#### UNIVERSITI MALAYSIA SABAH

BORANG PE	NGESAHAN STATUS TESIS@
JUDUL: INHIBITING TWO-CO	OMPONENT SIGNAL. TRANSPUCTION
SYSTEM IN MYC	OBACTERIUM TUBERCULOSIS
Ijazah: Sains Sarjana Muda	Sains Dengan Kepujian
SESI PEN	GAJIAN: 2002 - 2005
Saya JENIFER ROLLAND	
	(HURUF BESAR) Doktor Falsafab)* ini disimpan di Perpustakaan Universiti aan seperti berikut:
	sia Sabah. a dibenarkan membuat salinan untuk tujuan pengajian sahaja. aan tesis ini sebagai bahan pertukaran antara institusi pengajian (Mengandungi maklumat yang berdarjah keselamatan atau kepentingan Malaysia seperti yang termaktub di dalam
TERHAD	AKTA RAHSIA RASMI 1972) (Mengandungi maklumat TERHAD yang telah ditentukan oleh organisasi/badan di mana penyelidikan dijalankan)
LO.D.	Disahkan oleh
(TANDATANGAN PENULIS)	(TANDATANGAN PUSTAKAWAN)
Alamat Tetap: P/s 244, Kg. Tondulu; 89657 Tambunan, Sabah.	Prof. Ho Coy Choke Nama Penyelia
Tarikh: 31 Mac 2005	Tarikh: 31 Mac 2005

CATATAN: \* Potong yang tidak berkenaan.

.

. .

\*\* Jika tesis ini SULIT atau TERHAD, sila lampirkan surat daripada pihak berkuasa/organisasi berkenaan dengan menyatakan sekali sebab dan tempoh tesis ini perlu dikelaskan sebagai SULIT dan TERHAD.

@ Tesis dimaksudkan sebagai tesis bagi Ijazah Doktor Falsafah dan Sarjana secara penyelidikan, ztau disertasi bagi pengajian secara kerja kursus dan penyelidikan, atau Laporan Projek Sarjana Muda (LPSM).



230006



# INHIBITING TWO-COMPONENT SIGNAL TRANSDUCTION SYSTEM IN MYCOBACTERIUM TUBERCULOSIS

JENIFER ROLLAND L

# THIS DISSERTATION IS SUBMITTED TO FULFILL PARTIAL OF THE REQUIREMENT TO OBTAIN A DEGREE IN BACHELOR OF SCIENCE WITH HONOUR

# BIOTECHNOLOGY PROGRAMME SCHOOL OF SCIENCE AND TECHNOLOGY UNIVERSITY OF MALAYSIA SABAH

PERPUSTAKAAN UNIVERSITI MALAYSIA SABAH

2005







### DECLARATION

I hereby certify that this masterpiece of mine is my very own product except for the statements, abridgements and summaries that each of them has been explained its source.

March 29, 2005

er follal

JENIFER ROLLAND L HS2002-3047



Signature

1. SUPERVISOR

(Prof. Ho Coy Choke)

2. EXAMINER 1

(Dr. Jualang @ Azlan bin Gansau)

3. EXAMINER 2

(Dr. Zaleha Abdul Aziz)

4. DEAN

(Prof. Madya Dr. Amran Ahmed)

UNIVERS

Huleychok

### ACKNOWLEDGEMENT

Although I have put a lot of effort on succeeding this project, its outcome book of report would have never come together in this way without the contributions of many individuals. First and foremost, a thousand thanks to my project's supervisor, Prof. Dr. Ho Coy Choke, for all the taught, encouragement and supports that have brought me to this level. To my academic supervisor, Dr. Lee Ping Chin, thanks for all the advices along my journey of learning in University of Malaysia Sabah. Not forgetting all my other lecturers, Prof. Perumal Ramasamy, Prof. Paranjothy Karthigesu, Prof. Datuk Dr. Kamaruzaman Ampon, Dr. Jualang @ Azlan Abdullah bin Gansau, Dr. Zaleha Abdul Aziz, Dr. Roziah Kambul, Ms. Teoh Peik Lin, Dr. Vijay Kumar and Dr. Michael Wong, for sharing their knowledge to the students as well as giving care and supports. I also wish to acknowledge the post-graduates students in Pascasiswazah Laboratory of UMS -Mr. Ho Wei Loon offered inputs to my project, including several very useful reviews besides assisting me along the progress of my project. Others, Mr. Ong Si Mon for his brilliant tips and cheering humors, Ms. Puah Seok Hwa and Ms. Hew Chaw Sen for keeping the project moving in dealing with my requests, and last but not least, Mr. Foo Sek Hin for his encouraging taught and comments. This acknowledgement also goes to Sabah Foundation Group (Kumpulan Yayasan Sabah) for letting us to do research and collecting soil samples at Danum Valley Field Centre in Lahad Datu. Thanks a lot to these individuals, Mr. Bernadus Bala Ola (Senior Forest Ranger) and the other forest rangers, Mr. Herman Francis Tating, Mr. Nasir Abdul Majid and Mr. Johar Aribin for their great help in collecting soil samples and species determination. Thanks also for the subsequent species determination done by Forestry Research Center (FRC) in Sandakan. I also wish to acknowledge all the undergraduate students under my project's supervisor for their great contributions and very much helpful in progressing my project. My final acknowledgement goes to my friends and family, though I cannot state everyone's name, your supportive help and comments mean a lot to me. Again, thank you very, very much.



### ABSTRACT

In this work, my aim is to identify potential antimicrobial compounds that inhibit the regulation of two-component system in Mycobacterium tuberculosis over osmotic stress. Few diversity of actinomycetes were isolated from a total of 16 soil samples that had been collected from the virgin tropical rainforest of Danum Valley in Sabah. The soil samples were mainly collected under selected trees in the Newbery Plot area. A total of 46 strains of actinomycetes had been isolated from these soil samples through isolation using HV media and purification using Oatmeal media. Through fermentation process, different secondary metabolites were extracted from all these actinomycetes strains. The resulting acetone extracts were screened for K<sup>+</sup> and Na<sup>+</sup> signals in Mycobacterium twocomponent signal transduction system, on media that had been developed to apply osmotic pressure. Mycobacterium smegmatis was grown using the modified M9 minimal media with low and high concentration of potassium (K<sup>+</sup>) and sodium (Na<sup>+</sup>) ions. Wildtype Mycobacterium smegmatis mc<sup>2</sup>155 strain H8000 was used as targeted microorganism in modified M9 minimal media. The pH of media was adjusted to 6.7. Glucose, thiamine hydrochloride (Vitamin B<sub>1</sub>) and the inoculum of M. smegmatis were added into the media. The culture was incubated in 37°C for 3-4 days. Streptomycin was used as a negative control. A new simple but interesting discovery was found regarding streptomycin where its resulting inhibition zones were decreasing as the concentration of Na<sup>+</sup> signal in media arise. Besides reducing the growth of M. smegmatis, higher concentration of signals in the media had increased the media density, which might explain why the fixed concentration of streptomycin used as control cannot widen its inhibition zone. Only Extract H11588 showed antimicrobacterial activity on the twocomponent signal transduction system in Mycobacterium. However, it was later defined as toxic to M. smegmatis because it showed inhibition zones on both low and high concentrations (Na<sup>+</sup> concentration: Low: 0.8% (w/v); High: 4.0% (w/v); K<sup>+</sup> concentration: Low 100 µM; High: 1 mM) of both studied signals.



### ABSTRAK

Objektif kajian ini adalah untuk mengenalpasti sebatian antimikrob yang berpotensi merencat pengawalaturan sistem dua-komponen di dalam Mycobacterium tuberculosis terhadap tekanan osmotik. Pelbagai aktinomiset telah diasingkan daripada sejumlah 16 sampel tanah yang diperolehi daripada hutan hujan tropika di Lembah Danum, Sabah. Sampel-sampel tanah diambil pada bahagian bawah pokok-pokok yang terpilih di dalam kawasan Plot Newbery. Sejumlah 46 strain aktinomiset telah berjaya dipencilkan daripada sampel-sampel tanah ini melalui kaedah pengasingan menggunakan media selektif HV dan kaedah penulenan menggunakan media kompleks Oatmeal. Melalui proses penapaian, pelbagai metabolit sekunder diekstrak daripada keseluruhan strain-strain aktinomiset ini. Ekstrak-ekstrak aseton yang terhasil diuji untuk isyarat K<sup>+</sup> dan Na<sup>+</sup> terhadap sistem isyarat transduksi dua-komponen Mycobacterium ke atas media yang telah diubahsuai untuk tekanan osmotik. Mycobacterium smegmatis dikulturkan pada media minimum M9 yang telah diubahsuai dengan kepekatan ion kalium (K<sup>+</sup>) dan natrium (Na<sup>+</sup>) yang tinggi dan rendah. Mycobacterium smegmatis mc<sup>2</sup>155 jenis liar strain H8000 digunakan sebagai mikroorganisma sasaran. pH media diselaraskan kepada 6.7. Glukosa, thiamine hydrochloride (Vitamin B<sub>1</sub>) dan inokulum M. smegmatis ditambahkan ke dalam media. Kultur kemudian dieramkan pada suhu 37°C selama 3-4 hari. Streptomycin digunakan sebagai kawalan negatif. Satu penemuan ringkas tetapi memberangsangkan mengenai streptomycin adalah di mana ia menghasilkan zon perencatan yang semakin kecil apabila kepekatan Na<sup>+</sup> semakin meningkat di dalam media. Kepekatan ion isyarat yang semakin tinggi dalam media telah meningkatkan kepadatan media, yang mana menjelaskan pengurangan zon perencatan yang dihasilkan oleh streptomycin. Peningkatan kepekatan K<sup>+</sup> dan Na<sup>+</sup> juga merencat pertumbuhan M. smegmatis, Hanya Ekstrak H11588 menunjukkan aktiviti antimikrobacteria terhadap sistem isyarat transduksi dua-komponen dalam Mycobacterium. Walau bagaimanapun, ia adalah toksik kepada M. smegmatis kerana menghasilkan zon perencatan pada kepekatan tinggi dan rendah (Kepekatan Na<sup>+</sup>: Rendah: 0.8% (w/v); Tinggi: 4.0% (w/v); Kepekatan K<sup>+</sup>: Rendah: 100 µM; Tinggi: 1 mM) untuk isyarat K<sup>+</sup> dan Na<sup>+</sup>.



## **TABLE OF CONTENTS**

1 ugo	P	a	g	e	
-------	---	---	---	---	--

DEC	LARATION	ii
VER	IFICATION	iii
ACK	NOWLEDGEMENT	iv
ABS	TRACT	v
ABS	TRAK	vi
TAB	LE OF CONTENTS	vii
TAB	LES	xi
FIGU	JRES	xii
PHO	TOS	xiii
SYM	IBOLS, UNITS, ABBREVIATIONS, TERMS & FORMULAS	xiv
СНА	APTER 1 INTRODUCTION	1
СНА	APTER 2 LITERATURE REVIEW	4
2.1	Actinomycetes	4
	2.1.1 Nocardia	6
	2.1.2 Actinomyces	7
	2.1.3 Streptomyces	8
	2.1.4 Microbial secondary metabolites from actinomycetes	8
2.2	Two-Component System	11
	2.2.1 Two-component system of EnvZ-OmpR in E. coli	12
2.3	Research Location	17
2.4	Research Problems	20



CHAP	TER 3	3 MATERIALS AND METHODS	21
3.1	Sterili	zation Techniques	21
	3.1.1	Autoclave	22
	3.1.2	Heat Sterilization	22
	3.1.3	Sieving	23
3.2	Collec	cting Soil Samples	23
3.3	pH Determination		27
3.4	Isolati	on of Actinomycetes	27
5.1	3.4.1	Preparation of humic acid – Vitamin B	28
	3.4.2	Serial dilution	30
3.5	Purific	cation of Actinomycetes	31
	3.5.1	Preparation of Oatmeal media	32
	3.5.2	Purification of single colony of actinomycete	33
3.6	Prepa	ration of 20% Glycerol Stock Solution	34
3.7	Gram Staining		34
	3.7.1	Preparation of reagents	34
	3.7.2	Preparation of smear	36
	2.8.3	Gram Staining and preparation of slide	36
3.8	Prepar	ration of Extracts from Actinomycetes	37
	3.8.1	Preparation of fermentation media	37
	3.8.2	Fermentation	38
	3.8.3	Extraction	38
3.9	Screen	ning	38
	3.9.1	Hypothesis / Expected result	39



viii

	3.9.2 Preparation of seed culture of Mycobacterium smegmatis	41
	3.9.3 Preparation of paper discs	44
	3.9.4 Preparation of screening media	44
	3.9.5 Placement of paper discs onto screening media and incubation	45
	3.9.6 Scoring	46
СНА	PTER 4 RESULT AND DATA ANALYSIS	47
4.1	pH Determination	47
4.2	Origins of Soil Samples	48
4.3	Isolation and Purification	50
4.4	Glycerol Stock and Slant Agars	54
4.5	Extracts of Actinomycetes	55
4.6	Screening Result	56
	4.6.1 Seed culture	56
	4.6.2 Developing screening media	56
	4.6.3 Screening result for $K^+$ and $Na^+$ signals	61
СНА	PTER 5 DISCUSSION	64
5.1	Sterilization Techniques	64
5.2	Collecting Soil Samples	65
5.3	pH Determination	67
5.4	Isolation of Actinomycetes	67
5.5	Purification of Actinomycetes	68
5.6	Glycerol Stock Solution	69
5.7	Gram Staining	69
5.8	Extracts of Actinomycetes	70
5.9	Screening	71



CHAPTER 6	CONCLUSION	74
REFERENCES		76
APPENDIX A		78
APPENDIX B		80
APPENDIX C		81
APPENDIX D		82
APPENDIX E		84
APPENDIX F		85



x

## TABLES

## Pages

3.1	Expected results for screening method.	39
3.2	Chemicals involved in studying different concentration of Na <sup>+</sup> .	42
3.3	Chemicals involved in studying different concentration of K <sup>+</sup> .	43
4.1	Characteristics of isolates that have been isolated from Danum Valley.	52
4.2	The initial screening result on different concentration of Na <sup>+</sup> .	57
4.3	Recorded data of the following graph of Figure 4.1.	57
4.4	Screening result on different concentration of K <sup>+</sup> .	58
4.5	Screening result on extract from H11588 strain.	63
5.1	Some information about the originate tree of Extract H11588.	66



## FIGURES

Pages

2.1	Schematic presentation of the essential components			
	of a two-component system.	13		
2.2	Prokaryotic two-component signal transduction system.	14		
2.3	Map of Sabah shows the location of Danum Valley Conservation Area			
	from Lahad Datu.	19		
3.1	Summary of methods involved in the research.	23		
3.2	Location of sampling site from Segama River.	25		
3.3	Newbery Plot and locations of collected soil samples.	26		
3.4	Serial dilution and isolation.	31		
3.5	Patterns of streaks made onto the Oatmeal media.	33		
3.6	Steps involved in Gram Staining procedure.	36		
3.7	Expected result for screening.	40		
3.8	Position of paper discs with control and different extracts on			
	screening media.	45		
4.1	Inhibition zones on control (streptomycin) in different			
	concentration of Na <sup>+</sup> .	58		



## PLATES

## Pages

2.1	Segama River.	18
4.1	The collected soil samples were stored in universal bottles.	48
4.2	Features of some trees showing the origins of soil samples that were	
	taken from the sampling site at Danum Valley.	48
4.3	Some features of isolates on Oatmeal media.	
	Aerial mycelia colour (left) and substrate mycelia colour (right).	50
4.4	20% glycerol stock solutions were kept in 7 ml bijon bottles.	54
4.5	Features of some selected slant agars.	54
4.6	Some features of extracts kept in universal bottles.	55
4.7	Inhibition zones on different concentration of $K^+$ in M9 minimal media.	59
4.8	Inhibition zones occurred on both low (left) and high (right)	
	concentration of K <sup>+</sup> .	61
4.9	Screening result on extract from H11588 strain in low (left) and	
	high (right) concentration.	62



### SYMBOLS, UNITS, ABBREVIATIONS, TERMS & FORMULAS

- $\mu$  micro (10<sup>-6</sup>)
- % percentage
- °C degree Celsius
- g gram
- mg milligram
- kg kilogram
- cm centimeter
- km kilometer
- m meter
- M Molar
- ml milliliter
- L Liter
- K<sup>+</sup> potassium ion
- Na<sup>+</sup> sodium ion
- No. number
- DV Danum Valley
- Std. Standard
- Mr. Mister
- Ms. Miss
- Prof. Professor
- HV Humic acid Vitamin B
- r.p.m Rotation per minute
- ISO1 Isolation using Humic acid-Vitamin B agar media
- PUR1 Purification using Oatmeal agar media
- D extract was diluted
- UD no dilution on extract
- w/v weight over volume
- v/v volume over volume



### CHAPTER 1

### **INTRODUCTION**

The need for novel approaches to control the regulation of tuberculosis is increasing. Approximately, 2–3 million people die because of this disease and it is believed that one-third of the world's population have been infected with *Mycobacterium tuberculosis*. The only widely used vaccine for tuberculosis is *Mycobacterium bovis* BCG, however, does not provide adequate levels of protection in Africa, India, and some parts of the USA (Rickman *et. al.*, 2004). Moreover, the increasing of drug-resistant strains of *M. tuberculosis* makes the identification of new drug targets and vaccines more challenging. One approach to combat tuberculosis is to identify new potential inhibitor via screening method that could inhibit a specific two-component system in *Mycobacterium tuberculosis*. Many bacteria, including *Mycobacterium*, use two-component signal transduction systems to allow them to respond rapidly to changes in their environment such as pH, temperature and osmotic stress, and control a variety of bacterial processes.



Potassium  $(K^{+})$  and sodium  $(Na^{+})$  were used to create the osmotic effects. Potassium is an essential macronutrient for the growth of most organisms. In bacteria, potassium plays an important role in the maintenance of intracellular pH and cell turgor. The second signal is sodium which also basically similar to potassium in terms of osmotic effect. The following are the steps in brief, which involved in screening. The bacteria were grown in the modified M9 minimal media (refer Table 3.3) with high and low concentrations of Na<sup>+</sup>. Glucose (as the carbon source) and thiamine hydrochloride (Vitamin B<sub>1</sub>) were added. Seed culture (liquid culture) of *Mycobacterium smegmatis* was prepared using the modified M9 minimal media and the bacteria were grown at 37°C temperature for 3-4 days. The screening step (refer Appendix B) subsequently took place in the same but solid (agar) media. Extracts were pipetted to different paper disc with 0.6 cm diameter. Streptomycin was used as the negative control. The media was comprised of two layers of agar- the upper layer (0.7 % w/v) and the bottom layer (1.0 % w/v). Glucose and thiamine hydrochloride (Vitamin B1), as well as the seed culture of Mycobacterium smegmatis were added to upper layer only. The media were incubated at 37°C temperature for 4 days. These steps were repeated for studying K<sup>+</sup> signal (refer Appendix C) in modified M9 minimal media (refer Table 3.4) over osmotic stress.

For this work, the soil samples were collected from specific location of the Danum Valley Conservation Area. The Danum Valley Conservation Area is one of the last remaining preserves of primary lowland rainforest in Asia. Therefore, the search for novel potential inhibitors are greater as Danum Valley is one of Sabah's last strongholds of undisturbed lowland rain forest.



This work is purposely to identify a potential antimicrobial compound that inhibits the regulation of two-component system in *Mycobacterium tuberculosis* over osmotic stress. The scope is to grow *M. smegmatis* using the modified M9 minimal media and to identify potential inhibitor(s) in low concentration of potassium ( $K^+$ ) and sodium (Na<sup>2+</sup>) ions under optimum temperature of 37°C and almost neutral condition pH of 6.7. Other objectives of this work are (i) to collect a few soil samples from the virgin tropical rainforest of Danum Valley Conservation Area in Sabah, (ii) to isolate and to purify bacteria from the collected soil samples using respectively humic acid-Vitamin B agar and Oatmeal agar, (iii) to extract secondary metabolites from actinomycetes through fermentation process, (iv) to develop sufficient media for screening of actinomycetes for both signals, K<sup>+</sup> and Na<sup>+</sup>, over osmotic pressure, (vi) to implement screening method of actinomycete for inhibitors of two-component system in *Mycobacterium smegmatis*, and finally, (vii) to practice screening method on extracts of actinomycetes.



### CHAPTER 2

### LITERATURE REVIEW

#### 2.1 Actinomycetes

Actinomycetes are bacteria belonging to the order *Actinomycetales* and are characterized by the formation of branching filaments giving them a fungal appearance. Actinomycetes are widespread in nature and can be separated into two subgroups: oxidative forms, found mostly in soil habitants, and fermentative forms, living in natural cavities of man and animals. Actinomycete is a diverse and a large group of grampositive filamentous and/or branching bacilli. These bacteria are in a form of Fungi-like bacteria that produce long, thread-like branched filaments that look like spider webs stretching through the compost. The earthy smell of compost is caused by actinomycetes. They are the primary decomposers of tough plant materials like bark, newspapers and woody stems. Actinomycetes are especially effective at attacking tough, raw plant tissues (cellulose, chitin and lignin), and softening them up for their less enterprising relatives. Most actinomycetes form spores, the manner of spores varies and is used in separating subgroups. The DNA base compositions of most members fall within the range of 68-



78% GC, and organisms at the upper end of this range have the highest GC percentage of any bacteria known (Madigan *et al.*, 2003).

Actinomycetes are saprophytes whose compost organic compounds in soil and act as parasite in hosts including human, which caused diseases like tuberculosis and leprosi. Species under this group are always in worldwide scientists' view because of their potential in producing antibiotics. For example, streptomycin from Streptomyces griseus for treatment of tuberculosis, which caused by Mycobacterium tuberculosis, and immunosuppressive drugs of takrolimus (FK506), which produced by Streptomyces tsukubaensis (Lo et al., 2002). Actinomycetes are everywhere in nature, correct identification of Nocardia species is essential to rule out Streptomyces species that is usually considered nonpathogenic. Growth rate is from few days to 4 weeks. Nocardia species must also be compared with rapid grower mycobacteria that may mimic Nocardia both by macroscopically (culture) and microscopically (direct specimen or culture). Simple tests that are necessary to identify actinomycetes are growth rate, susceptibility to antibacterial agents, morphology information, distinct musty odor Gram stain, Modified Kinyoun Stain (MK), Ziehl-Neelsen Stain (ZN), Casein Hydrolysis and growth on selective and differential medium such as sodium pyruvate (PYR) for branching and fragmentation by Nocardia species. A recent study of two organisms (Stachybotrys chartarum and Streptomyces califonicus) commonly isolated from moisture-damaged structures indicated the organisms might increase their overall biomass when grown together in the laboratory as opposed to when each organism is grown individually. They are potentially hazardous individually and may also present another ser of problems when



growing together indoors, especially for prolonged periods of time. In contrast to the harmful effects, the actinomycetes also produce a majority of antibiotics that are beneficial to humans and animals (Rau, 2004).

### 2.1.1 Nocardia

An allergic pneumonitis called farmer's lung occurs among agricultural workers who have inhaled dust from moldy plant material. It has been traced to at least three actinomycetes - Actinomyces, Nocardia and Streptomyces. The most common organism in this group is Nocardia that is responsible to cause a variety of infections including Mycetoma. Sulfur granules may be seen by the naked eye in the specimen and under the microscope during direct microscopic examination of the specimen stained by H & E stain. In contrast to Actinomyces, species of Nocardia are inhabitants of the soil rather than commensals in animals and they are aerobic. Nocardia species are gram-positive and two species are pathogenic for man -N. asteroides, and N. brasiliensis, and are somewhat acid-fast. There are two common modes of infection by Nocardia, pulmonary nocardiosis arises from inhalation of the organisms, whereas chronic subcutaneous absesses (mycetomas) arise from contamination of skin wounds, usually on the feet, and hands of labourers. In pulmonary nocardiosis, the lesions may simulate miliary or pulmonary tuberculosis. N. asteroides lie scattered though the abscesses in the form of tangled, fine, branching filaments. Aggregations into granules do not occur. Different species of Nocardias are associated with mycetomas in different parts of the world, such as N. brasiliensis in Mexico. These abscesses are clinically very similar to those of



*Streptomyces* and to various fungi. *Nocardia* are widely distributed throughout temperate and tropical climates. The diseases it causes are seen frequently in association with immunosuppression or underlying chronic diseases such as Hodgkin's disease. Once nocardiosis becomes clinically evident, it tends to become progressive and fatal, even with aggressive therapy: about 50% of patients succumb. Various antibacterial drugs are used in the treatment of nocardiosis, and sulfonamides are reported to be most effective.

### 2.1.2 Actinomyces

Actinomycosis generally arise from endogenous inhabitants of the oral cavity, whereas nocardiosis results from inhalation of soil organisms. Several species of Actinomyces have been implicated as the cause of actinomycosis in humans and animals. A. israelli is usually responsible for disease in man. It is part of the normal oral flora. It can be cultured from the majority of human tonsils and is nearly always found in scrapings of gums and teeth. The conditions that lead the organism to become invasive are not definitely known but may involve trauma and dental surgery. In general, actinomycotic infections are accompanied by a mixed flora of gram-negative bacteria (actinobacillus, Eikenella, Fusobacterium, and Bacteroides). Actinomycosis is distributed worldwide but is relatively rare. Its incidence is higher in men than women and in persons over 20. Actinomycosis is characterised by chronic destructive abscesses of connective tissues; the abdomen (especially the caecum and appendix), the lungs, the chest wall, and the face and neck may be involved. Wherever the lesions occur, they are basically the same. Abscesses expand into contiguous tissues and eventually form



burrowing, tortuous sinuses to the skin surface, where they discharge purulent material. When pus from an abscess or infected sputum is examined carefully, yellow sulphur granules are occasionally seen. These are small colonies of actinomycetes, which may be up to several millimetres in diameter. Detection of granules is not required to establish a diagnosis of actinomycosis but their presence facilitates identification of the organism. Penicillin is the drug of choice. The organism is also sensitive to tetracycline.

### 2.1.3 Streptomyces

Streptomyces are characterised by the stability of their filaments and by the formation of spores on the aerial mycelia and by the formation of spores on the aerial mycelia that project above the surface of the culture medium. With increasing appreciation of the distinction between norcardiae and streptomycetes, it has been realised that both cause actinomycotic abscesses. Because streptomycetes are ubiquitous in soil, infection is attributed to contamination of scratches and penetrating wounds. Mycetomas caused by streptomycetes are indistinguishable clinically from other actinomycetes.

### 2.1.4 Microbial secondary metabolites from actinomycetes

Actinomycetes represent the microbial group richest in production of variable secondary metabolites. These mostly bioactive molecules are the end products of complex multi-step biosynthetic pathways. Recent progress in the molecular genetics and



biochemistry of the biosynthetic capacities of actinomycetes enables first attempts to redesign these pathways in a directed fashion. However, in contrast to several examples of designed biochemical improvement of primary metabolic processes in microorganisms, none of the products or strains derived from pathway engineering in actinomycetes discussed herein have reached pilot or production scale. The main reasons for this slow progress are the complicated pathways themselves, their complex regulation during the actinomycete cell cycle, and their uniqueness, as most pathways and products are specific for a strain rather than for a given species or larger taxonomic group (Piepersberg, 1994).

Microbial secondary metabolites include antibiotics, pigments, toxins, effectors of ecological competition and symbiosis, pheromones, enzyme inhibitors. immunomodulating agents, receptor antagonists and agonists, pesticides, antitumor agents and growth promoters of animals and plants. They have a major effect on the health, nutrition and economics of our society. They often have unusual structures and their formation is regulated by nutrients, growth rate, feedback control, enzyme inactivation, and enzyme induction. Regulation is influenced by unique low molecular mass compounds, transfer RNA, sigma factors and gene products formed during postexponential development. The synthases of secondary metabolism are often coded by clustered genes on chromosomal DNA and infrequently on plasmid DNA. Unlike primary metabolism, the pathways of secondary metabolism are still not understood to a great degree and thus provide opportunities for basic investigations of enzymology, control and differentiation. Secondary metabolism is brought on by exhaustion of a



#### REFERENCES

- Alves, A. M. C. R. (published year is not mentioned). Glucose metabolism in actinomycetes.
- Chang, C., and Stewart, R. C., 1998. The Two-Component System. *Plant Physiology* 117, 723-731.
- Barrett, J. F., and Hoch, J. A., 1998. Two-Component Signal Transduction as a Target for Microbial Anti-Infective Therapy. ANTIMICROBIAL AGENTS AND CHEMOTHERAPY 42 (7), 1529-1536.
- Demain, A. L., 1998. Induction of microbial secondary metabolism. INTERNATL MICROBIOL 1, 259–264.
- Deschenes, R. J., Lin, H., Ault, A. D., and Fassler, J. S., 1999. Antifungal Properties and Target Evaluation of Three Putative Bacterial Histidine Kinase Inhibitors. *ANTIMICROBIAL AGENTS AND CHEMOTHERAPY* **43** (7), 1700-1703.
- Euverink, G. J.W. (published year is not mentioned). AROMATIC AMINO ACID BIOSYNTHESIS IN ACTINOMYCETES.
- Fontan, P. A., Walters, S., and Smith, I., 2004. Cellular signaling pathways and transcriptional regulation in *Mycobacterium tuberculosis*: Stress control and virulence. *CURRENT SCIENCE* 86 (1), 122-134.
- Galperin, M. Y., Nikolskaya, A. N., Koonin, E. V., 2001. Novel domains of prokaryotic two-component signal transduction systems. *FEMS Microbiology Letters* 203, 11-21.



- Gaβel, M., and Altendorf, K., 2001. Analysis of KdpC of the K1-transporting KdpFABC complex of *Escherichia coli*. Eur. J. Biochem. FEBS 268, 1772-1781.
- Lo, C. W., Lai, N. S., Cheah, H-Y., Wong, N. K. I., and Ho, C. C., 2002. Actinomycetes isolated from soil samples from the Crocker Range Sabah. ARBEC.
- Madigan, M. T., Martinko, J. M., and Parker, J., 2003. Brock Biology of Microorganisms. Ed 10. Prentice Hall, USA.
- Oskay, M., Tamer, A. Ü., and Azeri, C., 2004. Antibacterial activity of some actinomycetes isolated from farming soils of Turkey. *African Journal of Biotechnology* **3** (9), 441-446.
- Parkinson, J. S., 1995. Genetic Approaches for Signaling Pathways and Proteins. Two-Component Signal Transduction. Hoch, J. A., and Silhavy, T. J. American Society for Microbiology, Washington, D. C.
- Piepersberg, W., 1994. Pathway engineering in secondary metabolite-producing actinomycetes. Crit Rev Biotechnol. 14 (3), 251-85.
- Pérez, E., Samper S., Bordas, Y., Guilhot, C., Gicquel, B., and Martin, C., 2001. An essential role for phoP in Mycobacterium tuberculosis virulence. Molecular Microbiology 41 (1), 179-187.
- Rau, S. M., 2004. In The Air Newsletter January 2004.
- Rickman, L., Saldanha, J. W., Hunt, D. M., Hoar, D. N., Colston, M. J., Millar, J. B. A., and Buxtona, R. S., 2004. A two-component signal transduction system with a PAS domain-containing sensor is required for virulence of *Mycobacterium*



tuberculosis in mice. Biochemical and Biophysical Research Communications 314, 259–267.

- Smith, I., 2003. Mycobacterium tuberculosis Pathogenesis and Molecular Determinants of Virulence. CLINICAL MICROBIOLOGY REVIEWS 16 (3), 463-496.
- Stephenson, K., and Hoch, J. A., 2002. Two-component and phosphorelay signaltransduction systems as therapeutic targets. Elsevier Science Ltd, USA.
- Tyagi, J. S., and Sharma, D., 2004. Signal transduction systems of mycobacteria with special reference to *M. tuberculosis*. *CURRENT SCIENCE* **86** (1), 93-102.
- Uozumi, N., 2001. *Esherichia coli* as an expression system for K<sup>+</sup> transport systems from plants. *AJP-Cell Physiol* **281**, 733-739.
- Véscovi, E. G., Ayala, Y. M., Cera, E. D., and Groisman, E. A., 1997. Characterization of the Bacterial Sensor Protein PhoQ. *THE JOURNAL OF BIOLOGICAL CHEMISTRY* 272 (3), 1440-1443.
- Via, L. E., Curcic, R., Mudd, M. H., Dandayuthapani, S., Ulmer, R. J., and Deretic, V. J., 1996. Elements of Signal Transduction in *Mycobacterium tuberculosis*: In Vitro Phosphorylation and In Vivo Expression of the Response Regulator MtrA. *Journal of Bacteriology* 178 (11), 3314-3321.
- Zimmann, P., Puppe, W., and Altendorf, K., 1995. Membrane Topology Analysis of the Sensor Kinase KdpD of Escherichia coli. The American Society for Biochemistry and Molecular Biology 270 (47), 28282-28288.

