (c)  $S3(4)10^{-1}(1)= 21.35$  mm; (d)  $N1(5)10^{-1}(3)= 10.93$  mm 84

Figure 4.22: Screening among 20 *Trichoderma* isolates on their ability to degrade lignin, cellulose and starch on tannic acid media, Jensen media and modified Melin-Nokrans media, respectively. Values are the means of three replicates. Vertical bars indicate standard errors of the mean 85



UMS  
UNIVERSITI MALAYSIA SABAH



## LIST OF SYMBOLS AND UNITS

<b>cfu.g<sup>-1</sup></b>	-	Colony forming unit per gram
<b>μM</b>	-	Micrometer
<b>mm</b>	-	Milimeter
<b>mg.mL<sup>-1</sup></b>	-	Milligram per millilitre
<b>ng</b>	-	Nanogram
<b>μg</b>	-	Microgram
<b>g</b>	-	Gram
<b>mL</b>	-	Mililiter
<b>μL</b>	-	Microliter
<b>mM</b>	-	Milimolar
<b>nm</b>	-	Nanometer
<b>min</b>	-	Minute
<b>sec</b>	-	Second
<b>°C</b>	-	Degree celcius
<b>%</b>	-	Percentage
<b>±</b>	-	Plus minus
<b>rpm</b>	-	Rotation per minute
<b>V</b>	-	Voltan



UMS  
UNIVERSITI MALAYSIA SABAH

## LIST OF ABBREVIATIONS

<b>6PP</b>	-	6-penthyl- $\alpha$ -pyr-one
<b>BCA</b>	-	Biological control agent
<b>BLAST</b>	-	Basic Local Alignment Search Tool
<b>bp</b>	-	Basepair
<b>C/N</b>	-	Carbon to nitrogen ratio
<b>CAZy</b>	-	Carbohydrate-active enzymes
<b>CFU</b>	-	Colony Forming Unit
<b>CMC</b>	-	Carboxymethyl cellulose
<b>CTAB</b>	-	Cetyltrimethylammonium Bromide
<b>DNA</b>	-	Deoxyribonucleotide acid
<b>dNTP</b>	-	Deoxynucleotides triphosphate
<b>EDTA</b>	-	Ethylenediaminetetraacetic acid
<b>EFB</b>	-	Empty fruit bunch
<b>EtBr</b>	-	Ethidium bromide
<b><i>tef1</i></b>	-	Translational elongation factor 1- $\alpha$
<b>ISTH</b>	-	International Subcommission on <i>Trichoderma</i> and <i>Hypocrea</i> Taxonomy
<b>ITS 1</b>	-	Internal transcribed spacer 1
<b>ITS 2</b>	-	Internal transcribed spacer 2
<b>ITS 4</b>	-	Internal transcribed spacer 4
<b>JM</b>	-	Jensen media
<b>Kb</b>	-	Kilo base

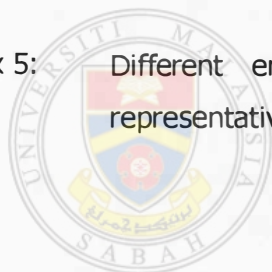
<b>LiPs</b>	-	Lignin peroxidases
<b>MEGA</b>	-	Molecular Evolutionary Genetic Analysis
<b>MMNM</b>	-	Modified-Melin Nokrans media
<b>MnPs</b>	-	Manganese-peroxidases
<b>NCBI</b>	-	National Center for Biotechnology Information
<b>PAHs</b>	-	Polycyclic aromatic hydrocarbons
<b>PCR</b>	-	Polymerase Chain Reaction
<b>PDA</b>	-	Potato Dextrose Agar
<b>PDB</b>	-	Potato Dextrose Broth
<b>PPO</b>	-	Polyphenol oxidase
<b>rDNA</b>	-	Ribosomal DNA
<b>rRNA</b>	-	Ribosomal RNA
<b>Sect.</b>	-	Section
<b>SMs</b>	-	Secondary metabolites
<b>spp.</b>	-	Species
<b>TAM</b>	-	Tannic acid media
<b>TBE</b>	-	Tris-Borate-EDTA
<b>tRNA</b>	-	Transfer ribonucleic acid
<b>TSM</b>	-	<i>Trichoderma</i> Selective Medium
<b>UPGMA</b>	-	Unweighted Pair Group with Arithmetic Mean



UMS  
UNIVERSITI MALAYSIA SABAH

## LIST OF APPENDICES

	Page
Appendix 1: Preparation of media and buffer.	124
Appendix 2: List of sequences from GenBank that used for the phylogenetic analysis of ITS, <i>tef1</i> and <i>cal</i> genes.	128
Appendix 3: The colony color of <i>Trichoderma</i> isolates after 6 days incubation on PDA at $28 \pm 2$ °C.	129
Appendix 4: Averaged colony radius of <i>Trichoderma</i> isolates cultures on PDA at 20, 25, 30 and 35 °C from day 1 to day 5.	133
Appendix 5: Different enzymatic activity exhibited by 20 representative strains of <i>Trichoderma</i> .	135



UMS  
UNIVERSITI MALAYSIA SABAH

# CHAPTER 1

## INTRODUCTION

### 1.1 General Introduction

The genus *Trichoderma* belongs to ascomycetic (Ascomycota, Hypocreales) fungi found in various ecosystems such as agricultural fields, forest, salt marshes and deserts, in almost all climatic zones (Roiger, Jeffers and Caldwell, 1991; Samuels, 2006; Kumar, Amaresan, Bhagat, Madhuri and Srivasta, 2010). Some *Trichoderma* spp. have economic importance because of its potential producers of enzymes, antibiotics and used as a biocontrol agent in the agricultural field (Harman and Björkman, 1998; Monte, 2001). Thus, precise identification and characterization of these fungi is a vital requirement. The early approach for identification of *Trichoderma* is on the morphological basis (Rifai 1969; Bissett, 1984; Dodd, Lieckfeldt and Samuels, 2000). Morphological descriptions such as colony appearance and microscopic characteristics which include phialide and conidia sizes, conidiophores and formation of chlamydospores were observed. However, morphological alone is insufficient to identify *Trichoderma* accurately, since they are genetically diverse and characterized by variable morphology particularly between the closely related species of *Trichoderma* (Chaverri and Samuels, 2003; Chaverri, Castlebury, Samuels and Geiser, 2003; Druzhinina, Kubicek, Komoń-Żelazowska, Mulaw and Bissett, 2010).

The advent in molecular methods and identification tools based on molecular data from DNA sequencing has led to satisfactory taxonomy identification. DNA-based methods based on DNA barcoding are now routinely used in *Trichoderma* identification. It has done based on multilocus DNA sequence analysis of internal transcribed spacers (ITS) 1 and 2 of the ribosomal deoxynucleic acid (rDNA), gene cluster and fragments of the translational elongation factor 1- $\alpha$

(*tef1*), RNA polymerase II subunit (*rpb2*), chitinase 18-5 (*chi18-5*), actin (*act*) or calmodulin (*ca1*) (Kindermann, El-Ayouti, Samuels and Kubicek, 1998; Dodd *et al.*, 2000; Druzhinina, Kopchinskiy, Komoń, Bissett, Szakacs and Kubicek, 2005; Gal-Hemed, Atanasova, Komoń-Żelaswoska, Druzhinina, Viterbo and Yarden, 2011; Blaszczyk, Popiel, Chelkowski, Koczyk, Samuels, Sobieralski and Siwulski, 2011; Atanasova, Druzhinina and Jaklitsch, 2013; Jaklitsch and Voglmayr, 2015). Most studies have been used the combination of ITS and *tef1* for the identification of new species of *Trichoderma* (Bissett, Szakacs, Nolan, Druzhinina, Gradinger and Kubicek, 2003; Kraus, Druzhinina, Gams, Bissett, Zafari, Szakacs, Kopchinskiy, Prillinger, Zare and Kubicek, 2004; Lu, Druzhinina, Fallah, Chaverri, Gradinger, Kubicek and Samuels, 2004). In addition, sequence data may useful for phylogeny study, providing valuable insights into their evolutionary relationships. Moreover, the correct identification serves as an efficient selection and use of such isolates for commercial applications.

Druzhinina *et al.* (2005) introduced an online program using oligonucleotide barcodes, based on ITS and *tef1* sequences, known as International Subcommittee on *Trichoderma* and *Hypocrea* Taxonomy (ISTH). At present, the database has listed more than 100 species which have been identified based at the molecular level. The basis of the method is an oligonucleotide barcode generated from a diagnostic combination of various oligonucleotides of ITS1 and ITS2 sequences. These online tools have been allowed the identification of most *Trichoderma* isolates or suggest them as representatives of potentially new species.

Lignocellulolytic enzymes are produced by a wide variety of fungi, including species in *Trichoderma* genus. Some *Trichoderma* spp. are great hydrolytic enzyme producer and therefore important for the biotechnological industry, such as *T. reesei* and *T. viride* (Mandels, Weber and Parizek, 1971; Domigues, Queiroz, Cabral and Fonseca, 2000). The search for potential biomass degrading enzymes also led to the isolation of these fungi (Sallenave-Namont, Pouchus, du Pont, Lossus and Verbis, 2000). Lignocellulolytic fungi have a potential to degrade a range of the lignocellulosic biomass. Many lignocellulosic materials such as wood, bagasse and

wheat straw have been studied as potential substrates for the production of lignocellulolytic enzymes (Duff and Murray, 1996; Ogel, Yarangumeli, Dundar and Ifrij, 2001; Kalogerist, Christakopoulos, Katapodis, Alexiou, Vlachou, Kekos and Macris, 2003). The purified lignocellulolytic enzymes are used for commercial applications such as in coffee production where the hydrolysis of cellulose occurs during the drying of beans (Mussatto, Carneiro, Silva, Roberto and Teixeira, 2011). Moreover, the application of lignocellulolytic fungi improve the composting process of biomass where the C:N ratio was not at the optimal rate (Hart, Leij, Kinsey and Lynch, 2002).

## 1.2 Problem Statement

Several potential strains of *Trichoderma* are beneficial features which have been used as a producers of a range important hydrolytic enzymes, as well as a biological control agents (Verma, Brar, Tyagi, Surampally and Valéro, 2007; Sant, Casanova, Segarra, Avilés, Reis and Trillas, 2010). Previously, *Trichoderma* spp. identification is dependent primarily on morphological characters such as growth rate, colony color, size and length of conidia and phialides. Alwindia and Hirooka (2015) claimed that the size of conidia and phialides overlapped between *T. catoptron* and *T. stramenium*, made identification more complicated. Additionally, the morphological and cultural characters are difficult to accurately define species. Gams and Bissett (1998) reported that variations among *Trichoderma* spp. could not be differentiated satisfactorily *via* morphological methods, thus making nomenclature placement uncertain.

DNA sequence analysis became a new paradigm in fungal systematics for *Trichoderma* (Samuels, 2006). The development of molecular tools has enabled the positive identification of any strains. Early stages of molecular research of *Trichoderma* identification rely on the sequencing of ITS regions of rDNA. In addition, a multigene approach using at least two unlinked loci is desirable for the molecular identification of closely related *Trichoderma* spp. Genes of *tef1*, *rpb2*, *cal* and *act*, for example, can be used in combination with ITS regions to reflect

differences between and within groups of closely related species. Hence, combination of morphological and DNA data will provide at the species level of *Trichoderma* identification. The result from this study stress the importance of combination both morphological and molecular identification tools to describe diversity of *Trichoderma* in paddy fields of Tuaran.

Microbial composting is an effective alternative for the recycling of biomass materials into compost. It promotes sustainable agriculture and environmental protection, as well improving soil's physical, chemical and biological properties (Mylavarapu and Zinati, 2009). Composting process required rapid biodegradation to break down complex lignin, cellulose and starch. *Trichoderma* spp. has the potential to degrade lignocellulosic materials efficiently. Therefore, this study also focused on the screening and evaluating the compatible lignocellulolytic potential among *Trichoderma* spp. for rapid and environmental friendly composting process.

### **1.3 Hypothesis**

Accurate identification of *Trichoderma* species requires both molecular genotyping and phenotypical characterization. Specific *Trichoderma* strains produced vast amount of lignocellulolytic enzymes such as cellulases and hemicellulase.

### **1.4 Research Objectives**

The main focus of this study is to characterize and identify *Trichoderma* spp. isolated from wet paddy field soil samples, Tuaran. The species identification was conducted by using morphological characterization and sequencing analysis of the ITS region, *tef1* and *cal* genes. Moreover, the lignocellulolytic activities of *Trichoderma* spp. were evaluated by screening their degradation towards lignin, cellulose and starch degradation.