

**ANALYSIS OF THE COLD ADAPTATION STRATEGY
OF ANTARCTIC YEAST
Glaciozyma antarctica PI12**



JOSEPH KOH SOON PENG

UMS
UNIVERSITI MALAYSIA SABAH

**BIOTECHNOLOGY RESEARCH INSTITUTE
UNIVERSITI MALAYSIA SABAH
2016**

**ANALYSIS OF THE COLD ADAPTATION STRATEGY
OF ANTARCTIC YEAST
Glaciozyma antarctica PI12**



JOSEPH KOH SOON PENG

UMS

**DISSERTATION SUBMITTED IN FULFILLMENT
FOR THE DEGREE OF DOCTOR OF PHILOSOPHY
IN BIOTECHNOLOGY**

**BIOTECHNOLOGY RESEARCH INSTITUTE
UNIVERSITI MALAYSIA SABAH
2016**

DECLARATION

I hereby declare that the material in this project is my own except for quotations, excerpts, equations, summaries and references, which have been duly acknowledged.

21st August 2015

Joseph Koh Soon Peng
PB 20119024



UMS
UNIVERSITI MALAYSIA SABAH

CERTIFICATION

NAME : **JOSEPH KOH SOON PENG**

MATRIC NO. : **PB2011-9024**

TITLE : **ANALYSIS OF THE COLD ADAPTATION
STRATEGY OF ANTARCTIC YEAST *Glaciozyma
antarctica* PI12**

DEGREE : **DOCTOR OF PHILOSOPHY IN BIOTECHNOLOGY**

VIVA DATE : **4th APRIL 2016**



DECLARED BY;
UMMS
UNIVERSITI MALAYSIA SABAH

1. SUPERVISOR

Prof. Dr. Clemente Michael Wong Vui Ling

Signature

2. CO-SUPERVISOR

Dr. Christopher Voo Luk Yung

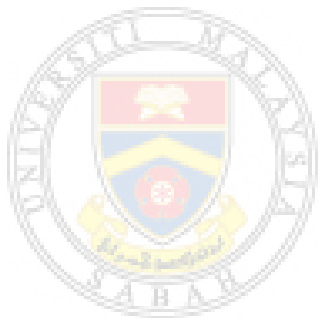
ACKNOWLEDGEMENT

Thank God for His Grace! I would like to express my deepest gratitude and sincere appreciation to Professor Dr. Michael Wong, a great supervisor, for his invaluable guidance and encouragement throughout this study. Also, my sincere appreciation conveyed to Dr. Christopher Voo, for his extensive advice and kind support in this study.

I am grateful to the Biotechnology Research Institute (BRI) for the comfortable working environment, and I am thankful to all the laboratory assistants for their support. We would like to thank Universiti Malaysia Sabah (UMS) and Malaysia Genome Institute for providing facilities and support, this work is supported by MOSTI (Project No: 08-05-MGI-GMB001).

My appreciation is extended to all the laboratory mates, especially to Raimi Redman, Suhaila from UKM, Dexter, for their sincerity helps and friendship throughout my entire research time.

Finally, I would like to express my deepest gratitude and love to my family, including my wife, Cassandra and my daughter, Melissa Jayne, for their love, patience, encouragement and support!



UMS
UNIVERSITI MALAYSIA SABAH

ABSTRACT

Analysis of the cold adaptation strategy of Antarctic yeast *Glaciozyma antarctica* PI12

Psychrophilic yeast *Glaciozyma antarctica* PI12 was isolated from Antarctica. However, the information related to psychrophilic yeast and genus *Glaciozyma* is limited. Therefore, characterization of growth, cell doubling time, cell division, aerobic and partial anaerobic respiration system, morphology and the growth at -12°C , -7°C and -5°C were carried out. Our result showed that *G. antarctica* PI12 formed whitish creamy colony on PDA media, and has an optimal growth temperature of 12°C in YPD media. Its cell doubling time is 15.8 hours per generation, and the cell division occurs on either poles of the cell. *G. antarctica* PI12 can grow under both aerobic and partially anaerobic conditions, but with a faster growth at aerobic condition. Little is known about the other genes which are involved in the cold adaptation of *G. antarctica* PI12. Therefore, to understand the adaptation strategies of *G. antarctica* PI12, RNA-seq was carried out followed by a *de novo* assembly of *G. antarctica* PI12 transcriptome using the Trinity assembly package. Thermal stresses such as -12°C , 0°C , 16°C and 20°C were used to induce a maximum number of expressed genes by *G. antarctica* PI12. We have obtained approximately 465 million of reads using the paired-end Illumina sequencing platform. These reads was assembled into 6,301 unique genes, which comprised of a total of 46,196 unique transcripts (UT) sequences (mean sequence length $\sim 1,555$ bp) including 29,885 UTs with coding sequence (CDS). Our data provide the first comprehensive sequence resource available for functional genomics studies in *G. antarctica* PI12. Besides, the gene expression patterns of *G. antarctica* PI12 in response to rapid temperature shifts were determined. 205 and 206 genes were affected when the cells were rapidly shifted from 12°C to 0°C or -12°C in minimal media, and YPD media. When the cells were rapidly shifted from 12°C to 16°C and 20°C , 116 genes were expressed. We grouped the genes obtained from minimal media and YPD into the early cold response (ECR, 0°C for six hours); late cold response (LCR, 0°C for 24 hours); early freeze response (EFR, -12°C for six hours); and late freeze response (LFR, -12°C for 24 hours). On the other hand, we grouped expressed genes in the heat shock response to the early heat response (EHR, 16°C for six hours); and late heat response (LHR, 16°C and 20°C for 24 hours); early heat response (EHR, 20°C for six hours); and late heat response (LHR, 20°C for 24 hours). Interestingly, there are groups of genes expressed consistently according to the time incubation at six and 24 hours. The result implies that the thermal specific early and late responses are mediated by a different and yet uncharacterized regulatory proteins. An adaptation model of *G. antarctica* PI12 which involved three components, namely the inactivation, the adaptive and the cell death was constructed based on the results, it indicates the complexity of the adaptation strategy of *G. antarctica* PI12 to adapt to a changing temperature.

ABSTRAK

Glaciozyma antarctica PI12 adalah sejenis yis basidiomycetes dan psikrofilik yang telah diasingkan daripada Antartika. Namun, maklumat yang berkaitan dengan yis psikrofilik dan genus *Glaciozyma* adalah terhad. Untuk memahami dengan lebih lanjut mengenai *G. antarctica* PI12, siasatan terhadap ciri-ciri pertumbuhan, masa gandaan sel, pembahagian sel, system respirasi aerobik dan anaerobik seprara, morfologi dan kadar tumbuh semasa dieram pada -12°C , -7°C dan -5°C . Perhatian kami menunjukkan bahawa *G. antarctica* PI12 mempunyai permukaan berkrum putih pada media Potato Dextrose Agar (PDA), dan mempunyai suhu pertumbuhan optimum pada suhu 12°C dalam media C dalam media Yeast Peptone Dextrose (YPD). Masa sel mengganda adalah 15.8 jam setiap generasi, dan pembahagian sel yang berlaku pada kedua-dua belah hujung sel. *G. antarctica* PI12 boleh bertumbuh bawah kedua-dua keadaan aerobik dan anaerobik, walaupun begitu, keadaan aerobik memberikan pertumbuhan yang lebih cepat. Maklumat terhadap gen-gen yang terlibat dalam adaptasi sejuk atau panas di dalam *G. antarctica* PI12 juga terhad. Oleh itu, kajian RNA-seq telah dilaksanakan dan diikuti oleh pemasangan RNA secara *de novo* menggunakan pakej pemasangan Trinity. Tegasan haba seperti -12°C , 0°C , 16°C dan 20°C telah digunakan untuk mendorong bilangan maksimum gen yang disalin oleh *G. antarctica* PI12. Sekira-kira 465 juta daripada penjujukan Illumina telah diperolehi. Termasuk 6301 gen yang unik, terdiri daripada sejumlah 46,196 transkrip unik (UT) urutan (min panjang urutan $\sim 1,555$ bp) termasuk 29,885 SUA dengan pengekodan urutan (CDS) yang diperlukan oleh *G. antarctica* PI12 semasa keadaan haba. Data kami merupakan sumber urutan komprehensif yang pertama yang ada untuk pelengkap data genomik *G. antarctica* PI12 yang sedia ada. Selain itu, corak ekspresi gen *G. antarctica* PI12 sebagai tindak balas kepada perubahan suhu pesat telah ditentukan. 205 gen dan 206 gen terjejas apabila sel-sel telah beralih secara pantas daripada 12°C kepada 0°C atau -12°C dalam YPD and MM. 116 gen telah terjejas apabila sel-sel telah beralih secara pantas daripada 12°C hingga 16°C atau 20°C . Gen-gen diperolehi daripada semua eksperimen adalah reaksi sejuk awal (ECR, 0°C selama enam jam); reaksi sejuk lewat (LCR, 0°C untuk 24 jam); reaksi membekukan awal (EFR, -12°C selama enam jam); dan reaksi pembekuan lewat (LFR, -12°C untuk 24 jam). Kami juga mengumpul gen dinyatakan dalam reaksi kejutan haba kepada reaksi hangat awal (EHR, 16°C selama enam jam); dan reaksi hangat lewat (LHR, 16°C dan 20°C selama 24 jam); reaksi haba awal (EHR, 20°C selama enam jam); dan reaksi haba lewat (LHR, 20°C selama 24 jam). Kami juga terkumpul gen bersalin konsisten mengikut masa pengeraman pada enam dan 24 jam. Keputusan kami menunjukkan bahawa reaksi tertentu berfungsi pada keadaan berubah awal dan lewat telah diantarai oleh protein-protein yang berbeza dan protein yang belum dicirikan. Selain itu, satu model penyesuaian *G. antarctica* PI12 mengandungi tiga komponen, iaitu komponen menyahaktifkan, komponen penyesuaian dan komponen kematian sel telah dibina berdasarkan keputusan yang didapati, model tersebut menunjukkan kerumitan strategi adaptasi daripada *G. antarctica* PI12 untuk menyesuaikan diri dengan suhu yang berubah-ubah.

TABLE OF CONTENTS

	Page
TITLE	i
DECLARATION	ii
CERTIFICATION	iii
ACKNOWLEDGEMENT	iv
ABSTRACT	v
ABSTRAK	vi
TABLE OF CONTENTS	vii
LIST OF TABLES	xiii
LIST OF FIGURES	xiv
LIST OF SYMBOLS & ABBREVIATIONS	xvii
LIST OF APPENDICES	xviii
CHAPTER 1: INTRODUCTION	
1.1 Preamble	1
1.2 Objectives	3
CHAPTER 2: LITERATURE REVIEW	
2.1 Antarctica, the extreme niche	4
2.2 Departure of the continent of Antarctica from the Supercontinent of Gondwana	4
2.3 Endemism of microorganisms found in Antarctica	5
2.4 Psychrophiles	6
2.4.1 Psychrophilic yeast	7
2.4.2 Psychrophilic basidiomycota found in Antarctica	8
2.4.3 Cell division in psychrophilic yeast	11
2.4.4 Respiration of psychrophilic yeast	11
2.5 Complete genome sequence of psychrophilic microorganisms	12
2.6 Enzymes, proteins and genes adapted to cold	13
2.7 General low temperature adaptation strategies	17
2.7.1 Control of molecular motion	19

2.7.2	Reactive oxygen species (ROS)	20
2.7.3	Membrane fluidity	26
2.7.4	Molecular sensors	27
2.7.5	Antifreeze proteins	28
2.8	Adaptation of psychrophilic yeast	30
2.9	<i>Glaciozyma antarctica</i> PI12	31
2.10	Transcriptome approaches	36
2.11	Research approaches	38

CHAPTER 3: CHARACTERIZATION OF ANTARCTIC YEAST *Glaciozyma antarctica* PI12

3.1	Introduction	40
3.2	Literature Review	41
3.3	Materials & Methods	42
3.3.1	Maintenance of strain	42
3.3.2	Growth measurement of <i>Glaciozyma antarctica</i> PI12	43
3.3.3	Growth rate of <i>Glaciozyma antarctica</i> PI12	43
3.3.4	Doubling time assessment of <i>Glaciozyma antarctica</i> PI12	43
3.3.5	Subzero temperature growth study of <i>Glaciozyma antarctica</i> PI12	44
3.3.6	Aerobic and anaerobic analysis of <i>Glaciozyma antarctica</i> PI12	44
3.3.7	Microscopic analysis	44
	a. DAPI counterstaining	44
	b. Scanning Electron Microscope (SEM)	45
3.3.8	Cell measurement	45
3.3.9	Molecular analysis of <i>Glaciozyma antarctica</i> PI12	45
	a. Genomic DNA extraction of <i>Glaciozyma antarctica</i> PI12	45
	b. Characterization of extrachromosomal DNA isolated from <i>Glaciozyma antarctica</i> PI12	46
	c. Rolling Circle Amplification (RCA)	46
	d. Ultrasonic fragmentation	47
	e. Cloning	47
	f. Plasmid miniprep	47

	g. Primer design and Polymerase Chain Reaction (PCR)	47
3.4	Results & Discussion	49
3.4.1	The growth of <i>Glaciozyma antarctica</i> PI12	49
3.4.2	Doubling time of <i>Glaciozyma antarctica</i> PI12	52
3.4.3	Aerobic and anaerobic analysis of <i>Glaciozyma antarctica</i> PI12	56
3.4.4	Microscopic Analysis of <i>Glaciozyma antarctica</i> PI12 Using Fluorescent Microscope And Scanning Electron Microscope (SEM)	58
3.4.5	Molecular characterization of <i>Glaciozyma antarctica</i> PI12	66
3.5	Conclusion	76

CHAPTER 4: *De Novo* Transcriptome Assembly And Genes From The Antarctic Yeast *Glaciozyma antarctica* PI12

4.1	Introduction	77
4.2	Literature Review	78
4.3	Materials & Methods	79
4.3.1	<i>Glaciozyma antarctica</i> PI12 strain and culture conditions	79
4.3.2	cDNA library construction and sequencing	79
4.3.3	<i>De novo</i> transcriptome assembly	80
4.3.4	RSEM for transcript abundance estimation	81
4.3.5	Differential expression analysis using EdgeR: Identification of Differentially expressed transcripts between pairs of samples	82
4.3.6	Functional annotation and KEGG pathway analysis	82
4.3.7	CDC prediction	82
4.4	Results & Discussion	83
4.4.1	Paired-end sequencing and <i>de novo</i> assembly	83
4.4.2	Characterization by similarity search	83
4.4.3	Functional characterization by GO annotation	87
4.4.4	Functional classification by KEGG	90
4.4.5	Analysis of genes encoding important functional properties in <i>G. antarctica</i> PI12 during stress conditions	91
	a. Aminoacyl tRNA biosynthesis	92

	b. Biosynthesis of unsaturated fatty acids	92
	c. Antifreeze proteins	94
	d. Cryoprotectants	95
	e. Oxidative stress-related genes	95
	f. Heat shock proteins	96
4.5	Conclusion	97

**CHAPTER 5: Global Transcriptomic Profile And Gene Expression
Patterns Of *Glaciozyma antarctica* PI12 In Response To
Cold Shocks Using Minimal Medium**

5.1	Summary	98
5.2	Background study	99
5.3	Materials & Methods	100
	5.3.1 Maintenance and growth of <i>Glaciozyma antarctica</i> PI12	100
	5.3.2 RNA extraction and sequencing	100
	5.3.3 Analysis of Illumina transcriptome sequencing results	101
	5.3.4 Abundance estimation using RSEM	101
	5.3.5 Differential expression analysis using EdgeR: Identification of differentially expressed transcripts between pairs of samples	101
	5.3.6 Functional annotation and KEGG pathway analysis	101
	5.3.7 CDS prediction	102
5.4	Results	102
	5.4.1 Cold response of <i>G. antarctica</i> PI12	102
5.5	Discussion	106
	5.5.1 Early Cold Response (ECR)	106
	5.5.2 Early Freeze Response (EFR)	108
	5.5.3 Late Cold Response (LCR)	109
	5.5.4 Late Freeze Response (LFR)	111
	5.5.5 Time-Dependent Stress Response	112
	a Early Time-Dependent Response	113
	b Late Time-Dependent Response	115
5.6	General cold stress responses	115
5.7	Conclusion	116

CHAPTER 6: GENE EXPRESSION PATTERNS OF *Glaciozyma antarctica* PI12 IN RESPONSE TO COLD- AND HEAT-SHOCKS

6.1	Summary	118
6.2	Background	119
6.3	Materials & Methods	120
	6.3.1 Maintenance and growth of <i>Glaciozyma antarctica</i> PI12	120
	6.3.2 RNA extraction and sequencing	120
	6.3.3 Analysis of Illumina transcriptome sequencing results	121
	6.3.4 Abundance estimation using RSEM	121
	6.3.5 Differential expression analysis using EdgeR: Identification of differentially expressed transcripts between pairs of samples	121
	6.3.6 Functional annotation and KEGG pathway analysis	122
	6.3.7 CDS prediction	122
	6.3.8 Gene ontology and functional categorization of differentially expressed genes	122
6.4	Results & Discussion	123
	6.4.1 Transcript-level changes in response to a changing temperature	123
	6.4.2 Temperature shifts response of <i>G. antarctica</i> PI12	127
	6.4.3 Early Cold Responses (ECR)	129
	6.4.4 Late Cold Responses (LCR)	130
	6.4.5 Early Freeze Responses (EFR)	131
	6.4.6 Late Freeze Responses (LFR)	132
	6.4.7 Early Time-Dependent Responses	136
	6.4.8 Late Time-Dependent Responses	138
	6.4.9 The heat responses in <i>Glaciozyma antarctica</i> PI12	139
	6.4.10 Early Heat Responses (Moderate Temperature, 16°C)	140
	6.4.11 Late Heat Responses (Moderate Temperature, 16° C)	141
	6.4.12 Early Heat Responses (20°C)	141
	6.4.13 Late Heat Responses (20°C)	142
	6.4.14 Heat Shock Time-dependent Responses	143

6.4.15	Analysis Of temperature-reciprocal transcriptional responses to high and low temperatures	143
6.5	Conclusion	148
CHAPTER 7: CONCLUSION		
7.1	Summary	150
7.2	Characterization of <i>Glaciozyma antarctica</i> PI12	152
	7.2.1 The doubling time of <i>Glaciozyma antarctica</i> PI12	152
	7.2.2 Aerobic and anaerobic of <i>Glaciozyma antarctica</i> PI12	154
	7.2.3 Microscopic analysis of <i>Glaciozyma antractica</i> PI12	154
	7.2.4 Molecular analysis of <i>Glaciozyma antarctica</i> PI12	155
7.3	Transcriptomic analysis of <i>Glaciozyma antarctica</i> PI12	156
	7.3.1 De novo transcriptome assembly of <i>Glaciozyma antarctica</i> PI12	157
	7.3.2 Gene expression of <i>Glaciozyma antarctica</i> PI12	158
	a. Cold shock response of <i>Glaciozyma antarctica</i> PI12	159
	b. Heat shock response of <i>Glaciozyma antarctica</i> PI12	160
	7.3.3 Advantages of RNA-seq	161
	7.3.4 Limitation of RNA-seq	163
7.4	The adaptation model of <i>Glaciozyma antarctica</i> PI12	165
7.5	Future perspectives of the study	170
7.6	Closing remarks	171
REFERENCES		176
APPENDICES		201

LIST OF TABLES

	Page
Table 2.1 Psychrophilic yeasts found in Antarctica	8
Table 2.2 Psychrophilic Basidiomycota found in Antarctica	10
Table 2.3 Psychrophilic microorganisms whose genomes have been sequenced (Margesin, 2009)	12
Table 2.4 Cold adapted enzymes	13
Table 2.5 Cold shock proteins	15
Table 2.6 Cold shock gene in <i>S. cerevisiae</i> (Takayuki et al., 2003)	16
Table 2.7 Oxidative stress defense systems (Jamieson, 1998)	22
Table 2.8 Yeast strains and their growth temperatures	30
Table 2.9 Summary of the protein encoding genes annotated in the genome of <i>G. antarctica</i> PI12	34
Table 3.1 Cloning ligation reaction	47
Table 3.2 Primers for LP7 amplification	48
Table 3.3 PCR master mix for amplification	48
Table 3.4 PCR conditions for amplification	49
Table 4.1 Gene encoding delta-9 fatty acid desaturases in <i>G. antarctica</i> PI12	93
Table 5.1 Total number of expressed transcripts	105
Table 6.1 Summary of the number of DEGs found under different cold and heat shock condition	124
Table 6.2 List of Early Time-dependent Responses DEGs	136
Table 6.3 List of Late Time-dependent Responses DEGs	138
Table 6.4 Environmental Stress Gene (ESG) found in <i>G. antarctica</i> PI12 under cold- and heat-shock	145

LIST OF FIGURES

	Page
Figure 2.1	Image obtained from Hashim et al. (2013) 30
Figure 3.1	The growth of <i>G. antarctica</i> PI12 in Yeast Peptone Dextrose (YPD) at 12°C shaking at 180rpm for 7 days 50
Figure 3.2	The growth of <i>G. antarctica</i> PI12 in Yeast Peptone Dextrose (YPD) at 12°C shaking at 180rpm for 6 days 51
Figure 3.3	The growth of <i>G. antarctica</i> PI12 in Yeast Peptone Dextrose (YPD) containing 15% (v/v) glycerol at -5oC and -7oC for 5 months 52
Figure 3.4	The cell count of <i>G. antarctica</i> PI12 using hemocytometer 54
Figure 3.5	The growth curve of <i>G. antarctica</i> PI12 at 12°C 55
Figure 3.6	The respiratory effect on the growth rate of <i>G. antarctica</i> PI12 in Yeast Peptone Dextrose (YPD) at 12°C shaking at 180rpm for 5 days 57
Figure 3.7	Standard deviation histogram of the respiratory effect on the growth rate of <i>G. antarctica</i> PI12 in Yeast Peptone Dextrose (YPD) at 12°C shaking at 180rpm for 5 days 58
Figure 3.8	Cell division occurred after 24 hours of incubation at 12°C 59
Figure 3.9	Cell division occurred after 48 hours of incubation at 12°C 60
Figure 3.10	Cell division occurred after 72 hours of incubation at 12°C 61
Figure 3.11	Scanning of <i>G. antarctica</i> PI12 after 72 hours of incubation at 12°C 62
Figure 3.12	Scanning of <i>G. antarctica</i> PI12 after 120 hours of incubation at 12°C 63
Figure 3.13	Illustration of the cell division and nucleic division in <i>G. antarctica</i> PI 12 64
Figure 3.14	Illustration of the length, diameter, area size, and perimeter of <i>G. antarctica</i> PI12 65
Figure 3.15	Genomic DNA of <i>G. antarctica</i> PI12 on 0.7% of 20 cm long agarose gel 66
Figure 3.16	Optimized DNA fragmentation 68
Figure 3.17	Fragmentized Rolling Circle Amplified (RCA) DNA fragments 69

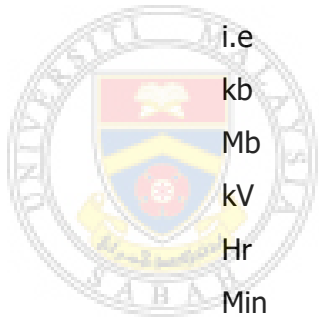
	on 0.7% agarose gel	
Figure 3.18	Digested fragments of <i>G. antarctica</i> PI12 from pJET plasmid with <i>BGLII</i> on 0.7% agarose gel	70
Figure 3.19	Sequencing chromatogram of the fragment of <i>G. antarctica</i> PI12 DNA molecule (776 bp) (LPF7)	71
Figure 3.20	DNA fragments after PCR amplification on 0.7% agarose gel using FP1 and RP	72
Figure 3.21	DNA fragments after PCR amplification on 0.7% agarose gel using FP2 and RPs	73
Figure 3.22	DNA fragment after the treatment of RNase on 0.7% agarose gel	74
Figure 3.23	DNA fragment after the treatment of Dnase on 0.7% agarose gel	75
Figure 4.1	<i>De novo</i> transcriptome assembly workflow	81
Figure 4.2	BioAnalyzer electrophoresis file run summary	84
Figure 4.3	BioAnalyzer electrophoresis file run summary	85
Figure 4.4	BioAnalyzer electrophoresis file run summary	86
Figure 4.5	Distribution of e-value, sequence length and mean similarity from Blast2GO analysis.	87
Figure 4.6	Gene ontology (GO) distribution based on biological process, molecular function, and cellular component in <i>G. antarctica</i> PI12	88
Figure 4.7	Species distribution of <i>G. antarctica</i> PI12 UTs from Trinity assembly	89
Figure 4.8	GO Ontology in <i>G. antarctica</i> PI12	90
Figure 5.1	Gene encoding delta-9 fatty acid desaturases in <i>G. antarctica</i> PI12	103
Figure 5.2	Venn diagram of the total number of DEGs found in <i>G. antarctica</i> PI12	113
Figure 6.1	GO classification of DEGs under cold shock conditions	125
Figure 6.2	GO classification of DEGs under heat shock conditions	126
Figure 6.3	The interaction of PH domain in P13K/AKT pathway and Ras-Raf-Mek-Erk pathway and the downstream regulation in <i>Glaciozyma antarctica</i> PI12 during cold shock	135

Figure 7.1	Summary of the growth and the number of DEGs	166
Figure 7.2	The adaptation model of <i>Glaciozyma antarctica</i> PI12	168



LIST OF SYMBOLS AND ABBREVIATIONS

G	-	Gram
Mg	-	Miligram
µg	-	Microgram
°C	-	Degree Celsius
L	-	Liter
ml	-	mililiter
µl	-	Microliter
Amp	-	Ampicilin
<i>Kar^r</i>	-	Geneticin Resistance Gene
Bp	-	Basepair
DNA	-	Deoxyribonucleic acid
dNTP	-	Deoxynucleotide triphosphate
<i>et al.</i>	-	<i>et alia</i> (and others)
i.e	-	That is
kb	-	Kilobase
Mb	-	megabase
kV	-	Kilovolt
Hr	-	Hour
Min	-	Minute
S	-	second
M	-	Molar
O.D	-	Optical density
PCR	-	Polymerase chain reaction
Rpm	-	Revolutions per minute
TBE	-	Tris borate EDTA
v/v	-	Volume per volume
w/v	-	Weight per volume
Nm	-	Nanometer



UMS
UNIVERSITI MALAYSIA SABAH

LIST OF APPENDICES

	Page
Appendix A Stress inducible genes identified from <i>G. antarctica</i> PI12 <i>de novo</i> transcriptome	201
Appendix B Cell doubling time measurement using OD600nm	215
Appendix C DEGs at -12°C for 6 hours using MM	217
Appendix D DEGs at -12°C for 24 hours using MM	219
Appendix E DEGs at 0°C for 6 hours using MM	222
Appendix F DEGs at 0°C for 24 hours using MM	224
Appendix G DEGs at 0°C for 6 hours using YPD	229
Appendix H DEGs at 0°C for 24 hours using YPD	232
Appendix I DEGs at -12°C for 6 hours using YPD	235
Appendix J DEGs at -12°C for 24 hours using YPD	240
Appendix K DEGs at 16°C for 6 hours using YPD	243
Appendix L DEGs at 16°C for 24 hours using YPD	245
Appendix M DEGs at 20°C for 6 hours using YPD	246
Appendix N DEGs at 20°C for 24 hours using YPD	250
Appendix O Cell measurement	251
Appendix P Sequence of the fragment of <i>G. antarctica</i> PI12 DNA molecule (776bp) (LPF7)	254
Appendix Q Multiplexing DNA oligonucleotide sequences	255

CHAPTER 1

INTRODUCTION

1.1 Preamble

Cold adapted microorganisms are excellent candidates to provide the understanding of molecular adaptations of a cell towards extreme conditions. Psychrophilic and psychrotrophic microorganisms are first referred to those cold adapted bacteria (Morita, 1975). However, the term is now generally refers to those organisms capable to survive, and proliferate at extremely cold condition. Psychrophiles show metabolic fluxes, which are comparable with those exhibited by mesophiles at moderate temperatures (Mihaela *et al.*, 2009). Moreover, the enzymes produce by the psychrophiles offer more novel opportunities for biotechnological applications (Zhao *et al.*, 2011).

In order to gain a better understanding of the cold adaptation of psychrophiles, more than 10 cold adapted microorganisms genome have been sequenced to achieve that purpose. Among the sequenced genomes are, *Colwellia psychrerythraea*, *Desulfotalea psychrophila*, *Methanococcoides burtonii*, *Methonagenium frigidum*, *Polaribacter filamentus*, *Polaribacter irgensii*, *Pseudoalteromonas haloplanktis* TAC125, *Psychrobacter arcticus* 273-4, *Psychrobacter cryohalolentis* K5 and *Psychromonas ingrahamii* (Auman *et al.*, 2006; Bakermans *et al.*, 2006; Corien *et al.*, 2009; Gosink *et al.*, 1998; Jeroen *et al.*, 1999; Medigue *et al.*, 2005; methe *et al.*, 2005; Rabus *et al.*, 2004 & Sauders *et al.*, 2003). The genome of the yeast used in this study, *Glaciozyma antarctica* PI12, also has been sequenced using the Roche, 454 and Illumina sequencing platforms. The genome size is about 2.2 million base pairs, sorted into 21 scaffolds, which consisted of a total of 7857 genes. About 10% of the genes found in the genome of *G. antarctica* PI12 are known to be novel genes. Some cold active and adaptation genes such as α -amylase (Ramli *et al.*, 2013), β -mannanase (Parvizpour *et*

al., 2014), antifreeze protein 1 (Hashim *et al.*, 2013), and antifreeze glycopeptides (AFGP) (Shah *et al.*, 2012) have been identified and cloned.

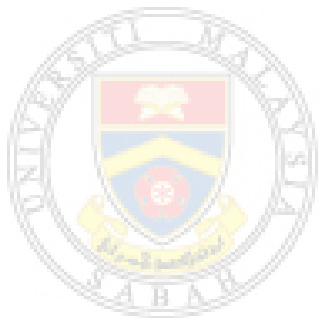
Margesin (2009) highlighted several cold adaptation strategies that are common among psychrophilic microorganisms, including changes in amino acid copiousness that favor protein mobility; production of RNA and protein chaperones; desaturated membrane lipids; expression of cold shock protein; and increasing the cell wall elasticity. Generally, cold adaptations are grouped into three categories: 1) control of molecular motion, 2) resource efficiency, and 3) temperature-specific alleles (Margesin, 2009). However, there are many more genes that are involved in thermal adaptation that have yet been identified.

The genome sequence data per se will not provide information on genes that are expressed during cold adaptation. Therefore, RNA-seq or transcriptomic sequencing can provide further information of genes that are involved in cold adaptation. The recent RNA-seq, or known as deep RNA sequencing, is based on the NGS (next generation sequencing) technology. RNA-seq analysis can be carried out with or without the genome information (Feng *et al.*, 2012).

The objectives of this study are to determine the adaptation mechanisms and strategies of *Glaciozyma antarctica* PI12 to thermal stresses, and to characterize the physiological profile of *G. antarctica* PI12. This thesis is divided into three chapters to address the above objectives. First, the characterization of *G. antarctica* PI12 will be determined based on the growth at its optimal growth temperature at 12°C, the growth at sub-zero temperatures -12°C, -7°C and -5°C, the doubling time of *G. antarctica* PI12 using cell counter, aerobic and anaerobic of *G. antarctica* PI12 and microscopic analyses based on fluorescent microscope and also Scanning Electron Microscope (SEM) to observe *G. antarctica* PI12 cell division and bud division. Moreover, the molecular techniques, such as genomic analysis and transcriptomic analysis will be applied to *G. antarctica* PI12. Transcriptomic analysis using RNA-seq is set to determine the genes that are involved during thermal stresses adaptation of *G. antarctica* PI12.

1.2 Objectives

1. To determine the growth rate, doubling time, anaerobic and aerobic effects of *G. antarctica* PI12 at their optimum growth temperature at 12°C,
2. To characterize the morphological features of *G. antarctica* PI12 using Scanning Electron Microscope (SEM),
3. To establish a *de novo* transcriptomic database of *G. antarctica* PI12,
4. To identify the differential gene expression (DEG) patterns of *G. antarctica* PI12 in respond to various temperature shifts using Minimal Medium (MM),
5. To identify the differential gene expression (DEG) patterns of *G. antarctica* PI12 in respond to various temperature shifts using Yeast Peptone Dextrose (YPD).



UMS
UNIVERSITI MALAYSIA SABAH

CHAPTER 2

LITERATURE REVIEW

2.1 Antarctica, the extreme niche

Antarctica is known to be the world's largest continent, with the area size around 14 million km². The continent is covered by two massive ice sheets, namely the East Antarctica, and the West Antarctica ice sheets. The two gigantic ice sheets are separated by a 3, 500 km long range, known as the Transantarctic Mountain, it is also known to be the largest ice-free area in the continent of Antarctica (~23, 000 km²).

Antarctica is the coldest region on Earth, this is due to the rarefied solar radiation expose to the continent, only 16% of that solar radiation at equatorial region is exposed to Antarctica. Not only that, the high average surface elevation surface of the ice sheets, which in most of the places exceed 4, 000 m. To date, the lowest temperature recorded in Antarctica was -89.4°C at Vostok (Krause & Flood, 1997).

Despite the fact it is the coldest region on Earth, some of the areas receive thermal increment based on the geographical differences in climate. It also depends on: 1) the length of the thaw period; 2) the length of the thaw period, and; 3) the number of the thaw day in summer.

2.2 Departure of the continent of Antarctica from the Supercontinent of Gondwana

Antarctica was a part of the supercontinent of Gondwana in more than 170 million years ago. The supercontinent of Gondwana consisted of the continents, which are known as Antarctica, Australia, New Zealand, South America, India and Africa. According to the continental shifting theory, the Gondwana broke apart into seven continents and shifting occurred. Fossils and rocks found in Antarctica also were found in other continents. The finding also suggests that Antarctica was once a much

warmer place before it separated from the Gondwana. It is because when Antarctica was still attached to Gondwana, West Antarctica was partially in the northern hemisphere, and East Antarctica was at the equator (Stonehouse, 2002).

The shifting of the Antarctica continent to the south is a lengthy process. The shifting slowly introducing coldness to the continent as it shifts toward the south. Therefore, all the living organisms that were once lived on this continent undergo multiple natural selections, especially natural selection based on the changing of temperatures. The idea of natural selection proposed by Charles Darwin in year 1859 is that, the organisms that successfully adapt or evolved with certain traits survived, whereas, those organisms failed to adapt were eliminated by the changing environment. Rogers (2007) also stated that a strong natural selection in Antarctica controlled by the environmental factors led to an adaptation of the Antarctic biota.

2.3 Endemism of microorganisms found in Antarctica

According to Cowan *et al.* (2011), even though Antarctica is geographically isolated, it has not been microbiologically isolated. This is because it constantly receives a population of non-indigenous microorganisms, mostly were transported from the southern hemisphere continents by a high altitude aeolian process (Pearce *et al.*, 2009; Hughes & Convey, 2010; Cowan *et al.*, 2011). Nevertheless, there is no quantitative method developed to measure the total inorganic, and organic inputs to the Antarctic, but Cowan *et al.* (2011) assuming the value would be larger per annum, with around 10^{10} - 10^{12} cells for 1 m² x 1-cm deep soil profile of non-indigenous microorganisms can be found (Cowan *et al.*, 2011).

Non-indigenous microorganisms are possibly introduced to Antarctica as an aeolian particle, or the anthropogenic impact (human activity) (Cowan *et al.*, 2011). Aeolian particle capture experiments have demonstrated that most of the non-indigenous microorganism is introduced to Antarctica as an aeolian particle (Pearce *et al.*, 2009; Cowan *et al.*, 2011). For example, the southern oceans generate aerosols, which serve to be a vehicle for transport of marine microorganism and marine aerosol nutrient input in the near-coastal terrestrial of the Antarctic continent (Bokhorst *et al.*, 2007). Besides, a growing number of human visitations to Antarctica are also known as the factor the non-indigenous microorganisms were introduced to Antarctica.