

**CHEMOPREVENTIVE EFFECTS OF SELECTED  
HERBAL PLANTS AGAINST CARBON  
TETRACHLORIDE-MEDIATED OXIDATIVE  
TISSUE DAMAGE IN RATS**



**KOH PEI HOON**

UNIVERSITI MALAYSIA SABAH

**BIOTECHNOLOGY RESEARCH INSTITUTE  
UNIVERSITI MALAYSIA SABAH  
2011**

**CHEMOPREVENTIVE EFFECTS OF SELECTED  
HERBAL PLANTS AGAINST CARBON  
TETRACHLORIDE-MEDIATED OXIDATIVE  
TISSUE DAMAGE IN RATS**

**KOH PEI HOON**



**UMS**

**THESIS SUBMITTED IN FULFILLMENT FOR  
THE DEGREE OF MASTER OF SCIENCE**

**BIOTECHNOLOGY RESEARCH INSTITUTE  
UNIVERSITI MALAYSIA SABAH  
2011**

## UNIVERSITI MALAYSIA SABAH

## BORANG PENGESAHAN TESIS

JUDUL : \_\_\_\_\_

\_\_\_\_\_

\_\_\_\_\_

IJAZAH : \_\_\_\_\_

\_\_\_\_\_

SAYA : \_\_\_\_\_ SESI PENGAJIAN : \_\_\_\_\_

(HURUF BESAR)

Mengaku membenarkan tesis \*(LPSM/Sarjana/Doktor Falsafah) ini disimpan di Perpustakaan Universiti Malaysia Sabah dengan syarat-syarat kegunaan seperti berikut:-

1. Tesis adalah hak milik Universiti Malaysia Sabah.
2. Perpustakaan Universiti Malaysia Sabah dibenarkan membuat salinan untuk tujuan pengajian sahaja.
3. Perpustakaan dibenarkan membuat salinan tesis ini sebagai bahan pertukaran antara institusi pengajian tinggi.
4. Sila tandakan (/)

SULIT (Mengandungi maklumat yang berdarjah keselamatan atau kepentingan Malaysia seperti yang termaktub di AKTA RAHSIA RASMI 1972)

TERHAD (Mengandungi maklumat TERHAD yang telah ditentukan oleh organisasi/badan di mana penyelidikan dijalankan)

TIDAK TERHAD

Disahkan oleh:

\_\_\_\_\_  
(TANDATANGAN PENULIS)\_\_\_\_\_  
(TANDATANGAN PUSTAKAWAN)

Alamat Tetap: \_\_\_\_\_

\_\_\_\_\_

\_\_\_\_\_

\_\_\_\_\_  
(NAMA PENYELIA)

TARIKH: \_\_\_\_\_

TARIKH: \_\_\_\_\_

## Catatan:

\*Potong yang tidak berkenaan.

\*Jika tesis ini SULIT dan TERHAD, sila lampirkan surat daripada pihak berkuasa/organisasi berkenaan dengan menyatakan sekali sebab dan tempoh tesis ini perlu dikelaskan sebagai SULIT dan TERHAD.

\*Tesis dimaksudkan sebagai tesis bagi Ijazah Doktor Falsafah dan Sarjana Secara Penyelidikan atau disertai bagi pengajian secara kerja kursus dan Laporan Projek Sarjana Muda (LPSM).

## DECLARATION

I hereby declare that the material in this thesis is of my own effort, except for quotations, excerpts, equations, references and summaries, which have been duly acknowledged and cited clearly its sources.

15 AUGUST 2011

---

KOH PEI HOON  
PB2009-8022



UMS  
UNIVERSITI MALAYSIA SABAH

## CERTIFICATION

NAME : **KOH PEI HOON**  
MATRIC NO : **PB2009-8022**  
TITLE : **CHEMOPREVENTIVE EFFECTS OF SELECTED HERBAL  
PLANTS AGAINST CARBON TETRACHLORIDE-  
MEDIATED OXIDATIVE TISSUE DAMAGE IN RATS**  
DEGREE : **MASTER OF SCIENCE**  
VIVA DATE : **27 MAY 2011**

## DECLARED BY

### 1. SUPERVISOR

ASSOC. PROF. DR. MOHAMMAD IQBAL

### 2. CO-SUPERVISOR

MR. RUZAIDI AZLI MODH. MOKHTAR



**UMS**  
UNIVERSITI MALAYSIA SABAH

## ACKNOWLEDGEMENT

I would like to deepest thank to my supervisor, Assoc. Prof. Dr. Mohammad Iqbal whom brought their professionalism and expertise in supervising, constant support and encouragement. His continuous support and encouragement provided me the strength and energy to complete the research and publish this thesis on time. Deepest thanks also to my co-supervisor, Mr. Ruzaidi Azli Mohd. Mokhtar for the guiding and supervision. He gave extensive comments on the thesis writing so that the thesis could be published successfully. I am also thankful to Prof. Datin Seri Panglima Dr. Ann Anton, Director of Biotechnology Research Institute for her support and encouragement

For the examination and analysis of histopathological study, I would like to express special thanks to Dr. Mie Mie Sein, School of Medicine, Universiti Malaysia Sabah for her professionalism and expertise on the analysis of histopathology slides. She has guided me along the interpretation of the histopathological examination. Furthermore, my special sense of gratitude to Prof. Dr. Amran Ahmed, School of Science and Technology, Universiti Malaysia Sabah for his valuable statistical analysis and interpretation guiding.

I would also like to thank Ministry of Higher Education, Malaysia for providing the grant-in-aid No. FRG 166-SP-2008 throughout the research project. In the mean time, special thanks to Ministry of Science, Technology and Innovation (MOSTI), Malaysia for offering me the University Postgraduate Research Scholarship Scheme. In addition, I would like to grateful thank Kebun Rimau Sdn. Bhd., Sabah for providing the fresh plant samples of *Cymbopogon citratus* and *Morinda citrifolia* throughout the research project.

For providing a stimulating research working environment, special thanks to lab assistants at Biotechnology Research Institute, Universiti Malaysia Sabah, Vidarita Maikin, Azian Awang Besar, Muhd. Adam Hairie Dailis Abdullah, Mony Mian and Emran Raga where the research began in earnest until the research project completed.

In the administration part, I would like to thank all the administrator and supportive staffs from Biotechnology Research Institute office, Universiti Malaysia Sabah for their grateful administration support. Special thanks to Junainah Ismail for helping in administration by completing the master project.

At home, Dr. Khoo Yong Pheng has all had a part to play in giving me to write the thesis, motivation and encouragement. Very thanks for his valuable comments.

## ABSTRACT

### CHEMOPREVENTIVE EFFECTS OF SELECTED HERBAL PLANTS AGAINST CARBON TETRACHLORIDE (CCl<sub>4</sub>)-MEDIATED OXIDATIVE TISSUE DAMAGE IN RATS

Oxidative damage of biomolecules is implicated in the pathogenesis of various chronic diseases including cancer. This has led to intensive investigation aimed at reducing the extent of such oxidative injury. The present study was aimed to evaluate the antioxidant and chemopreventive effects of selected herbal plants against carbon tetrachloride (CCl<sub>4</sub>)-mediated oxidative tissue damage in rats. Herbal plants viz *Aloe vera*, *Andrographis paniculata*, *Cymbopogon citratus* and *Morinda citrifolia* were selected and evaluated for their total phenolic and 2,2-diphenyl-2-picrylhydrazyl (DPPH) scavenging activity. Based on the results of *in vitro* studies, *A. paniculata* and *C. citratus* were selected further for *in vivo* studies due to their higher total phenolics and antioxidant activity. Rats were pretreated with ethanolic extract of *A. paniculata* and *C. citratus* accordingly to the selected doses (100 mg/kg b.w., 200 mg/kg b.w. and 300 mg/kg b.w.) for 14 days followed by two dosage of CCl<sub>4</sub> (1.2 ml/kg b.w.) via oral (gavage) on days 13 and 14. All of these animals were sacrificed 24 hours after the last dose of CCl<sub>4</sub> or saline. Blood, liver and kidney tissues were taken quickly for biochemical and histopathological studies to assess the derangement in the functioning of liver and kidneys. Challenge of CCl<sub>4</sub> induced oxidative stress both in the liver and kidneys, as evident from augmentation in lipid peroxidation (TBARS) which was accompanied by a decreased in antioxidant enzymes activities and depletion of glutathione reduced level. Parallel to these changes, CCl<sub>4</sub> enhanced hepatic damage as evidenced by a sharp increased in serum transaminases. However, blood urea nitrogen and serum creatinine were not elevated significantly. Additionally, serum biochemistry oxidative stress markers were consistent with the hepatic and renal histopathological studies. In liver, most of these changes were significantly alleviated by pretreatment of animals with 300 mg/kg b.w. *A. paniculata* and 200 mg/kg b.w. *C. citratus*. The ability of *A. paniculata* and *C. citratus* to scavenge the DPPH radical was determined and had an EC<sub>50</sub> value of 583.60 and 994.77 µg/ml respectively. In addition, the antioxidant activity was closely related to the total phenolic content as evident by *A. paniculata* and *C. citratus* showing the value of 65.37 and 30.74 mg GAE/g of extract. In contrast, the changes in the kidney were not significantly alleviated by the pretreatment with *A. paniculata* and *C. citratus* extracts. Present results indicate that the hepatoprotective effects of *A. paniculata* and *C. citratus* might be ascribable to its antioxidant and free radical scavenging properties. We concluded that *A. paniculata* and *C. citratus* could be used as hepatoprotective agents and possess the potential to be used to treat or prevent degenerative diseases where oxidative stress is implicated.

## ABSTRAK

Kerosakan oksidatif biomolekul menyebabkan pelbagai patogenesis penyakit kronik termasuk kanser. Ini membawa kepada penyelidikan intensif ditumpukan untuk mengurangkan kesebaran kerosakan oksidatif. Kajian ini adalah bertujuan untuk menilai antioksidan dan kesan pencegahan kemo dengan menggunakan tumbuhan herbal yang dipilih terhadap kerosakan tisu oksidatif akibat pendedahan carbon tetrachloride ( $CCl_4$ ) pada tikus. Aloe vera, *Andrographis paniculata*, *Cymbopogon citratus* dan *Morinda citrifolia* telah dipilih dan dinilai jumlah kandungan fenolik dan aktiviti penyingkiran radikal 2,2-diphenyl-2-picrylhydrazyl (DPPH). Berdasarkan keputusan *in vitro*, *A. paniculata* dan *C. citratus* mengandungi jumlah kandungan fenolik dan aktiviti antioksidan yang lebih tinggi dan dipilih seterusnya untuk kajian *in vivo*. Tikus diberikan rawatan ekstrak etanol *A. paniculata* dan *C. citratus* berdasarkan dos yang dipilih (100 mg/kg berat badan, 200 mg/kg berat badan dan 300 mg/kg berat badan) selama 14 hari dan diikuti dengan dua kali rawatan  $CCl_4$  (1.2 ml/kg berat badan) melalui mulut (gavage) pada hari ke-13 dan 14. Dua puluh empat jam selepas kali terakhir  $CCl_4$  atau saline, tikus dibunuh. Darah, hati dan ginjal dikeluarkan dengan segera untuk analisis biokimia dan histopatologi untuk menilai perubahan fungsi hati dan ginjal. Tikus yang diberi rawatan  $CCl_4$  mengalami tekanan oksidatif pada kedua-dua hati dan ginjal dengan menunjukkan perubahan dalam lipid peroxidation (TBARS) bersama penurunan aktiviti enzim antioksidan serta penurunan tahap glutathione. Selaras dengan perubahan itu, rawatan  $CCl_4$  meningkatkan kerosakan hati dengan menunjukkan peningkatan yang jelas pada serum transaminase. Namun demikian, nitrogen urea darah dan kreatinin tidak menunjukkan peningkatan yang signifikan. Tambahan pula, serum penunjuk tekanan oksidatif adalah selaras dengan histopatologi hati dan ginjal. Dalam hati, kebanyakan perubahan jelas dikurangkan oleh pemberian rawatan 300 mg/kg b.w. *A. paniculata* dan 200 mg/kg b.w. *C. citratus*. Keupayaan *A. paniculata* dan *C. citratus* untuk menyingkirkan radikal DPPH telah dinilai dan menunjukkan nilai  $EC_{50}$  583.60 dan 994.77  $\mu\text{g/ml}$  masing-masing. Selain itu, aktiviti antioksidan adalah berkaitan rapat dengan kandungan fenolik dengan *A. paniculata* dan *C. citratus* menunjukkan sebanyak 65.37 dan 30.74 mg GAE/g ekstrak masing-masing. Sebaliknya, tiada perbezaan signifikan pada ginjal apabila rawatan ekstrak *A. paniculata* dan *C. citratus* diberikan. Keputusan pada kajian ini menunjukkan bahawa kesan perlindungan hepatik *A. paniculata* dan *C. citratus* mungkin disebabkan oleh antioksidan dan keupayaan menyingkirkan radikal bebas. Kajian ini merumuskan bahawa *A. paniculata* dan *C. citratus* berupaya digunakan sebagai agen perlindungan hepatik dan berpotensi untuk mengubati atau mencegah penyakit berkaitan tekanan oksidatif.





<b>CHAPTER 2: LITERATURE REVIEW</b>	<b>5</b>
2.1 Carbon tetrachloride	5
2.1.1 Introduction	5
2.1.2 Mechanism Action	6
2.1.3 Carbon Tetrachloride Toxicity	10
2.2 <i>Aloe vera</i>	13
2.2.1 Introduction	13
2.2.2 Traditional Uses	14
2.2.3 Chemical Constituents	15
2.2.4 Biological Effects	16
2.3 <i>Andrographis paniculata</i>	18
2.3.1 Introduction	18
2.3.2 Traditional Uses	20
2.3.3 Chemical Constituents	21
2.3.4 Biological Effects	21
2.4 <i>Cymbopogon citratus</i>	25
2.4.1 Introduction	25
2.4.2 Traditional Uses	26
2.4.3 Chemical Constituents	26
2.4.4 Biological Effects	27
2.5 <i>Morinda citrifolia</i>	30
2.5.1 Introduction	30
2.5.2 Traditional Uses	32
2.5.3 Chemical Constituents	33
2.5.4 Biological Effects	35
<b>CHAPTER 3: METHODOLOGY</b>	<b>38</b>
3.1 Plant Materials	38
3.2 Chemicals	42
3.3 Preparation of Ethanolic Extract	43
3.4 Determination of Total Phenolic Content	43
3.5 Determination of 2,2-Diphenyl-2-picrylhydrazyl (DPPH) Assay	44
3.6 Animals	45
3.6.1 Treatment Conditions	45
3.6.2 Collection of Serum and Tissue	46
3.7 Determination of Final Body Weight	47
3.8 Assessment of Liver and Kidney Index	47
3.9 Serum Biochemistry	48
3.9.1 Determination of Serum Alanine	48

	Aminotransferase (ALT)	
3.9.2	Determination of Serum Aspartate Aminotransferase (AST)	48
3.9.3	Determination of Serum Urea Nitrogen	49
3.9.4	Determination of Serum Creatinine	50
3.9.5	Determination of Serum Lactate Dehydrogenase (LDH) Activity	51
3.10	Preparation of Tissues Post Mitochondrion Supernatant (PMS)	51
3.11	Determination of Total Protein Contents	52
3.12	Biochemical Analysis	53
3.12.1	Determination of Reduced Glutathione (GSH)	53
3.12.2	Determination of Lipid Peroxidation (LPO)	54
3.12.3	Determination of Catalase (CAT) Activity	55
3.12.4	Determination of Glutathione Peroxidase (GPx) Activity	56
3.12.5	Determination of Glutathione Reductase (GR) Activity	57
3.12.6	Determination of Quinone Reductase (QR) Activity	58
3.12.7	Determination of Glutathione-S-transferase (GST) Activity	59
3.12.8	Determination of Glucose-6-phosphate Dehydrogenase (G6PD) Activity	60
3.12.9	Determination of $\gamma$ -Glutamyl Transpeptidase ( $\gamma$ -GT) activity	61
3.13	Histopathological Examination	62
3.14	Statistical Analysis	65
	<b>CHAPTER 4: RESULTS</b>	66
4.1	Extraction Yield and Total Phenolic Content	66
4.2	2,2-Diphenyl-2-picrylhydrazyl (DPPH) Assay	67
4.3	Effects of <i>A. paniculata</i> Extract on Carbon Tetrachloride (CCl <sub>4</sub> )-Mediated Oxidative Tissue Damage in Rats	68
4.3.1	Body Weight	68
4.3.2	Liver and Kidney Index	69
4.3.3	Alanine Aminotransferase (ALT), Aspartate Aminotransferase (AST) and Lactate Dehydrogenase (LDH)	70

4.3.4	Blood Urea Nitrogen and Creatinine	71
4.3.5	Lipid Peroxidation (LPO)	72
4.3.6	Reduced Glutathione (GSH)	73
4.3.7	Quinone Reductase (QR)	74
4.3.8	Glutathione Peroxidase (GPx), Glutathione Reductase (GR) and Glutathione-S-transferase (GST)	75
4.3.9	Catalase (CAT), Glucose-6-phosphate Dehydrogenase (G6PD) and $\gamma$ -Glutamyl Transpeptidase ( $\gamma$ -GT)	78
4.3.10	Histopathological Examination	81
4.4	Effects of <i>C. citratus</i> Extract on Carbon Tetrachloride (CCl <sub>4</sub> )-Mediated Oxidative Tissue Damage in Rats	88
4.4.1	Body Weight	88
4.4.2	Liver and Kidney Index	89
4.4.3	Alanine Aminotransferase (ALT), Aspartate Aminotransferase (AST) and Lactate Dehydrogenase (LDH)	90
4.4.4	Blood Urea Nitrogen and Creatinine	91
4.4.5	Lipid Peroxidation (LPO)	92
4.4.6	Reduced Glutathione (GSH)	93
4.4.7	Quinone Reductase (QR)	94
4.4.8	Glutathione Peroxidase (GPx), Glutathione Reductase (GR) and Glutathione-S-transferase (GST)	95
4.4.9	Catalase (CAT), Glucose-6-phosphate Dehydrogenase (G6PD) and $\gamma$ -Glutamyl Transpeptidase ( $\gamma$ -GT)	98
4.4.10	Histopathological Examination	101
 <b>C H A P T E R 5 : D I S C U S S I O N</b>		
1	0	8
5.1	Extraction Yield and Total Phenolic Content	108
5.2	Free Radical Scavenging Activity	109
5.3	Effects of <i>A. paniculata</i> and <i>C. citratus</i> Extracts on Body Weight, Liver Index and Kidney Index in Rats	110
5.4	Effects of <i>A. paniculata</i> and <i>C. citratus</i> Extracts against CCl <sub>4</sub> -Mediated Oxidative Hepatic and Renal Damage in Rats	111

<b>CHAPTER</b>	<b>6:</b>	<b>CONCLUSION</b>
124		
<b>R</b>	<b>E</b>	<b>F</b>
<b>E</b>	<b>R</b>	<b>E</b>
<b>N</b>	<b>C</b>	<b>E</b>
<b>S</b>		
1	2	7
<b>APPENDIX</b>		
1	5	3

### LIST OF TABLES

	Page
Table 3.1 Parts, stages of maturity and sizes of plants used	39
Table 3.2 Procedure steps for tissue processing	63
Table 3.3 Procedure steps for auto-staining	64
Table 4.1 Extraction yields and total phenolic contents of <i>A. vera</i> , <i>A. paniculata</i> , <i>C. citratus</i> and <i>M. citrifolia</i> plant extracts	66
Table 4.2 DPPH free radical scavenging activity of <i>A. vera</i> , <i>A. paniculata</i> , <i>C. citratus</i> and <i>M. citrifolia</i> plant extracts compared with ascorbic acid	67
Table 4.3 Effects of <i>A. paniculata</i> extract on initial and final body weight	68
Table 4.4 Effects of <i>A. paniculata</i> extract on liver and kidney index	69
Table 4.5 Effects of <i>A. paniculata</i> extract on serum ALT, AST and LDH	70
Table 4.6 Effects of <i>A. paniculata</i> extract on blood urea nitrogen and creatinine	71

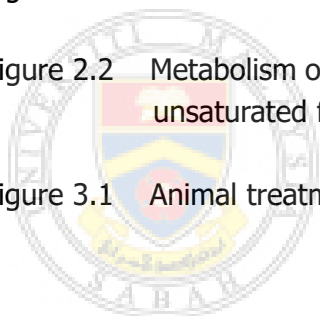
Table 4.7	Effects of <i>A. paniculata</i> extract on hepatic and renal LPO	72
Table 4.8	Effects of <i>A. paniculata</i> extract on hepatic and renal GSH	73
Table 4.9	Effects of <i>A. paniculata</i> extract on hepatic and renal QR	74
Table 4.10	Effects of <i>A. paniculata</i> extract on hepatic GPx, GR and GST	76
Table 4.11	Effects of <i>A. paniculata</i> extract on renal GPx, GR and GST	77
Table 4.12	Effects of <i>A. paniculata</i> extract on hepatic CAT, G6PD and $\gamma$ -GT	79
Table 4.13	Effects of <i>A. paniculata</i> extract on renal CAT, G6PD and $\gamma$ -GT	80
Table 4.14	Effects of <i>C. citratus</i> extract on initial and final body weight	88
Table 4.15	Effects of <i>C. citratus</i> extract on liver and kidney index	89
Table 4.16	Effects of <i>C. citratus</i> extract on serum ALT, AST and LDH	90
Table 4.17	Effects of <i>C. citratus</i> extract on blood urea nitrogen and creatinine	91
Table 4.18	Effects of <i>C. citratus</i> extract on hepatic and renal LPO	92
Table 4.19	Effects of <i>C. citratus</i> extract on hepatic and renal GSH	93
Table 4.20	Effects of <i>C. citratus</i> extract on hepatic and	94

renal QR

Table 4.21	Effects of <i>C. citratus</i> extract on hepatic GPx, GR and GST	96
Table 4.22	Effects of <i>C. citratus</i> extract on renal GPx, GR and GST	97
Table 4.23	Effects of <i>C. citratus</i> extract on hepatic CAT, G6PD and $\gamma$ -GT	99
Table 4.24	Effects of <i>C. citratus</i> extract on renal CAT, G6PD and $\gamma$ -GT	100

### LIST OF FIGURES

	Page	
Figure 2.1	Chemical structure of carbon tetrachloride	5
Figure 2.2	Metabolism of carbon tetrachloride (RH represents unsaturated fatty acids)	8
Figure 3.1	Animal treatment	45



UMMS  
UNIVERSITI MALAYSIA SABAH

## LIST OF PHOTOS

	Page
Photo 2.1 <i>Aloe vera</i>	13
Photo 2.2 <i>Andrographis paniculata</i>	19
Photo 2.3 <i>Cymbopogon citratus</i>	25
Photo 2.4 <i>Morinda citrifolia</i>	31
Photo 3.1 <i>Aloe vera</i>	40
Photo 3.2 <i>Andrographis paniculata</i>	40
Photo 3.3 <i>Cymbopogon citratus</i>	41
Photo 3.4 <i>Morinda citrifolia</i>	41
Photo 4.1 Histopathological sections of liver. (A) Control; (B) CCl <sub>4</sub> ; (C) <i>A. paniculata</i> 100 mg/kg b.w. and CCl <sub>4</sub> ; (D) <i>A. paniculata</i> 200 mg/kg b.w. and CCl <sub>4</sub> ; (E) <i>A. paniculata</i> 300 mg/kg b.w. and CCl <sub>4</sub> . Sections stained with haematoxylin and eosin. Black arrow, fatty changes. Red arrow, infiltration of inflammatory cells. Magnification 100X.	82



- Photo 4.2 Histopathological sections of kidney. 85  
(A) Control; (B) CCl<sub>4</sub>; (C) *A. paniculata* 100 mg/kg b.w. and CCl<sub>4</sub>; (D) *A. paniculata* 200 mg/kg b.w. and CCl<sub>4</sub>; (E) *A. paniculata* 300 mg/kg b.w. and CCl<sub>4</sub>.  
Sections stained with haematoxylin and eosin.  
Black arrow, necrosis. Blue arrow, glomerular atrophy.  
Magnification 100X.
- Photo 4.3 Histopathological sections of liver. 102  
(A) Control; (B) CCl<sub>4</sub>; (C) *C. citratus* 100 mg/kg b.w. and CCl<sub>4</sub>; (D) *C. citratus* 200 mg/kg b.w. and CCl<sub>4</sub>; (E) *C. citratus* 300 mg/kg b.w. and CCl<sub>4</sub>.  
Sections stained with haematoxylin and eosin.  
Black arrow, fatty changes.  
Red arrow, infiltration of inflammatory cells.  
Magnification 100X.
- Photo 4.4 Histopathological sections of kidney. 105  
(A) Control; (B) CCl<sub>4</sub>; (C) *C. citratus* 100 mg/kg b.w. and CCl<sub>4</sub>; (D) *C. citratus* 200 mg/kg b.w. and CCl<sub>4</sub>; (E) *C. citratus* 300 mg/kg b.w. and CCl<sub>4</sub>.  
Black arrow, necrosis. Blue arrow, glomerular atrophy.  
Sections stained with haematoxylin and eosin.  
Magnification 100X.

## LIST OF ABBREVIATIONS

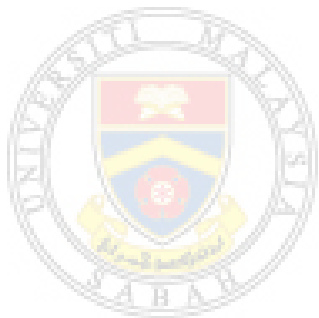
<b>Abs</b>	Absorbance
<b>Δ Abs</b>	Changes of absorbance
<b>ABTS</b>	2,2-Azinobis (3-ethyl benzothiazoline-6-sulphonic acid)
<b>AIDS</b>	Acquired immune deficiency syndrome
<b>ALP</b>	Alkaline phosphatase
<b>ALT</b>	Alanine aminotransferase
<b>ANOVA</b>	One-way analysis of variance
<b><i>A. paniculata</i></b>	<i>Andrographis paniculata</i>
<b>AST</b>	Aspartate aminotransferase
<b><i>A. vera</i></b>	<i>Aloe vera</i>
<b>BCA</b>	Bicinchoninic acid
<b>BCA1</b>	Bicinchoninic acid protein assay kit
<b>BHT</b>	Butylated hydroxyl toluene
<b>BSA</b>	Bovine serum albumin
<b>CAPE</b>	Caffeic acid phenethyl ester
<b>CAT</b>	Catalase
<b>CB2</b>	Cannabinoid 2
<b>•CCl<sub>3</sub></b>	Trichloromethyl radical
<b>CCl<sub>4</sub></b>	Carbon tetrachloride
<b><i>C. citratus</i></b>	<i>Cymbopogon citratus</i>
<b>CCl<sub>3</sub>OO•</b>	Trichloromethyl peroxy radical
<b>CD<sup>8+</sup></b>	Cluster of differentiation 8
<b>CDNB</b>	1-Chloro-2,4-dinitrobenzene
<b>cm</b>	Centimeter
<b>cm<sup>-1</sup></b>	Per centimeter
<b>CO<sub>2</sub></b>	Carbon dioxide

<b>Cu<sup>+</sup></b>	Copper (I) ion
<b>Cu<sup>2+</sup></b>	Copper (II) ion
<b>CYP2B1</b>	Cytochrome P450 2B1
<b>CYP2B2</b>	Cytochrome P450 2B2
<b>CYP2E1</b>	Cytochrome P450 2E1
<i>cyp2e1<sup>-/-</sup></i>	CYP2E1 knockout
<i>cyp2e1<sup>+/+</sup></i>	CYP2E1 wild-type
<b>CYP3A</b>	Cytochrome P450 3A
<b>DIAP</b>	14-Deoxy-11,12-didehydroandrographolide
<b>DCIP</b>	Dichlorophenolindophenol
<b>df</b>	Dilution factor
<b>DMSO</b>	Dimethyl sulfoxide
<b>DPPH</b>	1,1-Diphenyl-2-picryl-hydrazyl or 2,2-diphenyl-2-picrylhydrazyl
<b>DPX</b>	Di-N-butyle phthalate in xylene
<b>DNA</b>	Deoxyribonucleic acid
<b>DTNB</b>	5,5'-Dithio-bis-2-nitrobenzoic acid
<b>EC<sub>50</sub></b>	Concentration giving 50% inhibition
<b>EDTA</b>	Ethylenediamine tetra acetic acid
<b>FAD</b>	Flavin adenine dinucleotide
<b>FDA</b>	Food and Drug Administration
<b>g</b>	Gram
<b>G6PD</b>	Glucose-6 phosphate dehydrogenase
<b>GAE</b>	Gallic acids equivalents
<b>g/cm<sup>3</sup></b>	Gram per cubic centimeter
<b>GC-MS</b>	Gas chromatography-mass spectrometry
<b>g/l</b>	Gram per liter
<b>g/mol</b>	Gram per mole
<b>GPx</b>	Glutathione peroxidase
<b>GR</b>	Glutathione reductase
<b>G-SDNB</b>	1-Chloro-2,4-dinitrobenzene conjugate
<b>GSH</b>	Reduced glutathione
<b>GSSG</b>	Glutathione oxidized
<b>GST</b>	Glutathione-S-transferase
<b>GS-TNB</b>	Mixed disulphide (between reduced glutathione and 5-thionitrobenzoic acid)
<b>γ-GT</b>	γ-Glutamyl transpeptidase
<b>h</b>	Hour
<b>H<sup>+</sup></b>	Hydrogen ion
<b>HCl</b>	Hydrogen chloride
<b>HDL</b>	High-density lipoprotein
<b>HPLC</b>	High-performance liquid chromatography

<b>H&amp;E</b>	Haematoxylin and eosin
<b>H<sub>2</sub>O</b>	Water
<b>H<sub>2</sub>O<sub>2</sub></b>	Hydrogen peroxide
<b>IFN-γ</b>	Interferon-gamma
<b>IgA</b>	Immunoglobulin A
<b>IgG</b>	Immunoglobulin G
<b>IgM</b>	Immunoglobulin M
<b>IL-1β</b>	Interleukin-1β
<b>IL-2</b>	Interleukin-2
<b>IL -4</b>	Interleukin-4
<b>IL-6</b>	Interleukin-6
<b>IU/L</b>	International units per liter
<b>KCl</b>	Potassium chloride
<b>LC-MS</b>	Liquid chromatography-mass spectrometry
<b>LDH</b>	Lactate dehydrogenase
<b>LDL</b>	Low density lipoprotein
<b>LPO</b>	Lipid peroxidation
<b>m</b>	Meter
<b>M</b>	Molarity
<b>M<sup>-1</sup></b>	Per Molarity
<b>Mac-1</b>	Macrophage-1
<b><i>M. citrifolia</i></b>	<i>Morinda citrifolia</i>
<b>MDA</b>	Malondialdehyde
<b>mg</b>	Miligram
<b>Mg<sup>2+</sup></b>	Magnesium ion
<b>MgCl<sub>2</sub></b>	Magnesium chloride
<b>mg/dl</b>	Miligram per deciliter
<b>mg/kg b.w.</b>	Miligram per kilogram body weight
<b>mg/ml</b>	Miligram per milliliter
<b>min</b>	Minute
<b>ml</b>	Mililiter
<b>ml/kg b.w.</b>	Mililiter per kilogram body weight
<b>mm</b>	Milimeter
<b>mM</b>	Milimolarity
<b>mol/l</b>	Mole/liter
<b>N</b>	Normality
<b>n</b>	Number
<b>Na<sub>2</sub>CO<sub>3</sub></b>	Sodium carbonate
<b>NAD<sup>+</sup></b>	Nicotinamide adenine dinucleotide
<b>NADH</b>	Nicotinamide adenine dinucleotide reduced
<b>NADP</b>	Nicotinamide adenine dinucleotide phosphate

<b>NADP<sup>+</sup></b>	Nicotinamide adenine dinucleotide phosphate
<b>NADPH</b>	Nicotinamide adenine dinucleotide phosphate reduced
<b>NaOH</b>	Sodium hydroxide
<b>NF-kappa B</b>	Nuclear factor kappa-light-chain-enhancer of activated B cell
<b>NH<sub>3</sub></b>	Ammonia
<b>NH<sub>4</sub><sup>+</sup></b>	Ammonium ion
<b>(NH<sub>2</sub>)<sub>2</sub>CO</b>	Ammonium carbonate
<b>NK</b>	Natural killer
<b>nm</b>	Nanometer
<b>nmol</b>	Nanomole
<b>No.</b>	Number
<b>O<sub>2</sub></b>	Oxygen
<b>O<sub>2</sub>•<sup>-</sup></b>	Superoxide radical
<b>•OH</b>	Hydroxyl radical
<b>ONOO<sup>-</sup></b>	Peroxynitrite radical
<b>ORAC</b>	Oxygen radical absorbance capacity
<b>p</b>	Probability
<b>pH</b>	Negative decimal logarithm of the hydrogen ion activity in a solution
<b>PMS</b>	Post-mitochondrion supernatant
<b>PO<sub>4</sub><sup>3-</sup></b>	Phosphate ion
<b>POD</b>	Peroxidase
<b>PyOD</b>	Pyruvate oxidase
<b>QR</b>	Quinone reductase
<b>R•</b>	Alkoxy radical
<b>RH</b>	Unsaturated fatty acids
<b>ROO•</b>	Peroxy radical
<b>ROS</b>	Reactive oxygen species
<b>rpm</b>	Revolutions per minute
<b>RSA</b>	Radical scavenging activity
<b>s</b>	Second
<b>SD</b>	Standard deviation
<b>SnCl<sub>2</sub></b>	Stannous chloride
<b>SO<sub>3</sub><sup>2-</sup></b>	Sulphate ion
<b>SOD</b>	Superoxide dismutase
<b>SPSS</b>	Statistical package for the social sciences
<b>TBA</b>	Thiobarbituric acid
<b>TBARS</b>	Thiobarbituric acid reactive substance
<b>TCA</b>	Trichloroacetic acid
<b>TNB</b>	5-Thionitrobenzoic acid

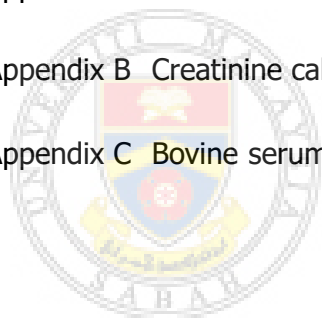
<b>TNF-<math>\alpha</math></b>	Tumor necrosis factor-alpha
<b>TRAP</b>	Total radical trapping antioxidant parameter
<b>tris-HCl</b>	Tris(hydroxymethyl)aminomethane hydrochloric acid buffer
<b>UMS</b>	Universiti Malaysia Sabah
<b>USA</b>	United State of America
<b>VLDL</b>	Very low-density lipoprotein
<b>v/v</b>	Volume per volume
<b>WHO</b>	World Health Organization
<b>w/v</b>	Weight per volume
<b><math>\mu\text{g/ml}</math></b>	Microgram per milliliter
<b><math>\mu\text{l}</math></b>	Microliter
<b><math>\mu\text{m}</math></b>	Micrometer
<b><math>\mu\text{mol}</math></b>	Micromole
<b><math>^{\circ}\text{C}</math></b>	Degree Celsius
<b>%</b>	Percent



**UMS**  
UNIVERSITI MALAYSIA SABAH

## LIST OF APPENDIX

	Page
Appendix A Gallic acid calibration curve	153
Appendix B Creatinine calibration curve	154
Appendix C Bovine serum albumin calibration curve	155



UMMS  
UNIVERSITI MALAYSIA SABAH

## CHAPTER 1

### INTRODUCTION

#### 1.1 Background of the Study

Reactive oxygen species (ROS) such as  $O_2\bullet^-$ ,  $H_2O_2$  and  $\bullet OH$ , are continuously generated inside the human body. They are generated due to the exposure of exogenous chemicals in our ambient environment and/or a number of endogenous metabolic processes involving redox enzymes and bioenergetics electron transfer. However, the ROS generated are being detoxified by the antioxidant in the body under normal circumstances. There is homeostasis equilibrium between the ROS generated and the antioxidants present. Imbalance equilibrium between the formation and inactivation of these ROS causes the detrimental effects. However, this equilibrium is affected and favoring the increase of ROS formation that culminates in oxidative stress. This phenomenon is due to ROS overproduction and/or inadequate antioxidant defense. The ROS attack to various biomolecules including proteins, lipids, mitochondria, lipoproteins and DNA which ultimately induce oxidative damage (Farber, 1994). This oxidative damage is an important etiological factor implicated in several chronic human diseases such as diabetes mellitus, cancer, atherosclerosis, arthritis, neurodegenerative diseases and also in ageing process (Hogg, 1998; Pong, 2003).

Liver and kidney disorders are the serious health problem in the worldwide. The Global Burden of Disease: 2004 Update (2008) reported that liver cirrhosis, nephritis and nephrosis were categorized in the 20 leading causes of deaths in 2004 for all WHO member state. In Peninsular Malaysia, liver cancer was the sixth most common cancers among population in 2006. There were 793 cases per