CHARACTERISATION OF THE GENETIC DIVERSITY WITHIN AND AMONG POPULATIONS OF THE ASIAN HORSESHOE CRAB IN SABAH USING CYTOCHROME OXIDASE 1 AND MICROSATELLITE MARKERS



BIOTECHNOLOGY RESEARCH INSTITUTE UNIVERSITY MALAYSIA SABAH 2015

CHARACTERISATION OF THE GENETIC DIVERSITY WITHIN AND AMONG POPULATIONS OF THE ASIAN HORSESHOE CRAB IN SABAH USING CYTOCHROME OXIDASE 1 AND MICROSATELLITE MARKERS

SAI KERISHA A/P KNTAYYA

THESIS SUBMITED FOR THE DEGREE OF MASTER OF SCIENCE

BIOTECHNOLOGY RESEARCH INSTITUTE UNIVERSITY MALAYSIA SABAH 2015

DECLARATION

I hereby declare that this dissection is of my own work except for the quotations and summaries from which the references are fully acknowledge.

8th August 2015



ACKNOWLEDEMENT

First and foremost, I would like to express my gratitude to my supervisor, Assoc. Prof. Dr. Vijay Kumar for his continuous supervision, criticisms, and support throughout this research. I would also like to thank my co-supervisor, Dr Kenneth Rodrigues for always being available to help and keep track of my lab work progress throughout these two and a half years.

I extend my gratefulness to the lab assistants, officers, and staff of the genomic laboratory in Biotechnology Research Institute for all the help and support given in providing me a conducive, comfortable and friendly working environment. Not forgetting my lab mates who have always been around not only to lend a helping hand with my research, but also as companions. I wish you guys all the best in your research and future undertakings!

I would like to especially thank my parents, my sister and dear friends who are close to me at heart. You guys know who you are. The journey would not have been possible if not for your constant encouragement and love. Thank You! I am deeply indebted to my father, who has not only guided me as a father, but also as a guru and teacher throughout my academic career.

To my friendly four-legged companion, I cannot imagine coming home after a stressful day of work and not see you greet me at the door with your wagging tail. Finally to You, Swami for all the love and blessings showered upon me throughout my life.

CERTIFICATION

- NAME : SAI KERISHA A/P KNTAYYA
- MATRIC NO : **PB20118057**
- TITLE : CHARACTERISATION OF THE GENETIC DIVERSITY WITHIN AND AMONG POPULATIONS OF THE ASIAN HORSESHOE CRAB IN SABAH USING CYTOCHROME OXIDASE 1 AND MICROSATELLITE MARKERS
- DEGREE : MASTER OF SCIENCE (CONSERVATION GENETICS)
- VIVA DATE : **13th February 2015**



Signature

ABSTRACT

Today, only four Horseshoe Crab species exists which includes Tachypleus tridentatus, Tachypleus gigas, Carcinoscorpius rotundicauda (found in Malaysian waters) and Limulus polyohemus (found only in American waters). In recent years, a decline in the number of Horseshoe Crabs has occurred due to various phenomena's including habitat degradation which has prompted studies on the genetic architecture and population structure of the Horseshoe crabs. Consequently, this study was carried out using cytochrome oxidase 1 (C01) gene and microsatellite markers to understand the genetic structure of Horseshoe Crabs in Sabah. A total of 86 Horseshoe Crab samples were collected from five districts in Sabah namely Bongawan (21), Kota Kinabalu (15), Kota Belud (26), Kudat (14) and Tuaran (10). Sequence analysis of the cytochorome oxidase 1 (C01) gene identified the existence of three species namely Tachypleus tridentatus, Tachypleus gigas, and Carcinoscorpius rotundicauda when tested for similarity using BLAST. Amplification of 10 microsatellite loci (Tt01, PLbp2-2-2, PLbp11-10-2, TTLR1A, TTLR1B2, PLbp11-9-2, HCMS069, TTLR151, TTLR152, and TTLR121) on *T. gigas*, C. rotundicauda and T. tridentatus was successfully carried out. All loci were observed to be polymorphic except for locus TTLR1A (Kudat) and locus TTLR151 (Tuaran). The number of alleles ranged from 2 to 6 per locus. The mean observed heterozygosities (Ho) and the mean expected heterozygosities (He) were between 0.55 and 0.72, and between 0.40 and 0.59, respectively indicating a satisfactory level of genetic diversity of the T.gigas population in Sabah. Hardy-Weinberg Equilibrium analysis showed deviation with the highest χ^2 value in locus TTLR1A (93.003) and the lowest in locus TTLR151 (10.108). The dendrogram constructed based on Nei's (1978) genetic distance using UPGMA method showed a distinct cluster of samples from all five locations with samples from Bongawan forming one cluster, samples from Kota Kinabalu and Tuaran forming the second cluster, and Kota Belud and Kudat forming the third cluster, indicating consistency with geographical locations of each population. This study shows moderately high levels of genetic diversity of the Horseshoe Crab population in Sabah. However, population density is constantly declining. This research provides molecular information that could be used to implement different conservation strategies for the Horseshoe Crab population in Sabah.

ABSTRAK

PENCIRIAN KEPELBAGAIAN GENETIK DI ANTARA DAN DI DALAM POPULASI BELANGKAS DI SABAH MENGGUNAKAN LOKUS CYTOCHROM OXIDASE 1 DAN DINUKLEOTIDA MIKROSATELIT

Pada masa kini, hanya terdapat empat jenis spesis belangkas yang terdiri daripada Tachypleus tridentatus, Tachypleus gigas, dan Carcinoscorpius rotunticauda yang boleh dijumpai di perairan Malaysia dan Limulus Polyphemus yang hanya terdapat di perairan America. .Sejak kebelakangan ini, kemerosotan bilangan belangkas terjadi disebabkan pelbagai faktor termasuk kemusnahan habitat yang telah mendorong kajian genetik dan struktur populasi belangkas. Kajian ini telah dijalankan menggunakan gen Cytochrome Oxidase 1 (C01) dan petanda-petanda mokrosatelit. Dalam kajian ini, sejumlah 86 sampel belangkas telah diperolehi dari lima daerah di Sabah iaitu Bongawan (21), Kota Kinabalu (15), Tuaran (10), Kota Belud (26) dan Kudat (14). Analisis jujukan gen Cytochrome Oxidase 1 (C01) menunjukkan terdapat tiga spesis belangkas iaitu Tachypleus tridentatus, Tachypleus gigas, and Carcinoscorpius rotundicauda apabila diuji untuk kesamaan menggunakan BLAST. Proses amplifikasi ke atas 10 lokus telah berjaya dilakukan untuk populasi T. gigas, C. rotundicauda dan T. tridentatus di Sabah dan kesemua 10 loki adalah bersifat polimorfik kecuali lokus TTLR1A (Kudat) dan lokus TTLR151 (Tuaran), Bilangan alel yang ditemui adalah di antara dua hingga enam bagi setiap lokus. Nilai min heterozigositi yang dicerap (Ho) dan nilai min heterozigositi iangkaan (He) masing-masing adalah di antara 0.55 dan 0.72, dan di antara 0.40 dan 0.59. Ini meunjukkan tahap kepelbagaian genetic belangkas yang memuaskan di Sabah. Analisis keseimbangan Hardy-Weinberg menunjukkan sisihan dengan nilai x² tertinggi dalam lokus TTLR1A iaitu 93.003 dan nilai x² terendah dalam lokus TTLR151 iaitu 10.108. Dendogram yang dibentuk berdasarkan jarak genetic Nei (1978) dengan menggunakan kaedah UPGMA menunjukkan perkelompokan sampel yang berbeza pada umumnya yang agak selari dengan lokasi geografi setiap populasi. Sampel dari Bongawan membentuk kelompok pertama, sampel dari Kota Kinabalu dan Tuaran membentuk kelompok kedua manakala sampel dari Kota Belud dan Kudat membentuk kelompok ketiga. Hasil kajian ini menunjukkan tahap kepelbagaian genetic belangkas yang agak tinggi di belangkas. Walaubagaimanapun, kepadatan populasi belangkas didapati merosot secara berterusan. Kajian ini berjaya menghasilkan maklumat molekular yang dapat digunakan untuk melaksanakan strategi pemuliharaan ke atas populasi belangkas di Sabah.

TABLE OF CONTENT

CHAPTER 1: INTRODUCTION 1 1.1 Introduction to Research 1 1.2 Issue of Extinction, Conservation Genetics and the Horseshoe 1 1.3 Significance of Study 4 1.4 Objectives 5 CHAPTER 2: LITERATURE REVIEW 6 2.1 General Biology, Ecology & Life History of the Horseshoe Crab 6 2.1.1 Phylogeny, Taxonomy and Distribution 6 2.1.2 Morphology 9 1.3 Diet and Predators 15
1.2 Issue of Extinction, Conservation Genetics and the Horseshoe crab 1 1.3 Significance of Study 4 1.4 Objectives 5 CHAPTER 2: LITERATURE REVIEW 2.1 General Biology, Ecology & Life History of the Horseshoe Crab 6 2.1 General Biology, Ecology & Life History of the Horseshoe Crab 6 2.1.1 Phylogeny, Taxonomy and Distribution 6 2.1.2 Morphology 9 2.1.3 Diet and Predators 15
1.3 Significance of Study 4 1.4 Objectives 5 CHAPTER 2: LITERATURE REVIEW 2.1 General Biology, Ecology & Life History of the Horseshoe Crab 6 2.1 General Biology, Ecology & Life History of the Horseshoe Crab 6 2.1.1 Phylogeny, Taxonomy and Distribution 6 2.1.2 Morphology 9 2.1.3 Diet and Predators 15
1.3 Significance of Study 4 1.4 Objectives 5 CHAPTER 2: LITERATURE REVIEW 2.1 General Biology, Ecology & Life History of the Horseshoe Crab 6 2.1.1 General Biology, Ecology & Life History of the Horseshoe Crab 6 2.1.2 Morphology 9 2.1.3 Diet and Predators 15
2.1General Biology, Ecology & Life History of the Horseshoe Crab62.1.1 Phylogeny, Taxonomy and Distribution62.1.2 Morphology92.1.3 Diet and Predators15
2.1.1 Phylogeny, Taxonomy and Distribution62.1.2 Morphology92.1.3 Diet and Predators15
2.1.4 Habitat172.1.5 Life Cycle and Reproduction20
2.1.5 Life Cycle and Reproduction202.2Importance of the Horseshoe Crab232.2.1 Ecology232.2.2 The Biomedical Industry252.2.3 Other Importance's27
2.3Status and Management of the Horseshoe Crab30
2.4Cytochrome Oxidase 1 (C01) Gene322.5Microsatellite DNA33
2.6Population Genetics of the Horseshoe Crab3338
CHAPTER 3: MATERIALS AND METHODOLOGY 41
3.1Sample Collection413.2DNA Extraction43vii

3.3	CO1 Gene Analysis – Species Identification	44
	3.3.1 COI-PCR Amplification	44
	3.3.2 PCR Product Cloning Using TA Cloning	45
	3.3.3 Plasmid Minipreparation	46
	3.3.4 DNA Sequencing	47
	3.3.5 DNA Barcoding	48
3.4	Microsatellite-PCR Analysis – Population Structure	48
	3.4.1 Microsatellite-PCR Amplification	48
	3.4.2 Fragment Analysis using QIAxel	50
	3.4.3 Hardy-Weinberg Equilibrium	50
	3.4.4 Degree of Heterozygosity, H	51
	3.4.5 Fis Inbreeding Factor	51
	3.4.6 Genetic Distance, Identity and Dendogram	52

CHAPTER 4: RESULTS

4.1 4.2 4.3	DNA Extraction C01 Gene Analysis – Species Identification 4.2.1 C01-PCR Amplification 4.2.2 Cloning and Plasmid Minipreparation 4.2.3 DNA Sequencing and BLAST 4.2.4 Pairwise Nucleotide Difference 4.2.5 ClustalW Dendrogram and Consensus Tree 4.2.6 Haplotype Distribution Microsatellite DNA Analysis – Population Structure 4.3.1 Primer Screening and Optimisation 4.3.2 Fragment Analysis and Allelic Frequencies 4.3.3 Hardy-Weinberg Equilibrium 4.3.4 Degree of Heterozygosity, H 4.3.5 Genetic Distance and Identity	
CHAPTER	5: DISCUSSION	112
5.1 5.2	Species Identification Population Structure	112 114
CHAPTER 6: CONCLUSION 121		

REFERENCES

APPENDIX			
Appendix A	CO1 Sequences	for	Horse

Appendix A	CO1 Sequences for Horseshoe crab Species from GeneBank	145
Appendix B	CO1 Sequence alignment of 86 Horseshoe crab samples	149

LIST OF TABLES

		Page
2.1	Distribution of the four remaining horseshoe Crab species	8
2.2	Organisms known to prey on Horseshoe crabs	16
3.1	Sample collection and collection period of horseshoe crabs	43
3.2	Sequence of forward and reverse primers used in cytochrome	44
	oxidase 1 gene amplification	44
3.3	Cycle sequencing reaction mixture	47
3.4	Primer sequences and annealing temperatures	49
4.1	Top four BLAST hits for <i>Tachypleus gigas</i> (BG5)	57
4.2	Top four BLAST hits for <i>Tachypleus tridentatus</i> (TN5)	58
4.3	Top four BLAST hits for <i>Carcinoscorpius rotundicauda</i> (KK7)	58
4.4	Number of species from each location	59
4.5	GenBank accession numbers and codes used in this study	61
4.6	Percentage pairwise nucleotide differences among horseshoe	64
	crab species	
4.7	Haplotype distribution of <i>T.gigas</i> CO1 region	68
4.8	Haplotype distribution of <i>T. tridentatus</i> CO1 region	69
4.9	Haplotype distribution of <i>C.rotundicauda</i> CO1 region	70
4.10	Number of haplotypes in each species	71
4.11	Number of haplotypes distributed in five locations	74
4.12	Number of <i>T.gigas</i> haplotypes in Bongawan, Kota Belud and	75
P	Kudat	
4.13	Haplotype and nucleotide diversity in horseshoe crabs in Sabah	76
4.14	Optimum annealing temperatures of 10 microsatellite loci used	78
4.15	Allele frequencies for <i>T.gigas</i> samples from Bongawan	91
4.16	Allele frequencies for <i>T. gigas</i> samples from Kota Belud	92
4.17	Allele frequencies for <i>T. gigas</i> samples from Kudat	93
4.18	Overall allele frequencies of <i>T. gigas</i> samples	94
4.19	Allele frequencies for C. rotundicauda samples from Kota Kinabalu	95
4.20	Allele frequencies for <i>T. tridentatus</i> samples from Tuaran	96
4.21	Chi-square test and likelihood ratio for Hardy-Weinberg for 10	
1.21	loci of <i>T. gigas</i> samples from Bongawan	97
4.22	Chi-square test and likelihood ratio for Hardy-Weinberg for 10	00
	loci of <i>T. gigas</i> samples from Kota Belud	98
4.23	Chi-square test and likelihood ratio for Hardy-Weinberg for 10	00
	loci of <i>T. gigas</i> samples from Kudat	99
4.24	Overall Chi-square test and likelihood ratio for Hardy-Weinberg	
	for 10 loci of <i>T. gigas</i> samples from Bongawan, Kota Belud and	100
	Kudat	
4.25	Chi-square test and likelihood ratio for Hardy-Weinberg for 10	101
	loci of <i>C. rotundicauda</i> samples from Kota Kinabalu	101
4.26	Chi-square test and likelihood ratio for Hardy-Weinberg for 10	102
	loci of <i>T. tridentatus</i> samples from Tuaran	102

4.27	Mean expected and observed heterozygosities for <i>T.gigas</i> population in Bongawan	103
4.28	Mean expected and observed heterozygosities for <i>T.gigas</i> population in Kota Belud	104
4.29	Mean expected and observed heterozygosities for <i>T.gigas</i> population in Kudat	105
4.30	Overall mean expected and observed heterozygosities for <i>T.gigas</i> population in Bongawan, Kota Belud and Tuaran	106
4.31	Mean expected and observed heterozygosities for <i>C.</i> <i>rotundicauda</i> population in Kota Kinabalu	107
4.32	Mean expected and observed heterozygosities for <i>T. tridentatus</i> population in Tuaran	108
4.33	Mean F_{IS} inbreeding values of 10 loci in Bongawan, Kota Kinabalu, Tuaran, Kota Belud and Kudat	109
4.34	Nei's original measure of genetic identity and distance of horseshoe crab samples from all locations	110



LIST OF FIGURES

		Page
2.1	Distribution of the four remaining horseshoe crab species	9
2.2	Dorsal view of the horseshoe crab	11
2.3	Ventral view of the horseshoe crab	11
2.4	Side view of the horseshoe crab	13
2.5	Differences among the four horseshoe crab species	14
2.6	<i>T.gigas</i> on a sandy beach during spawning period	19
2.7	<i>C. rotundicauda</i> in a mangrove during spawning period	19
2.8	Spawning activity of the horseshoe crab	20
2.9	Migratory shorebirds feeding on horseshoe crab eggs in New Jersey	24
2.10	Collection of blood (Haemolymph) from horseshoe crabs	26
2.11	Stacks of horseshoe crabs collected to be ground up for fertilizers at Delaware in 1924	29
3.1	Map of Sabah showing five populations from where Horseshoe crab samples were collected	42
3.2	Collection of tissue samples from the horseshoe crab	42
4.1	Horseshoe crab DNA bands observed on a 1.0% agarose gel after using the CTAB method of DNA extraction	53
4.2	PCR product band observed on a 2% agarose gel using	
- 143K	<i>cytochrome oxidase 1</i> (CO1) primers for the Horseshoe crab	54
4.3	White and blue colonies observed on the LB plate with X-Gal	55
4.4	Plasmid DNA bands observed on a 1.0% agarose gel	56
4.5	Consensus tree for 86 horseshoe crab samples using PHYLIP's	60
11	bootstrap method with 1000 replications	
4.6	Sequence alignment of the 12 COI gene nucleotide sequences of the horseshoe crab species	62
4.7	UPGMA Dendogram of the four horseshoe crab species	66
4.8	Number of haplotypes between and among <i>T.gigas</i> , <i>T.tridentatus</i> and <i>C.rotundicauda</i>	72
4.9	Neighbour-Joining (NJ) analysis of Kimura 2 parameter of	72
	genetic distance	73
4.10	Haplotype distribution of <i>T.gigas</i> in Bongawan, Kota Belud and Kudat	76
4.11	Amplification of the 10 microsatellite loci on 2% agarose gel	78
4.12	Amplification of 10 <i>T.gigas</i> loci using primer PLbp11-9-2	79
4.13	Amplification of 10 <i>T.gigas</i> loci using primer HCMS069	79
4.14	Fragment analysis of 10 <i>T.gigas</i> samples from Bongawan using	
	Tt01	81
4.15	Fragment analysis of 10 <i>T.gigas</i> samples from Kota Belud using PLbp2-2-2	82
4.16	Fragment analysis of 10 <i>T.gigas</i> samples from Kota Belud using PLbp11-10-2	83
4.17	Fragment analysis of 10 <i>T.gigas</i> samples from Bongawan using TTLR1A	84
4.18	Fragment analysis of 10 <i>T.gigas</i> samples from Kudat using TTLR1B2	85

4.19	Fragment analyses of 10 <i>T.gigas</i> samples from Kota Belud using PLbp11-9-2	86
4.20	Fragment analyses of 10 T.gigas samples from Kota Belud using HCMS069	87
4.21	Fragment analysis of 10 <i>T.gigas</i> samples from Bongawan using TTLR151	88
4.22	Fragment analysis of 10 <i>T.gigas</i> samples from Kota Belud using TTLR152	89
4.23	Fragment analysis of 10 <i>T.gigas</i> samples from Kota Belud using TTLR121	90
4.24	Dendrogram of horseshoe crab population in Sabah	111



LIST OF ABBREVASIONS AND SYMBOLS

-	Negative
%	Percent
+	Positive
°C	Degree Celsius
μg	Microgram
μL	Microlitre
A	Adenine nucleotide
bp	Base pair
c	Cytosine nucleotide
CBOL	Consortium for the Barcode of Life
COI	Cytochrome oxidase I
ddH ₂ O	Deionised Distilled Water
DNA	Deoxyribonucleic Acid
dNTP	Deoxyribonucleotide-triphosphate
Ed.	Edition
eds.	Editors
EDTA	Ethylenediaminetetraacetic acid
g A	Gram
G 🖉 📕	Guanine nucleotide
He	Expected Heterozygosity
Ho	Observed Heterozygosity
HCI	Hydrochloric Acid
HWE	Hardy-Weinberg Equilibrium
kb	NIODASE PAIL
KCI	Potassium Chloride
LAL	Limulus Amebocyte Lysate
Μ	Molar
mg	Milligram
MgCl ₂	Magnesium Chloride
MgSO₄	Magnesium Sulphate
mL	Millilitre
mM	Millimolar Mite eksendrick DNA
mtDNA	Mitochondrial DNA
	Sodium Chloride
NaOH PCR	Sodium hydroxide Polymerase Chain Reaction
	Page
pg. pM	Picomolar
RNA	Ribonucleic Acid
rpm	Revolutions per minute
s	Seconds
-	

SDS	Sodium dodecyl sulphate
SSRs	Simple Sequence Repeats
т	Thymine nucleotide
TBE	Tris/Borate/EDTA buffer solution
Tm	Melting temperature
Tris	Trishydroxymethylaminomethane
Tris-HCl	Tris-hydrochloric acid
UPGMA	Unweighted Pair Group Method with Arithmetic mean
V	Volts
w/v	Weight over volume
X	Times
h	Haplotype diversity
п	Nucleotide diversity
F _{IS}	F statistics
IUCN	the International Union for the Conservation of Nature
ERDG	Ecological Research and Development Group



CHAPTER 1

INTRODUCTION

1.1 Introduction to Research

Conservation biology is a multidisciplinary science that emphasises biodiversity and its maintenance that has developed as a result of growing awareness of biodiversity loss (Soule, 1985). Conservation genetics in particular, incorporates genetic management of small populations, resolution of taxonomic uncertainties and management units and the application of genetic analysis to preserve and understand species' biology (Frankham, 2003). This research falls in the scope of conservation genetics. The present study aims to understand the population structure of the horseshoe crabs in Sabah using microsatellite markers.

1.2 Issue of Extinction, Conservation Genetics and the Horseshoe crab

Extinction is a natural part of the evolutionary process. For example, the mass extinction at the end of Cretaceous 65 million years ago eliminated much of the previous flora and fauna including the dinosaurs (Cracraft, 2001). Nevertheless, this made way for propagation of the mammals and flowering plants which contributed to the world's biodiversity. In contrast to this, currently the world's biological diversity is rapidly depleting as both direct and indirect consequences of human action and this causes loss of species at rates that far outruns the creation of new species. An unknown but large number of species are already extinct, while many others have reduced population sizes that put them at risk (WCMC, 1992). As a result of this, human intervention is crucial to improve the management and to ensure the survival of many species today.

Conservation genetics is motivated by the need to reduce current rates of extinction and to preserve biodiversity. It is the application of genetics as a tool to preserve species as dynamic entities which are capable of coping with environmental changes. It includes genetic management of populations, resolution of taxonomic uncertainties, defining management units within species, and the use of molecular genetic analysis in forensics and understanding species biology. It also aims to minimize the risk of extinction due to genetic factors.

Horseshoe crabs, also known as living fossils are marine arthropods that are in need of conservation genetic measures. They are facing a rapid decline in the number of populations around the world due to factors such as water pollution, loss of living and spawning habitats and human exploitation (Li *et al.* 2009). Today, only four species of the horseshoe crab exists. Often used as main examples of organisms that survived long periods without any significant changes in their anatomy, horseshoe crabs have puzzled evolutionary biologists for centuries, earning them the name 'living fossils' and 'phylogenetic relicts' (Obst *et al.* 2012). The oldest Horseshoe crab fossil was found in Manitoba, Canada and is 445 million years old (Brut, 2014).

For most of its history, the horseshoe crab was regarded as junk from the sea. Bounties of horseshoe crabs were placed upside down to avoid them from feasting on clams. According to the Atlantic States Marine Fisheries Commission, approximately two million horseshoe crabs were caught yearly to be consumed as fertilizers and livestock feed in the mid 80's. In 1956, Frederik Bang, a John Hopkins biologist discovered that Limulus Amoebocyte Lysate (LAL), refined from the copper-based blood of horseshoe crabs, could be used to detect trace amounts of endotoxin contaminants which is what makes it useful for testing sterility of medical equipment and drugs for human use (Odell et al. 2005).

Aside from the medical benefits, horseshoe crabs also remain as a valuable part of the ecosystem. Many shorebirds, migratory birds, turtles and fish use horseshoe crab eggs as an important part of their diet. In Malaysia, horseshoe crabs are a delicacy in restaurants and command a high price. All this has led to poaching and over-harvesting to feed the market. This, along with onshore development and coastal disturbance is threatening the horseshoe crab population around the world. Since then, this has led to extensive studies on the horseshoe crab.

While there have been several studies done concerning the general biology and life history of the horseshoe crab, information concerning its status and population dynamics is not adequately known (Walls *et al.* 2002). However, there has been indication of a decline in the number of horseshoe crabs in Asia. As of now, India and Japan have listed the horseshoe crab as an endangered species (The Hindu Business Line, 2005). In addition to this, in Asia, horseshoe crabs are only listed under the data deficiency category (IUCN). In Malaysia, there is still a lack of reliable report on the status of the horseshoe crabs (Tan *et al.* 2010). Thus, in order to prevent further decline in the horseshoe crab population, studies on their population genetics is crucial. It will aid in the knowledge of their variations among different populations and later promote the establishment of conservation strategies for the horseshoe crab species in Malaysia.

In all biological research, the identification of species is critical. Accurate identifications reveal known information about each organism, its ecological roles, its physiological and biochemical properties, and its societal risks and benefits (Seifert *et al.* 2006). In addition to this, to the untrained eye, the differences in morphology and taxonomic characteristics of the four horseshoe crab species are subtle (Avise *et al.* 1994) and scientists are unable to distinguish one species from another. Besides that, morphological similarities have caused a struggle in understanding phylogenetic relationships among the horseshoe crab lineages (Xia, 2000).

Species identification, such as for the horseshoe crab in this study, requires the usage of a specific gene region. The region should be able to distinguish different species through polymorphism in the DNA sequences and should have a universal conserved primer region that can be used for different species. The *cytochrome oxidase I* (COI) gene region satisfies this requirement and it has been proven useful in numerous barcoding research (Kress *et al.*, 2005).

Microsatellites, a highly versatile genetic marker which are also known as Simple Sequence Repeats (SSRs) are stretches of short DNA sequence in which a motif is tandemly repeated. Due to its habitually high variability, microsatellites have been a useful marker to address matters such as discrimination, relationships, structure and classification, not only at the population level, but also at the individual level (Wan *et al.* 2004).

In the past two decades, such genetic markers have been used to determine the status of several endangered species and have provided significant understandings that have critically influenced management decisions and created benefits for several species (Wan *et al.* 2004). In this study, the *cytochrome oxidase 1* (CO1) gene region was used in the identification of the horseshoe crab species, while microsatellite markers were used in the assessment of the population structure of the horseshoe crabs in Sabah.

1.3 **Problem Statement**

There is significant decrease in the number of horseshoe crabs in Sabah.

1.4 Significance of Study

Taking into consideration the various interconnected issues affecting the horseshoe crab population and its possibility of becoming an endangered species in Malaysia, developing a successful horseshoe crab management strategy is important. More so, with its proven biomedical benefits, horseshoe crabs need to be protected through proper understanding of its population status encompassing its breeding pattern and its genetic diversity. In order to have conservation management plans, research and documentation of information is vital. This research is one step towards the documentation of scientific information of the horseshoe crab population in Sabah. This kind of information is also important considering the future prospects for Malaysia to start its own biomedical industry in which horseshoe crab bleeding can be carried out for the production of LAL. In this

context, future cultivation of horseshoe crabs can also benefit economically. As such, scientific information of this species has medical, ecological, and economical importance.

1.4 Objectives

The aim of this study is to identify the genetic structure of the wild populations of the horseshoe crabs in Sabah. Three specific objectives were taken to achieve this goal.

- 1. To identify the species of the Asian Horseshoe Crabs in Sabah using the primer sequences for *cytochrome oxidase I* (COI) gene of Asian horseshoe crab.
- 2. To characterize the genetic population structure of the Horseshoe crab populations through DNA profiling of single-locus microsatellite markers.
- 3. To obtain a phylogenetic tree of the horseshoe crab population through microsatellite markers and sequence homology of the *cytochrome oxidase I* (COI) gene.

UNIVERSITI MALAYSIA SABAH

CHAPTER 2

LITERATURE REVIEW

2.1 General Biology, Ecology & Life History of the Horseshoe Crab

2.1.1 Phylogeny, Taxonomy and Distribution

Horseshoe crabs are marine chelicerates closely related to arachnids. Horseshoe crabs are the closest living relatives to the trilobites (Shuster, 1982). They have lived for more than 500 million years since the Palaeozoic Devonian era. The oldest fossil found dates back between 360 and 405 million years ago (Chatterji *et al.*, 1992). The ancestor of the present species of the Horseshoe crab originated from the Mesozoic waters of Europe.

Up to date, only one consensus has been reached in which the Atlantic species (*Limulus polyphemus*) is a sister taxon to the three Indo-Pacific species (*Tachypleus gigas, Tachypleus tridentatus* and *Carcinoscorpius rotundicauda*). The consensus is consistent with the current taxonomy in which *Limulus* belongs to the subfamily Limulinae and the three other extant species belonging to Tachypleinae. All four species of Limulidae is the only living representatives of Merostomata (Yamaksai, 1988).

At present, the phylogenetic relationship between the three Indo-Pacific species remains unsolved despite the multitude of analyses conducted. Results were highly differed, leading to some research suggesting that the three species constitute a phylogenetically irresolvable Trichotomy which could have resulted from all three Indo-Pacific species forming within a short geological time (Avise *et al., 1994*).

Nevertheless, re-examination on phylogenetic relationships of the horseshoe crab species was conducted by Obst *et al.* (2012) using an intra-specific analysis involving 18S rDNA, 28S rDNA and mitochondrial gene cytochrome *c* oxidase I (COI). The results suggest:

- 1) Strong support for a monophyletic genus *Tachypleus* and a diversification of the three Asian species during the Paleogene period. "Speciation events were temporally well-separated by several million years" (Obst *et al., 2012*).
- 2) Tree topology suggests that the "three Asian species originated in central South East Asia from a marine stem group that inhabited the shallow coastal waters between the Andaman Sea, Vietnam, and Borneo".
- 3) "*C. rotundicauda* probably separated from the *Tachypleus* stem group by invading estuarine habitats, while *T. tridentatus* most likely migrated northeast along the Southern coast of China and towards Japan" (Obst *et al., 2012*).

Horseshoe crabs belong to the phylum of Arthropods, consisting of animals that have articulated bodies and limbs. The three major classes of Arthropods are Insects, Arachnids and Crustaceans. The horseshoe crabs belong to its own class called Merostomata. The term merostomata means "legs attached to the mouth". Horseshoe crabs are most closely related to trilobites that existed 544 million years ago (Shuster, 1982). The taxonomic classification of the horseshoe crabs is shown below.

Kingdom	:	Animalia
Phylum	:	Arthropoda
Class	:	Meristomata
Order	:	Xhiphosurida
Family	:	Limulidae
Genus	:	Limulus, Tachypleus, Carcinoscorpius
Species	:	Limulus polyphemus, Tachypleus gigas, Tachypleus tridentatus,
		Carcinoscorpius rotundicauda

The present distribution of Horseshoe crabs suggest they migrated when the shallow seas disappeared while the European land mass was formed. One group moved to the east and one moved to the west (Shuster, 1982). Today, Horseshoe crabs are found in only two regions of the world; the coastal waters of Asia, from India to Japan, and along the Atlantic coastlines of North America (Table 2.1 and Figure 2.1)

Species	Distribution
Limulus polyphemus	Atlantic coast of America and distributed extensively from Maine to the Florida Keys and around the periphery of the Gulf of Mexico up to south of Yucaton Peninsula
Carcinoscorpius rotundicauda	Found on the West coast of Malaysia and Thailand (in the mouth of Mae am River). Also found on the East coast of Orissa, India
Tachypleus gigas	Occurs along both the coasts of Malay Peninsula, around West coast of Singapore and along the coast of Orissa (India) to Indo-China, Borneo, Java (Indonesia), Torres Straits and Celebes
Tachypleus rotundicauda	It is the most common Asian species and found rich in the western and southern coasts of Japan. Also found along the coast of Vietnam, China (South and East coasts), Taiwan, Philippines, North Borneo and in the Indian Ocean side of Sumatra

 Table 2.1 : Distribution of the four remaining horseshoe crab species

Source : Biomedical Potentials of the Indian Horseshoe Crab. INFOFISH International. (Lakshmanan & Venkateshvaran, 1999).

Tachypleus tridentatus and *Carcinoscorpius rotundicauda* are from the kingdom Metazoa, phylum Arthropoda, subphylum Chelicerata, class Merostomata, order Xiphosura, and family Limulidae. *Tachypleus tridentatus* and *Carcinoscorpius rotundicauda* is also known as the Asian horseshoe crab. *Tachypleus tridentatus* is found mostly on the west and south coast of Japan and also found along the coast of Vietnam, south and east coast of China, Taiwan, Philippines and also north of Borneo (Lakshmanan & Venkateshvaran, 1999). *Carcinoscorpius rotundicauda* is found on the west coast of Malaysia, Thailand (in the mouth of Mae Nam river) and also on the east coast of Orissa of India (Lakshmanan & Venkateshvaran, 1999).

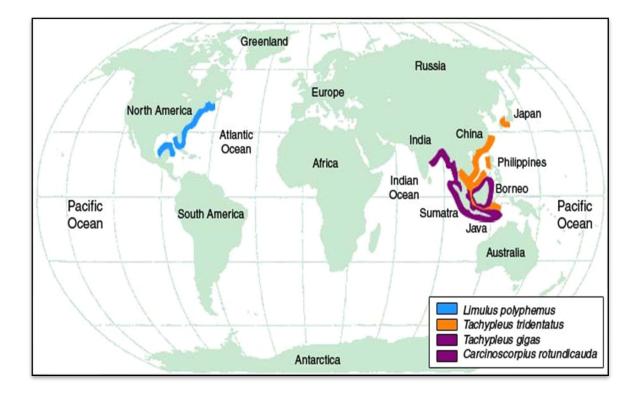


Figure 2.1: Distribution of the four remaining horseshoe crab species.

Source : University of Delaware Graduate College of Marine Studies and the Sea Grant College Program, Project Galathea, 2008.

2.1.2 Morphology

UNIVERSITI MALAYSIA SABAH

One of the most distinguished features of the horseshoe crab is its alien-like body. Despite its appearance, the horseshoe crab is harmless. The body of the horseshoe crab is divided into three regions: the prosoma, the opisthosoma and the teson (Rudkin and Young, 2009). These are sometimes referred to as the cephalothorax, the abdomen, and the tail.

The cephalothorax, also referred to as the carapace or shell, is the large anterior segment of the horseshoe crab. The carapace protects the legs and organs of the horseshoe crab and also keeps the animal upright in rough waters. The prosoma contains an intestinal tract with an esophagus and proventriculus which is used by the crab to grind food. It also contains a nervous system concentrated into a brain, a tubular heart, excretory glands and connective tissues and cartilaginous plates. The