## MICROPROPAGATION OF *Etlingera coccinea* (ZINGIBERACEAE)



## SCHOOL OF SCIENCE AND TECHNOLOGY UNIVERSITI MALAYSIA SABAH 2013

## MICROPROPAGATION OF *Etlingera coccinea* (ZINGIBERACEAE)

NURUL HUMAIRA ABDULLAH THADDEUS

# THESIS SUBMITTED IN PARTIAL FULFILLMENT OF THE REQUIREMENT FOR THE DEGREE OF MASTER OF SCIENCE

## SCHOOL OF SCIENCE AND TECHNOLOGY UNIVERSITI MALAYSIA SABAH 2013

#### PUMS 99:1

### **UNIVERSITI MALAYSIA SABAH**

BORANG PI	ENGESAHAN TESIS
JUDUL :	
IJAZAH :	
SAYA :	SESI PENGAJIAN :
(HURUF BESAR)	
Mengaku membenarkan tesis *(LPSM/Sarjana/Dokto Sabah dengan syarat-syarat kegunaan seperti berikut:	r Falsafah) ini disimpan di Perpustakaan Universiti Malaysia -
<ol> <li>Tesis adalah hak milik Universiti Malaysia Saba</li> <li>Perpustakaan Universiti Malaysia Sabah diben</li> <li>Perpustakaan dibenarkan membuat salinan t tinggi.</li> </ol>	ah. Jarkan membuat salinan untuk tujuan pengajian sahaja. esis ini sebagai bahan pertukaran antara institusi pengajian
4. Sila tandakan (/)	mat yang berdarjah keselamatan atau kepentingan Malaysia
seperti yang termaktu TERHAD (Mengandungi maklu mana penyelidikan di	ıb di AKTA RAHSIA RASMI 1972) mat TERHAD yang telah ditentukan oleh organisasi/badan di jalankan)
TIDAK TERHAD	Disahkan oleh:
 (TANDATANGAN PENULIS) Alamat Tetap:	(TANDATANGAN PUSTAKAWAN)
 TARIKH:	(NAMA PENYELIA) TARIKH:
Catatan: *Potong yang tidak berkenaan. *Jika tesis ini SULIT dan TERHAD, sila lampirkan sur menyatakan sekali sebab dan tempoh tesis ini perlu *Tesis dimaksudkan sebagai tesis bagi Ijazah Dokto bagi pengajian secara kerja kursus dan Laporan Pro	at daripada pihak berkuasa/organisasi berkenaan dengan u dikelaskan sebagai SULIT dan TERHAD. r Falsafah dan Sarjana Secara Penyelidikan atau disertai ıjek Sarjana Muda (LPSM).

### DECLARATION

I hereby declare that the material in this thesis is my own except for quotations, excerpts, equations, summaries and references, which have been duly acknowledged.

15 August 2013

Nurul Humaira Abdullah Thaddeus PS20068491



### CERTIFICATION

- NAME : NURUL HUMAIRA ABDULLAH THADDEUS
- MATRIC NO : **PS20068491**
- TITLE : MICROPROPAGATION OF ETLINGERA COCCINEA (ZINGIBERACEAE)
- DEGREE : MASTER OF SCIENCE (BIOTECHNOLOGY)
- VIVA DATE :4 JUNE 2013



#### ACKNOWLEDGEMENT

All the praise is the Almighty Allah, the Merciful and the Compassionate. Due to His willingness, the completion of this study was made possible. First of all, I would like to express my deep sense of gratitude to my research supervisor, Prof. Madya Dr. Jualang Azlan Gansau, for his guidance, constant support and valuable ideas, useful advises, reviewing of the thesis dissertation and for the pertinent suggestions to improve it. I also gratefully acknowledge for his patience with all about my mess to complete my thesis dissertation, for his patient listening and for the freedom of work he provided from the very beginning and all throughout my research work. Thanks also to Prof. Madya Dr. Baba Musta and Dr. Zaleha Abd. Aziz for gave me a meaningful chance to complete my study.

It is my pleasure to offer my special appreciation to my husband Mr. Rusphin Mohd.Asyraf for his deep understanding, patience and sacrifices, financial and moral support, useful advice and a lot of favor. Not forgot to my pair of twins, Farwez Hayyan and Farweza Hana who always by my side.

I am very grateful to my parents Thaddeus Gousi and Rimi Gurisang, my parents in law Mahidin Amat and Lamiah Samadal for their encouragement and moral support throughout my work. My gratitude also towards my sister Mrs. Nurhidayah and husband Mr. Subir Musta for being helpful, to my sister in law Mrs. Muhina and husband Mr. Sali who help me gathering the plant material, Mrs. Kasirani and husband Mr. Japius, Mrs. Haini and husband Mr. Lajimin, Hajah Aineh and husband Hj. Ibnu, Mrs. Shahidah and husband Mr. Kasa, Mrs. Zaida and husband Mr. Jasri, my brother in law Mr. Zulhimat and wife Mrs. Jati, Mr. Rymin and wife Mrs. Muna, and Ms. Adiba for their understanding support as well give me a hand to care my babies as I am in critical moment during this study. To all my brothers, Mark Donny, Kennedy, Mark Ronald, Bonny Face, Stanley, Frankie, Herman and Steve. It was a very special moments we have through.

To my collegue, Aina Syafiqa, Yanteh, Hartini, Roslina, Devina, Ainul, Rosmah, Cyril, Xcelom, Judy, Bong, Bella and Ozie for valuable suggestion and their help in the lab. My thanks also to lab assistants of Biotechnology Programme, Mrs. Mary for her cooperation during the work in the lab. It is impossible to thank all those separately who are responsible for this day as it is dependent on several yesterdays. I am personally thankful to all those known and unknown faces that directly or indirectly helped me during the phase of work towards my thesis dissertation. The financial support in the form of research fellowship by Universiti Malaysia Sabah (UMS) is duly acknowledged.

#### ABSTRACT

#### MICROPROPAGATION OF *Etlingera coccinea* (ZINGIBERACEAE)

An effective protocol was developed for *in vitro* regeneration of valuable food additive plant Etlingera coccinea (zingiberaceae) through different methods of plant tissue culture. Micropropagation was possible via shoot regeneration from rhizome bud explants, callus induction and proliferation derived from leaf explants, plantlets regeneration from callus culture and shoot multiplication through pseudo stem explants. This study was involved the effect of Plant Growth Regulators (PGRs), types and strengths of basal media, and types and concentrations of carbon sources. The results obtained showed that plant growth factors manipulation as stated markedly influence shoot regeneration, callus induction and proliferation, plantlets regeneration from callus culture, and shoot multiplication of *Etlingera coccinea*. Rhizome buds were sterilized and cultured on Murashige and Skoog (MS) basal medium containing 0.1-2.0 uM of thidiazuron (TDZ), benzylaminopurine (BAP) or kinetin (KIN) supplied with 3% (w/v) of sucrose. High percentage of shoot regeneration (100%) was achieved by  $(\sim 1\pm 0.5 \text{ cm})$  of the whole bud cultured on MS medium containing 1.0  $\mu$ M TDZ after 20 weeks of culture. Shoots were transferred to medium containing 5.0 µM BAP for shoot elongation and multiplication. High frequency of rooting (90%) was observed on medium supplemented with 1.0 µM IBA. The maximum percentage (100%) of callus formation was obtained after 10 weeks of culture from in vitro leaf explants cultured on half strength MS basal medium supplemented with 5.0 µM 2,4dichlorophenoxyacetic acid (2,4-D) and 10 µM Benzylaminopurine (BAP). In media selection, half strength MS medium was most suitable for callus induction while 3% (w/v) of sucrose was the best carbon source among other sugar types tested. Callus cell proliferation was rapidly increased on MS basal medium supplemented with 2.5 µM 2,4-D within 5 weeks of culture. Combination of 2.5 µM 2,4-D and 5.0 µM BAP improved callus proliferation. 3% (w/v) of sucrose was most effective for callus proliferation and long term maintenance of callus culture. Plantlets regeneration with highest frequency of shoot formation (88%) from callus culture were obtained on MS medium containing 2.5 µM BAP and 0.1 µM IAA with a mean of 5.5 and 7.0 shoots and roots per explant after 6 weeks of culture. The most responsive explants were observed on full strength MS basal medium while 3% (w/v) of sucrose was the best source of carbon among other carbon sources tested. Roots regeneration were observed along with the shoot formation, 2.5 µM BAP was superior to KIN and ZEA on shoot multiplication from pseudo stem explants. Combination of 2.5 µM BAP and 0.7 µM NAA enhance shoots regeneration percentage up to 100% over a 6 weeks period of culture and produced 12.0 shoots per explant with 100% of rooting percentage. Full strength of MS basal medium and 3% (w/v) of sucrose were the best supplements for shoot multiplication via pseudo stem explants. Micropropagated plantlets were grown healthy after transplanted to the pots containing a mixture of 3:1 (w/w) soil: sand with 85% of survival explants after 6 month of transplantation. The procedure reported here offers a potential system for micropropagation of Etlingera coccinea and expected to be useful in improvement and conservation of this species in the future.

#### ABSTRAK

Kaedah terbaik regenerasi secara in vitro bagi tumbuhan berkepentingan makanan Etlingera coccinea (zingiberaceae) telah dikaji melalui pelbagai kaedah kultur tisu tumbuhan. Mikropropagasi telah dilakukan melalui regenerasi pucuk daripada eksplan tunas rizom, induksi dan proliferasi kalus daripada eksplan daun, regenerasi plantlet daripada kultur kalus, dan penambahbilangan pucuk daripada eksplan menyerupai batang. Kajian ini melibatkan kesan pengawalaturan tumbuhan, jenis dan kepekatan medium, serta jenis dan kepekatan sumber karbon. Hasil kajian yang diperolehi menunjukkan manipulasi faktor pertumbuhan yang dinyatakan mempengaruhi regenerasi pucuk, induksi dan proliferasi kalus, regenerasi plantlet daripada kultur kalus, dan penambahbilangan pucuk Etlingera coccinea. Tunas rizom disterilkan dan dikulturkan ke dalam medium MS yang mengandingi 0.1-2.0 µM TDZ, BAP atau KIN secara berasingan dan dibekalkan dengan 3% (w/v) sukrosa. Peratus tertinggi regenerasi pucuk (100%) diperolehi daripada tunas rizom bersaiz (~1±0.5 cm) tanpa pembelahan, selepas 20 minggu dikulturkan dalam medium MS yang mengandungi 1.0 µM TDZ. Mikropucuk dipindahkan ke dalam medium yang mengandungi 5.0 µM BAP untuk pemanjangan dan proliferasi pucuk. Frekuensi tertinggi pertumbuhan akar (90%) didapati dalam medium vang dibekalkan 1.0 uM IBA. Peratus maksimum (100%) pembentukan kalus diperolehi daripada eksplan daun in vitro selepas 10 minggu dikulturkan dalam 1/2 kepekatan medium MS yang mengandungi 5.0 µM asid diklorofenoksiasetik (2,4-D) dan 10 µM benzilaminopurina (BAP). Dalam pemilihan media, 1/2 kepekatan medium MS didapati paling sesuai untuk induksi kalus manakala 3% (w/v) kepekatan sukrosa membekalkan sumber karbon yang terbaik. Proliferasi sel kalus meningkat dengan cepat dalam tempoh 5 minggu dalam medium MS yang dibekalkan 2.5 µM 2,4-D. Kombinasi 2.5 µM 2,4-D dan 5.0 µM BAP menambah peningkatan proliferasi kalus. 3% (w/v) sukrosa adalah paling efektif untuk proliferasi kalus da<mark>n pengekala</mark>n kultur kalus dalam jangka masa panjang. Regenerasi plantlet dengan frekuensi tertinggi pembentukkan pucuk (88%) daripada kultur kalus diperolehi dalam medium MS yang mengandungi 2.5 µM BAP dan 0.1 µM IAA dengan min 5.5 dan 7.7 pucuk dan akar per eksplan selepas 6 minggu dikulturkan. Tindakbalas eksplan yang terbaik didapati dalam medium MS pada kepekatan penuh manakala 3% (w/v) sukrosa membekalkan sumber karbon yang terbaik. Regenerasi akar terbentuk bersamaan dengan pembentukkan pucuk. 2.5 µM BAP mengatasi KIN dan ZEA dalam penambahbilangan pucuk daripada eksplan menyerupai batang. Kombinasi 2.5 µM BAP dan 0.7 µM NAA meningkatkan peratus regenerasi pucuk kepada 100% selama 6 minggu pengkulturan dan menghasilkan 12.0 pucuk per eksplan dengan 100% pembentukkan akar. Medium MS pada kepekatan penuh dan 3% (w/v) sukrosa adalah suplemen terbaik untuk penambahbilangan pucuk daripada eksplan menyerupai batang. Plantlet yang dipropagasikan bertumbuh dengan baik selepas dipindahkan ke dalam pasu yang mengandungi 3:1 (w/w) tanah:pasir dengan 85% eksplan berkembang selepas 6 bulan pemindahan. Kaedah yang dijelaskan ini menawarkan sistem mikropropagasi yang berpotensi untuk perkembangan dan pemuliharaan spesies Etlingera coccinea pada masa hadapan.

### TABLE OF CONTENTS

Ρ	a	q	e
	-	_	_

TIT	LE		i
DEC	LARATI	ON	ii
CER		TION	iii
ACK		DGEMENTS	iv
ABS	RACT		v
ABS	STRAK		vi
TAB	LE OF C	CONTENTS	vii
LIS	T OF TA	BLES	xii
LIS	T OF FIC	GURE	xvi
LIS	T OF PL	ATES	xviii
LIS	T OF AB	BREVIATIONS	xxiii
LIS	T OF SY	MBOLS	xxiv
GLC	<b>SSARY</b>		xxv
KEYWORDS			xxvii
	AA		
CH/	PTER 1	: INTRODUCTION	1
	1	UNIVERSITI MALAYSIA SABAH	
1.1	Introdu	ction	1
CHA	PTER 2	: LITERATURE REVIEW	
2.1	The Far	nily Zingiberaceae	4
2.2	The <i>Eth</i>	ingera coccinea	5
2.3	Importa	ance and Economical Potential of Etlingera coccinea	7
2.4	Previou	s Study on Micropropagation of Zingiberaceae Species	9
	2.4.1	Establishment of aseptic culture	9
	2.4.2	Callus induction and plant regeneration	12
	2.4.3	Shoot multiplication	14
	2.4.4	In vitro rooting	15
	2.4.5	Hardening	16

Factors	Affecting Micropropagation	16
2.5.1	Effect of culture medium	16
2.5.2	Effect of Plant Growth Regulators (PGRs)	21
2.5.3	Effect of carbon sources	27
PTER 3	: MATERIALS AND METHODS	29
Establi	ishment of Aseptic Culture and Shoot Regeneration	29
From F	Rhizome Bud Explants	
3.1.1	Plant materials and sterilization method	29
3.1.2	Media preparation	30
3.1.3	Culture conditions	30
3.1.4	Effect of TDZ, BAP and kinetin concentrations on shoot	30
	regeneration	
3.1.5	Effect of explants size on shoot regeneration	30
3.1.6	Effect of explants sectioning on shoot regeneration	31
3.1.7	Effect of BAP concentrations on shoot elongation	32
3.1 <mark>.8</mark>	Effect of IBA and NAA concentrations on rooting of shoots	32
3.1.9	Data collection and statistical analysis	32
3.1.10	Experimental design IVERSIII MALAYSIA SABAR	33
3.1.11	Maintenance of stock culture	33
Callus	Induction of Leaf Explants	34
3.2.1	Plant materials	34
3.2.2	Media preparation	34
3.2.3	Explants preparation, culturing method and	34
	growth conditions	
3.2.4	Effect of Plant Growth Regulators (PGRs) on	34
	callus induction of leaf explants	
3.2.5	Effect of basal media on callus induction of leaf explants	35
3.2.6	Effect of carbon sources on callus induction of leaf explants	35
3.2.7	Data observation and statistical analysis	35
	Factors 2.5.1 2.5.2 2.5.3 <b>PTER 3</b> <b>Establ</b> <b>From 1</b> 3.1.1 3.1.2 3.1.3 3.1.4 3.1.5 3.1.6 3.1.7 3.1.6 3.1.7 3.1.8 3.1.7 3.1.8 3.1.9 3.1.10 3.1.10 3.1.10 3.1.11 <b>Callus</b> 3.2.1 3.2.2 3.2.3 3.2.4 3.2.5 3.2.6 3.2.7	<ul> <li>Factors Affecting Micropropagation</li> <li>2.5.1 Effect of culture medium</li> <li>2.5.2 Effect of Plant Growth Regulators (PGRs)</li> <li>2.5.3 Effect of carbon sources</li> </ul> APTER 3 : MATERIALS AND METHODS Establishment of Aseptic Culture and Shoot Regeneration From Rhizome Bud Explants 3.1.1 Plant materials and sterilization method 3.1.2 Media preparation 3.1.3 Culture conditions 3.1.4 Effect of TDZ, BAP and kinetin concentrations on shoot regeneration 3.1.5 Effect of explants size on shoot regeneration 3.1.6 Effect of explants size on shoot regeneration 3.1.7 Effect of BAP concentrations on shoot regeneration 3.1.8 Effect of IBA and NAA concentrations on rooting of shoots 3.1.9 Data collection and statistical analysis 3.1.10 Experimental design 3.1.11 Maintenance of stock culture Callus Induction of Leaf Explants 3.2.2 Media preparation 3.2.3 Explants preparation, culturing method and growth conditions 3.2.4 Effect of Plant Growth Regulators (PGRs) on callus induction of leaf explants 3.2.5 Effect of axis on callus induction of leaf explants 3.2.6 Effect of carbon sources on callus induction of leaf explants 3.2.7 Data observation and statistical analysis

	3.2.8	Experimental design	36
3.3	Callus	Proliferation Derived From Leaf Explants	36
	3.3.1	Plant materials	36
	3.3.2	Media preparation	36
	3.3.3	Explants preparation, culturing method and growth conditions	36
	3.3.4	Effect of 2,4-D concentrations on callus proliferation	37
	3.3.5	Effect of 2,4-D and BAP combinations on callus proliferation	37
	3.3.6	Effect of sucrose concentrations on callus proliferation	37
	3.3.7	Effect of sucrose concentrations on long term maintenance	38
		of callus culture	
	3.3.8	Callus mass measurement and statistical analysis	38
	3.3.9	Experimental design	38
	150		
3.4	Plantle	ts Regeneration from Callus Culture	38
	7		
K	3.4 <mark>.1</mark>	Plant materials	38
	3.4.2	Media preparation	38
	3.4.3	Explants preparation, culturing method and	38
		growth conditions	
	3.4.4	Effect of BAP, kinetin and zeatin on plantlets regeneration	39
		from callus culture	
	3.4.5	Effect of BAP and kinetin with combinations of NAA or IAA	39
		on plantlets regeneration from callus culture	
	3.4.6	Effect of basal media on plantlets regeneration from	39
		callus culture	
	3.4.7	Effect of carbon sources on plantlets regeneration	39
		from callus culture	
	3.4.8	Data collection and statistical analysis	40
	3.4.9	Experimental design	41

Shoot	multiplication Trough Pseudo Stem Explants	41
3.5.1	Plant materials	41
3.5.2	Media preparation	41
3.5.3	Explants preparation, culturing method and growth conditions	41
3.5.4	Effect of BAP, kinetin and zeatin concentrations on shoot	41
	multiplication	
3.5.5	Effect of BAP with combinations of NAA, IBA and IAA on	42
	shoot multiplication	
3.5.6	Effect of basal media on shoot multiplication	42
3.5.7	Effect of carbon sources on shoot multiplication	42
3.5.8	Data collection and statistical analysis	42
3.5.9	Experimental design	43
Preli	minary Study On Plantlets Acclimatization	43
B		
PTER 4	: RESULTS AND DISCUSSION	44
2		
Establ	ishment of Aseptic Culture and Shoot Regeneration	44
from F	UNIVERSITI MALAYSIA SABAH	
4.1.1	Effect of TDZ, BAP and kinetin concentrations on shoot	44
	regeneration	
4.1.2	Effect of explants size on shoot regeneration	53
4.1.3	Effect of explants sectioning on shoot regeneration	58
4.1.4	Effect of BAP concentrations on shoot elongation	62
4.1.5	Effect of IBA and NAA concentrations on rooting of shoots	67
Callus	Induction of Leaf Explants	71
4.2.1	Effect of Plant Growth Regulators (PGRs) on callus induction	71
	of leaf explants	
4.2.2	Effect of basal media on callus induction of leaf explants	85
4.2.3	Effect of carbon sources on callus induction of leaf explants	89
	Shoot 3.5.1 3.5.2 3.5.3 3.5.4 3.5.5 3.5.6 3.5.7 3.5.8 3.5.9 Preli Preli 4.1.1 4.1.2 4.1.3 4.1.4 4.1.5 Callus 4.2.1 4.2.2 4.2.3	<ul> <li>Shoot multiplication Trough Pseudo Stem Explants</li> <li>3.5.1 Plant materials</li> <li>3.5.2 Media preparation</li> <li>3.5.3 Explants preparation, culturing method and growth conditions</li> <li>3.5.4 Effect of BAP, kinetin and zeatin concentrations on shoot multiplication</li> <li>3.5.5 Effect of BAP with combinations of NAA, IBA and IAA on shoot multiplication</li> <li>3.5.6 Effect of basal media on shoot multiplication</li> <li>3.5.7 Effect of arbon sources on shoot multiplication</li> <li>3.5.8 Data collection and statistical analysis</li> <li>3.5.9 Experimental design</li> </ul> PTER 4: RESULTS AND DISCUSSION PTER 4: RESULTS AND DISCUSSION A.1.1 Effect of TDZ, BAP and kinetin concentrations on shoot regeneration 4.1.2 Effect of explants size on shoot regeneration 4.1.3 Effect of Explants size on shoot regeneration 4.1.4 Effect of BAP concentrations on shoot regeneration 4.1.5 Effect of BAP concentrations on shoot shoots Callus Induction of Leaf Explants 4.2.1 Effect of Plant Growth Regulators (PGRs) on callus induction of leaf explants 4.2.2 Effect of basal media on callus induction of leaf explants 4.2.3 Effect of carbon sources on callus induction of leaf explants

4.3	Callus	Proliferation Derived From Leaf Explants	94
	4.3.1	Effect of 2,4-D concentrations on callus proliferation	94
	4.3.2	Effect of 2,4-D and BAP combinations on callus proliferation	100
	4.3.3	Effect of sucrose concentrations on callus proliferation	105
	4.3.4	Effect of sucrose concentrations on long term maintenance	111
		of callus culture	
4.4	Plantle	ets Regeneration from Callus Culture	117
	4.4.1	Effect of BAP, kinetin and zeatin concentrations on plantlets	117
		regeneration from callus culture	
	4.4.2	Effect of BAP and kinetin with combinations of NAA or IAA on	123
		plantlets regeneration from callus culture	
	4.4.3	Effect of basal media on plantlets regeneration from callus	130
	A	culture	
	4.4.4	Effect of carbon sources on plantlets regeneration from	135
	9 L	callus culture	
	1 -		
4.5	Shoot	Multiplication through Pseudo Stem Explants	143
	4.5.1	Development stages of shoot multiplication through SABA-	143
		pseudo stem explants	
	4.5.2	Effect of BAP, kinetin and zeatin concentrations on	145
		shoot multiplication	
	4.5.3	Effect of BAP with combinations of NAA, IBA and IAA	153
		on shoot multiplication	
	4.5.4	Effect of basal media on shoot multiplication	160
	4.5.5	Effect of carbon sources on shoot multiplication	166
4.6	Prelimi	inary Study On Plantlets Acclimatization	172
СНА	PTER 5	: CONCLUSION	175

#### 4.3 Callus Proliferation Derived From Leaf Explants

REFFERENCES		177
APPENDICES		208
APPENDIX 1	NUTRIENTS COMPOSITION OF DIFFERENT BASAL MEDIA	209
APPENDIX 2	PLANT CULTURE SUPPLEMENTS	212
APPENDIX 3	PREPARATION OF PGRS STOCK SOLUTIONS	214



#### LIST OF TABLES

Page

- Table 4.1Effect of BAP, TDZ and kinetin concentrations on shoot49regeneration of *Etlingera coccinea* rhizome buds after 20weeks cultured on MS basal medium supplemented with 3%(w/v) of sucrose, 0.8% (w/v) agar, pH 5.8 and grown at 25±2°C with 16h photoperiod.
- Table 4.2Effect of explants size on shoot regeneration of *Etlingera*56coccinea rhizome buds after 20 weeks cultured on MS basal<br/>medium supplemented with 3% (w/v) of sucrose, 0.8%<br/>(w/v) agar, pH 5.8 and grown at 25±2°C with 16h<br/>photoperiod
- Table 4.3Effect of explants sectioning of  $\sim 1.0 \pm 0.5$  cm explant on<br/>shoot regeneration of *Etlingera coccinea* rhizome buds after<br/>20 weeks cultured on MS basal medium supplemented with<br/>3% (w/v) of sucrose, 0.8% (w/v) agar, pH 5.8 and grown at<br/> $25\pm2^{\circ}$ C with 16h photoperiod
- Table 4.4Effect of BAP concentrations on elongation of shoots after 665weeks cultured on MS basal medium supplemented with 3%(w/v) of sucrose, 0.8% (w/v) agar, pH 5.8 and grown at 25±2°C with 16h photoperiod65
- Table 4.5Effect of IBA and NAA concentrations on rooting of shoots<br/>after 6 weeks cultured on MS basal medium supplemented<br/>with 3% (w/v) of sucrose, 0.8% (w/v) agar, pH 5.8 and<br/>grown at 25±2°C with 16h photoperiod69
- Table 4.6Effect of auxins (2,4-D, NAA, IBA and IAA) concentrations on<br/>callus induction of leaf explants after 20 weeks cultured on<br/>MS basal medium supplemented with 3% (w/v) of sucrose,<br/>0.8% (w/v) agar, pH 5.8 and grown at 25±2°C in the dark<br/>condition77
- Table 4.7The effect of cytokinins (BAP, kinetin, TDZ and zeatin)78concentrations on callus induction of *Etlingera coccinea* leaf<br/>explants after 20 weeks cultured on MS basal medium<br/>supplemented with 3% (w/v) of sucrose, 0.8% (w/v) agar,<br/>pH 5.8 and grown at 25±2°C in the dark condition
- Table 4.8The effect of 2,4-D with combination of BAP or kinetin on<br/>callus induction of *Etlingera coccinea* leaf explants after 20<br/>weeks cultured on MS basal medium supplemented with 3%<br/>(w/v) of sucrose, 0.8% (w/v) agar, pH 5.8 and grown at<br/>25±2°C in the dark condition

xiii

- Table 4.9The effect of NAA with combination of BAP or kinetin on<br/>callus induction of *Etlingera coccinea* leaf explants after 20<br/>weeks cultured on MS basal medium supplemented with 3%<br/>(w/v) of sucrose, 0.8% (w/v) agar, pH 5.8 and grown at<br/>25±2°C in the dark condition
- Table 4.10The effect of different basal media containing 5.0  $\mu$ M 2,4-D87and 10.0 $\mu$ MBAP on callus induction of *Etlingera coccinea*leaf explants after 20 weeks cultured on MS basal mediumsupplemented with 3% (w/v) of sucrose, 0.8% (w/v) agar,<br/>pH 5.8 and grown at 25±2°C in the dark condition
- Table 4.11The effect of different carbon sources on callus induction of<br/>*Etlingera coccinea* leaf explants after 20 weeks cultured on<br/>MS basal medium supplemented with 5.0 μM 2,4-D and<br/>10.0μM BAP 0.8% (w/v) agar, pH 5.8 and grown at 25±2°C<br/>in the dark condition92
- Table 4.12The effect of BAP, kinetin and zeatin concentrations on<br/>plantlets regeneration from callus culture of *Etlingera*<br/>*coccinea* on MS basal medium supplemented with 3% (w/v)<br/>of sucrose, 0.8% (w/v) agar, pH 5.8 and grown at 25±2°C<br/>with 16h photoperiod after 6 weeks of culture120
- Table 4.13The effect of BAP and kinetin with combination of various<br/>concentrations of NAA and IAA individually on plantlets<br/>regeneration from callus culture of *Etlingera coccinea* on MS<br/>basal medium supplemented with 3% (w/v) of sucrose,<br/>0.8% (w/v) agar, pH 5.8 and grown at 25±2°C with 16h<br/>photoperiod after 6 weeks of culture127
- Table 4.14 The effect of media types and nutrients supplement on 133 plantlets regeneration from callus culture of *Etlingera coccinea* supplemented with 2.5  $\mu$ M BAP with combination of 0.1  $\mu$ M IAA, 3% (w/v) of sucrose, 0.8% (w/v) agar, pH 5.8 and grown at 25±2°C with 16h photoperiod after 6 weeks ofculture.
- Table 4.15The effect of carbon sources on plantlets regeneration from<br/>callus culture of *Etlingera coccinea* on MS basal medium<br/>supplemented with 2.5  $\mu$ M BAP with combination of 0.1  $\mu$ M<br/>IAA, 0.8% (w/v) agar, pH 5.8 and grown at 25±2°C with<br/>16h photoperiod after 6 weeks of culture.
- Table 4.16The effect of BAP, kinetin and zeatin concentrations on shoot151multiplication through pseudo stem explants of Etlingera<br/>coccinea after 6 weeks culture period on MS basal medium<br/>supplemented with 3% (w/v) of sucrose, 0.8% (w/v) agar,<br/>pH 5.8 and grown at 25±2°C with 16h photoperiod

- Table 4.17The effect of BAP with combination of various concentrations158of NAA, IBA and IAA on shoot multiplication through pseudo<br/>stem explants of *Etlingera coccinea* after 6 weeks culture<br/>period on MS basal medium supplemented with 3% (w/v) of<br/>sucrose, 0.8% (w/v) agar, pH 5.8 and grown at 25±2°C<br/>with16h photoperiod
- Table 4.18The effect of different basal media on shoot multiplication<br/>through pseudo stem explants of *Etlingera coccinea* after 6<br/>weeks culture period supplemented with 2.5  $\mu$ M BAP and 0.7<br/> $\mu$ M NAA, 3% (w/v) of sucrose, 0.8% (w/v) agar, pH 5.8 and<br/>grown at 25±2°C with 16h photoperiod
- Table 4.19The effect of different carbon sources on shoot multiplication170through pseudo stem explants of *Etlingera coccinea* after 6weeks culture period on MS medium supplemented with 2.5 $\mu$ M BAP and 0.7  $\mu$ M NAA, 0.8% (w/v) agar, pH 5.8 andgrown at 25±2°C with 16h photoperiod
- Table 4.20*Ex-vitro* performance of *Etlingera coccinea* plantlets after 3173months of transplantation.



#### LIST OF FIGURES

- Figure 4.1 The effect of 2,4-D concentrations on growth of callus 98 [based on fresh weight (FW)] derived from leaf explants of *Etlingera coccinea* on MS basal medium supplemented with 3% (w/v) of sucrose, 0.8% (w/v) agar, pH 5.8 and grown at 25±2°C with 16h photoperiod within 5 weeks of culture
- Figure 4.2 The effect of 2,4-D concentrations on growth of callus 99 [based on dry weight (DW)] derived from leaf explants of *Etlingera coccinea* on MS basal medium supplemented with 3% (w/v) of sucrose, 0.8% (w/v) agar, pH 5.8 and grown at 25±2°C with 16h photoperiod within 5 weeks of culture
- Figure 4.3 The effect of 2,4-D and BAP combinations on growth of 103 callus [based on fresh weight (FW)] derived from leaf explants of *Etlingera coccinea* on MS basal medium supplemented with 3% (w/v) of sucrose, 0.8% (w/v) agar, pH 5.8 and grown at 25±2°C with 16h photoperiod within 5 weeks of culture.
- Figure 4.4 The effect of 2,4-D and BAP combinations on growth of 104 callus [based on dry weight (DW)] derived from leaf explants of *Etlingera coccinea* on MS basal medium supplemented with 3% (w/v) of sucrose, 0.8% (w/v) agar, pH 5.8 and grown at 25±2°C with 16h photoperiod within 5 weeks of culture
- Figure 4.5 The effect of sucrose concentrations on growth of callus 108 [based on fresh weight (FW)] derived from leaf explants of *Etlingera coccinea* on MS basal medium supplemented with 0.8% (w/v) agar, pH 5.8 and grown at 25±2°C with 16h

Page

photoperiod within 7 weeks of culture.

- Figure 4.6 The effect of sucrose concentrations on growth of callus 109 [based on dry weight (DW)] derived from leaf explants of *Etlingera coccinea* on MS basal medium supplemented with 0.8% (w/v) agar, pH 5.8 and grown at 25±2°C with 16h photoperiod within 7 weeks of culture.
- Figure 4.7 The effect of sucrose concentrations on growth of callus 114 based on fresh weight (FW) for consecutively 12 months with 12 subcultures on MS basal medium supplemented with 0.8% (w/v) agar, pH 5.8 and grown at 25±2°C with 16h photoperiod.
- Figure 4.8 The effect of sucrose concentrations on growth of callus 115 based on dry weight (DW) for consecutively 12 months with 12 subcultures on MS basal medium supplemented with 0.8% (w/v) agar, pH 5.8 and grown at 25±2°C with 16h photoperiod.

## UNIVERSITI MALAYSIA SABAH

### LIST OF PLATES

		Page
Plate 2.1	Nature of <i>Etlingera coccinea</i> in the field (Picture taken in the field from Kg. Kemburongoh Ranau, Sabah.	6
Plate 2.2	Inflorescences of Etlingera coccinea.	6
Plate 2.3	Fresh 'tuhau' after remove the outer scales (contained parts of rhizomes and leafy shoots)	7
Plate 2.4	Products of Etlingera coccinea.	8
Plate 3.1	(A) The young rhizome of <i>Etlingera coccinea</i> ; (B) Sprout bud of rhizome with outer scales; (C) Bud after removing outer scales.	29
Plate 3.2	(A) 1 week old rhizome bud ( $\sim$ 1±0.5cm); (B) 2 weeks old rhizome bud ( $\sim$ 3±0.5cm); (C) 3 weeks old rhizome bud ( $\sim$ 5±0.5cm)	31
Plate 3.3	(A) Non-sliced of aseptic single bud (Bar: 1mm) ;(B) Single bud cut into two half slices (Bar: 1mm); (C) Single bud cut into three slices (Bar: 1mm)	31
Plate 3.4	Explant preparation from leaf derived callus of <i>Etlingera coccinea</i> . (A) Callus derived from leaf explant (Bar: 1 mm) (B) Callus removed from leaf explant (Bar: 1 mm) (C) Callus dissect into approx. 0.5 x 0.5 cm size (Bar: 0.5 mm)	37
Plate 4.1	Shoot regeneration of <i>Etlingera coccinea</i> rhizome buds within 20 weeks cultured on MS basal medium supplemented with various concentrations of TDZ, $3\%$ (w/v) of sucrose, $0.8\%$ (w/v) agar, pH 5.8 and grown at $25\pm2^{\circ}$ C with 16h photoperiod	50
Plate 4.2	Shoot regeneration of <i>Etlingera coccinea</i> rhizome buds within 20 weeks cultured on MS basal medium supplemented with various concentrations of BAP, $3\%$ (w/v) of sucrose, 0.8% (w/v) agar, pH 5.8 and grown at $25\pm2^{\circ}$ C with 16h photoperiod	51
Plate 4.3	Shoot regeneration of <i>Etlingera coccinea</i> rhizome buds within 20 weeks cultured on MS basal medium supplemented with various concentrations of kinetin, $3\%$ (w/v) of sucrose, 0.8% (w/v) agar, pH 5.8 and grown at $25\pm2^{\circ}$ C with 16h photoperiod	52

- Plate 4.4 Effect of explants size on adventitious shoots regeneration of 57 *Etlingera coccinea* rhizome buds within 20 weeks cultured on MS basal medium supplemented with 1.0 μM TDZ, 3% (w/v) of sucrose, 0.8% (w/v) agar, pH 5.8 and grown at 25±2°C with 16h photoperiod
- Plate 4.5Effect of explants sectioning on adventitious shoots61regeneration of *Etlingera coccinea* rhizome buds within 20weeks cultured on MS basal medium supplemented with 1.0μM TDZ, 3% (w/v) of sucrose, 0.8% (w/v) agar, pH 5.8 andgrown at 25±2°C with 16h photoperiod
- Plate 4.6 Shoot elongation of *Etlingera coccinea* within 6 weeks 66 cultured on MS basal medium supplemented with different concentrations of BAP, 3% (w/v) of sucrose, 0.8% (w/v) agar, pH 5.8 and grown at 25±2°C with 16h photoperiod
- Plate 4.7 Roots formation of *Etlingera coccinea* after 6 weeks cultured 70 on MS basal medium treated with different concentrations of IBA and NAA individually, 3% (w/v) of sucrose, 0.8% (w/v) agar, pH 5.8 and grown at 25±2°C with 16h photoperiod
- Plate 4.8 Morphology of leaf explants of *Etlingera coccinea* after 20 79 weeks treated on different concentrations of 2,4-D, NAA, IBA and IAA alone on MS basal medium supplemented with 3% (w/v) of sucrose, 0.8% (w/v) agar, pH 5.8 and grown at 25±2°C in the dark condition
- Plate 4.9 Morphology of leaf explants of *Etlingera coccinea* after 20 80 weeks treated on different concentrations of BAP, kinetin,TDZ and zeatin alone on MS basal medium supplemented with 3% (w/v) of sucrose, 0.8% (w/v) agar, pH 5.8 and grown at 25±2°C in the dark condition
- Plate 4.10 Stages of callus formation from leaf explant of *Etlingera* 83 *coccinea*
- Plate 4.11 Morphology of leaf explants of *Etlingera coccinea* after 4 84 weeks treated on different combinations of 2,4-D or NAA with BAP or kinetin on MS basal medium supplemented with 3%(w/v) of sucrose, 0.8% (w/v) agar, pH 5.8 and grown at 25±2°C in the dark condition
- Plate 4.12 Morphology of leaf explants of *Etlingera coccinea* after 4 88 weeks treated on different basal media supplemented with 5.0  $\mu$ M 2,4-D and 10.0 $\mu$ M BAP, 3% (w/v) of sucrose, 0.8% (w/v) agar, pH 5.8 and grown at 25±2°C in the dark condition

- Plate 4.13 Development of callus formation from leaf explants of 93 *Etlingera coccinea* within 20 weeks of culture period treated with different carbon sources on MS basal medium supplemented with 5.0 μM 2,4-D and 10.0μM BAP, 0.8% (w/v) agar, pH 5.8 and grown at 25±2°C in the dark condition
- Plate 4.14 Callus morphology in control treatment and different 97 concentrations of 2,4-D on MS basal medium with 3% (w/v) of sucrose, 0.8% (w/v) agar, pH 5.8 and grown at 25±2°C with 16h photoperiod after 5 weeks culture
- Plate 4.15 Callus morphology in control treatment and different 102 combinations of 2,4-D and BAP on MS basal medium with 3% (w/v) of sucrose, 0.8% (w/v) agar, pH 5.8 and grown at 25±2°C with 16h photoperiod after 5 weeks culture
- Plate 4.16 Morphology of callus treated with different concentrations of 110 sucrose on MS basal medium supplemented with 2.5  $\mu$ M 2,4-D and 5.0  $\mu$ M BAP, 0.8% (w/v) agar, pH 5.8 and grown at 25±2°C with 16h photoperiod after 6 weeks culture
- Plate 4.17 Morphology of callus treated with different concentrations of sucrose on MS basal medium supplemented with 2.5  $\mu$ M 2,4-D and 5.0  $\mu$ M BAP, 0.8% (w/v) agar, pH 5.8 and grown at 25±2°C with 16h photoperiod after 10 months of culture with 10 subcultures
- Plate 4.18 Development stages of plantlets regeneration from callus 121 culture of *Etlingera coccinea* within 20 weeks of culture on MS basal medium supplemented with 2.5 µM BAP, 3% (w/v) of sucrose, 0.8% (w/v) agar, pH 5.8 and grown at 25±2°C with 16h photoperiod
- Plate 4.19 Regenerated plantlets from callus culture of *Etlingera* 122 *coccinea* after 20 weeks of culture treated with different cytokinins on MS basal medium supplemented with 3% (w/v) of sucrose, 0.8% (w/v) agar, pH 5.8 and grown at 25±2°C with16h photoperiod. Bar: (A-D) 2.0 cm.
- Plate 4.20 Development stages of plantlets regeneration from callus 128 culture of *Etlingera coccinea* within 20 weeks of culture on MS basal medium treated with 2.5 μM BAP and 0.1 μM IAA and 3% (w/v) of sucrose, 0.8% (w/v) agar, pH 5.8 and grown at 25±2°C with 16h photoperiod
- Plate 4.21 Regenerated plantlets from callus culture of *Etlingera* 129 *coccinea* after 20 weeks of culture treated with different combinations of cytokinin and auxin on MS basal medium supplemented with 3% (w/v) of sucrose, 0.8% (w/v) agar, pH 5.8 and grown at 25±2°C with 16h photoperiod.

- Plate 4.22 Development stages of plantlets regeneration of leaf derived 134 callus of *Etlingera coccinea* within 20 weeks of culture on MS basal medium supplemented with 2.5 µM BAP and 0.1 µM IAA and 3% (w/v) of sucrose, 0.8% (w/v) agar, pH 5.8 and grown at 25±2°C with 16h photoperiod
- Plate 4.23 Regenerated plantlets from leaf derived callus of *Etlingera* 134 *coccinea* after 20 weeks of culture treated on MS and B5 basal medium supplemented with 3% (w/v) of sucrose, 0.8% (w/v) agar, pH 5.8 and grown at 25±2°C with 16h photoperiod; (A) MS basal medium; (B) B5 basal medium.
- Plate 4.24 Development stages of plantlets regeneration from callus 141 culture of *Etlingera coccinea* treated with 3% sucrose within 20 weeks of culture on MS basal medium supplemented 0.8% (w/v) agar, pH 5.8 and grown at 25±2°C with 16h photoperiod
- Plate 4.25 Regenerated plantlets from callus culture of *Etlingera* 142 *coccinea* after 20 weeks of culture treated with different carbon sources on MS basal medium supplemented 0.8% (w/v) agar, pH 5.8 and grown at 25±2°C with 16h photoperiod.
- Plate 4.26 Development stages of multiple shoots formation from pseudo stem explants of *Etlingera coccinea* within 6 weeks cultured on MS basal medium supplemented with 2.5 µM of BAP, 3% (w/v) of sucrose, 0.8% (w/v) agar, pH 5.8 and grown at 25±2°C with 16h photoperiod.
- Plate 4.27 Multiple shoots formation from pseudo stem explants of 152 *Etlingera coccinea* after 6 weeks cultured on MS basal medium supplemented with various concentrations of BAP, kinetin and zeatin, 3% (w/v) of sucrose, 0.8% (w/v) agar, pH 5.8 and grown at 25±2°C with 16h photoperiod.
- Plate 4.28 Multiple shoots formation from pseudo stem explants of 159 *Etlingera coccinea* after 6 weeks cultured on MS basal medium supplemented with 2.5 μM BAP with combination of various concentrations of NAA, IBA and IAA, 3% (w/v) of sucrose, 0.8% (w/v) agar, and pH 5.8 and grown at 25±2 ° C with 16h photoperiod.
- Plate 4.29 Multiple shoots formation from pseudo stem explants of 165 *Etlingera coccinea* after 6 weeks cultured treated on different basal media supplemented with 2.5 μM BAP and 0.7 μM NAA, 3% (w/v) of sucrose, 0.8% (w/v) agar, pH 5.8 and grown at 25±2°C with 16h photoperiod.

- Plate 4.30 Multiple shoots formation from pseudo stem explants of 171 *Etlingera coccinea* after 6 weeks cultured treated with different carbon sources on MS basal medium supplemented with 2.5  $\mu$ M BAPand 0.7  $\mu$ M NAA, 0.8% (w/v) agar, pH 5.8 and grown at25±2°C with 16h photoperiod.
- Plate 4.31 Acclimatization stages of *Etlingera coccinea* plantlets. (A) *In* 174 *vitro* derived plantlets transferred to the plastic cup and exposed to external condition; (B) Rooting plantlet; (C) Plantlets after transferred to the pot and covered with the sterilized plastic; (D-F) Plantlets within 1-3 months growing in the mixture of 3:1 (w/w) soil: sand.

