

**SCREENING FOR MICROBIAL INHIBITORS OF  
SIGNAL TRANSDUCTION PARTICULARLY  
THE AKT/GSK-3 $\beta$  PATHWAY**

**FOO SEK HIN**

PERPUSTAKAAN  
UNIVERSITI MALAYSIA SABAH

**THESIS SUBMITTED IN FULFILMENT FOR THE  
DEGREE OF MASTER OF SCIENCE**

**SCHOOL OF SCIENCE AND TECHNOLOGY  
UNIVERSITI MALAYSIA SABAH**

**2006**



**UMS**  
UNIVERSITI MALAYSIA SABAH

UNIVERSITI MALAYSIA SABAH

BORANG PENGESAHAN STATUS TESIS®

JUDUL: SCREENING FOR MICROBIAL INHIBITORS OF SIGNAL TRANSDUCTION PARTICULARLY THE AKT/GSK-3 $\beta$  PATHWAY

IJAZAH: SARJANA SAINS (BIOTEKNOLOGI)

SESI PENGAJIAN: 2002 – 2006

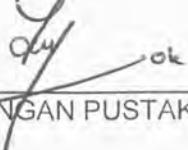
Saya, FOO SEK HIN mengaku membenarkan tesis Sarjana ini disimpan di Perpustakaan Universiti Malaysia Sabah dengan syarat-syarat kegunaan seperti berikut:

1. Tesis adalah hak milik Universiti Malaysia Sabah.
2. Perpustakaan Universiti Malaysia Sabah dibenarkan membuat salinan untuk tujuan pengajian saya.
3. Perpustakaan dibenarkan membuat salinan tesis ini sebagai bahan pertukaran antara institusi pengajian tinggi
4. TIDAK TERHAD

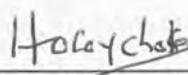
Disahkan oleh

  
(Penulis: FOO SEK HIN)

PENPUTAKAAN  
UNIVERSITI MALAYSIA SABAH

  
(TANDATANGAN PUSTAKAWAN)

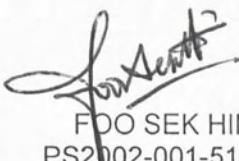
Alamat:  
Sekolah Sains dan Teknologi  
Universiti Malaysia Sabah  
Beg Berkunci 2073  
88999 Kota Kinabalu  
Sabah, MALAYSIA.

  
(Penyelia: Prof. Dr. Ho Coy Choke)

CATATAN:® Tesis dimaksudkan sebagai tesis ijazah Doktor Falsafah dan Sarjana secara penyelidikan atau disertasi bagi pengajian secara kerja kursus dan penyelidikan, atau laporan Projek Sarjana Muda (LPSM)

**DECLARATION**

The materials in this thesis are original except for quotations, excerpts, summaries and references which have been duly acknowledged.



FOO SEK HIN  
PS2002-001-514  
27 September 2006

PERPUSTAKAAN  
UNIVERSITI MALAYSIA SABAH

## ACKNOWLEDGEMENT

Firstly I wish to express my gratitude to my supervisor Prof. Ho Coy Choke. His scientific guidance and support in each phases of this project are crucial for the completion of this thesis.

I would like to thank Dr. Tomoko Andoh of Kumamoto University and Dr. Masaki Mizunuma of Hiroshima University for the yeast mutants and transformants, which marked an important start for this project.

This study involved many different specific tasks, which were in part performed in cooperation with other research groups. For this work, thanks go to:

WWF organization and Institute of Tropical Biology and Conservation (ITBC), UMS for granting us participation in scientific expeditions - Long Pasia, Lower Segama and Melalap.

Sabah Forest Centre (FRC) and Mr Johnny Gisil for their assistance in plant identifications.

Prof. Seow Heng Fong of the Immunology Unit (Faculty of Medicine and Health Sciences, UPM), for the facilities provided for mammalian cell culture work as well as Western blotting analyses.

Sir Philip Cohen and Dr. Jennifer Bain of University of Dundee, for the *in vitro* kinase assays of H7530 and H7667.

Dr. Laurent Meijer and Dr. Olivier Lozach of Station Biologique de Roscoff, C.N.R.S. for the testing of H7667 and H7530 against GSK-3 $\beta$  and CDK5.

I extend my thanks to colleagues Hew Chaw Sen, Puah Seok Hwa and Ong Si Mon for their friendship and support throughout my stay here. I would like to also thank other fellow postgraduates, Mr Chong Tong Seng, Mr. Lum Mok Sum and Ms Tan Su Hui for their help in many practical things.

I also thank all postgraduate students in the Immunology Unit (UPM) for creating such a friendly environment and stimulating atmosphere.

Last but not least, to my parents and the rest of the family members for their undivided support, love and patience.

This study was financially supported by IRPA and UMS Fundamental grants.

## ABSTRACT

This study aims to screen for microbial inhibitor of GSK-3 $\beta$  using yeast-based screening systems. *gsk-3* null mutant with four yeast GSK-3 homologs (MCK1, MDS1, MRK1 and YOL128C) disrupted, leads to (1) growth defect at 37°C, which is rescued in transformed H10075 (GSK-3 $\beta$ ) and H10079 (MCK1), alternatively by inclusion of 1.2M D-sorbitol into the growth medium. (2) Inability to utilize galactose for growth (at 25°C and 37°C), which is suppressed in H10079 (MCK1). Growth of *bul1bul2* mutant is also defective at 37°C, but distinguished in the inability to utilize glycerol. Both phenotypes of *bul1bul2* mutant are suppressed in H10082 (BUL1). Inhibitor of GSK-3 $\beta$  was therefore expected to mimic phenotypes of *gsk-3* null mutant, detected by (A) growth inhibition of H10075 (GSK-3 $\beta$ ) and H10079 (MCK1) only at 37°C, confirmed in inhibition of H10084 (MCK1) only at 37°C, and non-inhibition of H10085 ( $\Delta$ mck1). (B) Growth inhibition of H10079 (MCK1) in galactose medium (at 25°C and 37°C) and (C) Non-inhibition of H10082 (BUL1) in glycerol medium (at 25°C and 37°C). Thirty actinomycetes strains previously screened as toxic to yeast were tested on H10075 (GSK-3 $\beta$ ). Two strains H7530 and H7667 inhibited H10075 (GSK-3 $\beta$ ) only at 37°C, with minimal 1.2M D-sorbitol rescue. Identical inhibition was observed when H7530 and H7667 were tested on H10079 (MCK1) and H10084 (MCK1). In galactose utilization screening, both strains inhibited H10079 (MCK1) in SG-Ura medium, 25°C with stronger inhibition at 37°C. Another 287 actinomycetes strains, isolates from soils of Long Pasia, Lower Segama and Melalap (Crocker Range) were screened on H10075 (GSK-3 $\beta$ ) for inhibition at only 37°C, and only two weak inhibitors were found H11329 and H11364. These weak inhibitors were not further studied. Crude extracts of H7530 and H7667 did not inhibit GSK-3 $\beta$  activity *in vitro*. When tested *in vivo* using Western blotting, H7667 inhibited p-GSK-3 $\beta$  (Ser-9) in MCF-7 cells. H7530 had no effect. H10082 (BUL1) and PP1 screenings distinguished these two strains. H7530 inhibited H10082 (BUL1) in glycerol medium at 25°C and 37°C and inhibited H10018 (GLC7) of PP1 screening at 25°C and 37°C. H7667 was negative in the H10082 (BUL1) screen and inhibited H10018 (GLC7) only at 37°C resembled inhibition seen on H10075 (GSK-3 $\beta$ ) at 37°C. H7530 and H7667 were purified and bioactive peaks were identified. H7530 (F6) did not reduce p-GSK-3 $\beta$  (Ser-9) in MCF-7 cells. H7667 (F21) inhibited p-GSK-3 $\beta$  (Ser-9) but did not alter p-Akt (Thr-308) indicating H7667 (F21) could probably act directly at the Akt without affecting upstream kinases of Akt. This was substantiated by decrease in p-BAD (Ser-136). Though *in vitro* Akt assay did not detect any significant inhibition, H7667 (F21) may inhibit Akt in a non-ATP competitive manner or by other novel mechanisms. Despite the fact that no inhibitor of GSK-3 $\beta$  was found in this study, an inhibitor, H7667 (F21) that affects the Akt/GSK-3 $\beta$  pathway was identified. This justifies further investigation of H7667 (F21); to determine its specific kinase target and chemical identification of the active compound.



## ABSTRAK

### MENYARING PERENCAT MIKROBIAL TERHADAP TRANSDUKSI ISYARAT TERUTAMANYA LALUAN AKT/GSK-3 $\beta$

Kajian ini bertujuan untuk menyaring perencat mikrobial GSK-3 $\beta$  dengan sistem penyaringan yis. Mutan nul gsk-3 yang dibuntukan empat homolognya (MCK1, MDS1, MRK1 dan YOL128C) akan menyebabkan (1) kecacatan pertumbuhan pada 37°C, dipulihkan dalam transforman H10075 (GSK-3 $\beta$ ) and H10079 (MCK1), atau dengan penambahan 1.2M D-sorbitol ke dalam medium pertumbuhan. (2) Ketidakupayaan menggunakan galaktos untuk pertumbuhan (25°C and 37°C), dipulihkan dalam H10079 (MCK1). Pertumbuhan H10082 (BUL1) juga dihalang pada 37°C, tetapi dibezaikan dengan ketidakupayaan menggunakan gliserol. Kedua-dua fenotip ini dipulihkan dalam H10082 (BUL1). Maka perencat GSK-3 $\beta$  dijangka akan (A) merencatkan H10075 (GSK-3 $\beta$ ) and H10079 (MCK1) hanya pada 37°C, dipastikan dengan perencatan H10084 (MCK1) hanya pada 37°C and ketiadaan perencatan pada H10085 ( $\Delta$ mck1). (B) Perencatan H10079 (MCK1) di medium galaktos (25°C and 37°C) dan (C) ketiadaan perencatan pada H10082 (BUL1) di medium gliserol (25°C and 37°C). Tiga puluh strain aktinomiset yang didapati toksik terhadap yis diuji pada H10075 (GSK-3 $\beta$ ). Dua strain H7530 and H7667 didapati merencatkan H10075 (GSK-3 $\beta$ ) hanya pada 37°C, dengan pemulihan 1.2M D-sorbitol minima. Perencatan adalah identikal apabila diuji pada H10079 (MCK1) and H10084 (MCK1). Kedua-dua strain merencat H10079 (MCK1) dalam SG-Ura, dengan perencatan yang lebih kuat pada 37°C. Sebanyak 287 strain aktinomiset dipencarkan daripada tanah Long Pasia, Hilir Segama dan Melalap (Banjaran Crocker) disaring terhadap H10075 (GSK-3 $\beta$ ) untuk perencatan hanya pada 37°C, dan hanya dua perencat lemah ditemui, H11329 and H11364. Kedua perencat lemah ini tidak diuji secara lanjut. Ekstrak kasar H7530 and H7667 tidak merencat aktiviti *in vitro* GSK-3 $\beta$ . Dalam Western blotting, H7667 merencatkan p-GSK-3 $\beta$  (Ser-9) sel MCF-7. H7530 tidak memberi sebarang kesan. Penyaringan H10082 (BUL1) and PP1 membezakan kedua-dua strain. H7530 merencatkan H10082 (BUL1) di medium gliserol pada 25°C and 37°C, dan merencatkan H10018 (GLC7) pada 25°C dan 37°C. H7667 adalah negatif dalam penyaringan H10082 (BUL1) dan merencatkan H10018 (GLC7) hanya pada 37°C, menyerupai perencatan dalam H10075 (GSK-3 $\beta$ ) pada 37°C. Fraksi aktif H7530 and H7667 diasingkan dan peak bioaktif dikenalpasti. H7530 (F6) tidak merencatkan p-GSK-3 $\beta$  (Ser-9). H7667 (F21) merencatkan p-GSK-3 $\beta$  (Ser-9) tetapi tidak mengubah p-Akt (Thr-308), menandakan H7667 (F21) mungkin merencat Akt tanpa mempengaruhi kinase di atas Akt. Ini dibuktikan dalam pengurangan p-BAD (Ser-136). Asai *in vitro* Akt tidak menunjukkan perencatan yang ketara, yakni H7667 (F21) mungkin merencat Akt bukan secara ATP-kompetitif atau melalui mekanisma lain yang baru. Sungguhpun tiada perencat GSK-3 $\beta$  ditemui, kajian ini menemui H7667 (F21) yang merencatkan laluan Akt/GSK-3 $\beta$ , maka adalah penting untuk mengenalpasti kinase sasaran spesifik serta pengenalan struktur kimia bagi sebatian aktif tersebut.



## ABBREVIATIONS

Akt	protein kinase B (also called PKB)
AMPK	AMP-activated protein kinase
ATP	adenosine 5'-triphosphate
BSA	bovine serum albumin
CAMK	calmodulin-dependent protein kinase
CDK	cyclin-dependent protein kinase
CHK	checkpoint kinase
CK	casein kinase
CREB	cAMP-response element binding protein
CSK	C-terminal Src kinase;
DTT	dithiothreitol
DYRK	dual-specificity, tyrosine-phosphorylated and regulated kinase;
EGF	epidermal growth factor
ERK	extracellular-signal-regulated kinase
Exp.	Experiment
FBS	fetal bovine serum
g	gram
GAPDH	glyceraldehyde-3-phosphate dehydrogenase
GSK-3	glycogen synthase kinase-3
hr	hour(s)
HV	humic acid-B vitamins
IC <sub>50</sub>	concentration giving half maximal inhibition
IDV	Integrated density value
IGF-I	insulin-like growth factor I
ILK	integrin-linked kinase
JNK	c-Jun N-terminal kinase
kDa	kilodalton
L	litre
LCK	lymphocyte kinase
M	molar
MAPK	mitogen-activated protein kinase
MAPK	mitogen activated protein kinase
MAPKAP-K1	MAPK-activated protein kinase-1

MAPKAP-K2	MAPK-activated protein kinase 2
mg	milligram
min	minute
MKK	MAPK kinase (also called MEK)
MLCK	myosin light chain kinase
mm	millimeter
MNK	MAPK-integrating kinase
$M_r$	relative molecular mass (unitless)
MSK1	mitogen- and stress-activated protein kinase 1
mTOR	mammalian target of rapamycin
nM	nanomolar
PAGE	polyacrylamide gel electrophoresis
PBS	phosphate-buffered saline
PDGF	platelet-derived growth factor
PDK1	3-phosphoinositide-dependent protein kinase 1
PHK	phosphorylase kinase
PHLPP	PH domain Leucine-rich repeat protein phosphatase
PI3-kinase	phosphatidylinositol 3-kinase
PKA	cAMP-dependent protein kinase
PKC	protein kinase C
PKG	cGMP-dependent protein kinase
PMSF	phenylmethylsulfonyl fluoride
PRAK	p38-regulated/activated kinase
PRK	protein kinase C-related protein kinase
ROCK	Rho-dependent protein kinase
S6K1	p70 ribosomal protein S6 kinase
SAPK	stress-activated protein kinase
SAPK2a	stress-activated protein kinase 2a (also called p38)
SAPK2b	stress-activated protein kinase 2b (also called p38b2)
SAPK3	stress-activated protein kinase 3 (also called p38c)
SAPK4	stress-activated protein kinase 4 (also called p38d)
SGK	serum- and glucocorticoid-induced kinase;
$t_R$	retention time
$\mu\text{g}$	microgram
$\mu\text{l}$	microlitre
$\mu\text{M}$	micromolar

## TABLE OF CONTENTS

	Page No.
<b>RESEARCH TITLE</b>	I
<b>DECLARATION</b>	II
<b>ACKNOWLEDGEMENT</b>	III
<b>ABSTRACT</b>	IV
<b>ABSTRAK</b>	V
<b>ABBREVIATIONS</b>	VI
<b>TABLE OF CONTENTS</b>	VIII
<b>LIST OF TABLES</b>	XIII
<b>LIST OF FIGURES</b>	XVI
<b>CHAPTER 1: INTRODUCTION</b>	1
1.1    Small molecule inhibitors of signal transduction: sources and applications	1
1.2    Kinase inhibitors: From receptor protein tyrosine kinases to serine/threonine kinases	6
1.2.1    Inhibitors of the Akt/GSK-3 $\beta$ pathway	9
1.3    Molecular-targeted screening using the yeast – <i>Saccharomyces cerevisiae</i>	11
1.4    Objectives of study and significant results	14
<b>CHAPTER 2: LITERATURE REVIEW</b>	18
2.1    Glycogen synthase kinase-3 (GSK-3)	18
2.1.1    Structural aspects of GSK-3 on its activation and substrate specificity	20
2.1.2    Mechanism of inhibition by Ser-9 phosphorylation	22
2.1.3    GSK-3 and diabetes	23
2.1.4    GSK-3 and Wnt signaling	25
2.1.5    Small molecule inhibitors that affect GSK-3	29
2.1.5.1    Lithium	29
2.1.5.2    Natural inhibitors of GSK-3 $\beta$	30
2.1.5.3    Other synthetic inhibitors of GSK-3	35
2.2    Akt/Protein kinase B	40



2.2.1	Structural aspects of PKB (Akt) on its activation and substrate specificity	41
2.2.2	Mechanism of activation and self-inhibition of PKB	42
2.2.3	Role of Akt in diabetes	44
2.2.4	Role of Akt in cancer	45
2.2.5	Small molecule inhibitors of Akt	48
	2.2.5.1 Natural inhibitors of Akt signaling pathway	48
	2.2.5.2 Synthetic small molecule inhibitors of Akt	50
2.3	Akt/GSK-3 $\beta$ pathway in yeast signal transduction	54
2.3.1	Identification of yeast GSK-3 homologs and its physiological roles in yeast	54
	2.3.1.1 Involvement of MCK1 and BUL1 in cell integrity pathway	58
	2.3.1.2 MCK1 and BUL1 in utilization of alternative carbon sources and glycogen accumulation	60
2.3.2	Identification of yeast Akt homolog and its role in longevity and stress resistance	62
2.3.3	Identification of upstream kinases of Akt in yeast and their physiological functions	64
2.4	Soil actinomycetes: its biodiversity and potential source of novel inhibitors of signal transduction	66

### **CHAPTER 3: MATERIALS AND METHODS**

3.0	Outline of methodology used in study	69
3.1	Materials	70
	3.1.1 Reagents for microbial isolation, culture and screening	70
	3.1.2 Instrument and materials for extraction and HPLC work	70
	3.1.3 Reagents and materials for mammalian cell culture	71
	3.1.4 Instruments and materials for Western blotting	71
	3.1.5 Antibodies	72
	3.1.6 Cell line and yeast strains	72
	3.1.7 Actinomycetes strains that were toxic to yeast as detected in one or more yeast-based screening systems	73
	3.1.8 Media, solutions and buffers	75
3.2	Methods	79
	3.2.1 Soil sampling	79



3.2.2 Isolation of actinomycetes from soil samples	79
3.2.2.1 Isolation <i>Streptosporangiaceae</i> family of actinomycetes	79
3.2.3 Production of secondary metabolites	80
3.2.4 Yeast-based screening systems	80
3.2.5 Isolation of bioactive fraction	85
3.2.5.1 Method used for purification of aqueous extract of H7530	85
3.2.5.2 Method used for purification of H7667	85
3.2.6 Mammalian cell culture	86
3.2.7 Western blotting	86
3.2.8 Cover slip method	87
3.2.9 Testing of inhibitory activity in protein kinase assays <i>in</i> <i>vitro</i>	88
<b>CHAPTER 4: RESULTS</b>	<b>101</b>
4.1 Actinomycetes from Sabah rainforests for the screening of GSK- 3 $\beta$ inhibitor	101
4.1.1 Actinomycetes previously isolated from various terrestrial sites in Sabah and screened as toxic in other yeast-based screening systems	101
4.1.2 Sampling at Long Pasia	105
4.1.3 Sampling at Lower Segama	108
4.1.4 Sampling at Melalap, Crocker Range	113
4.2 Effects of H7530 and H7667 in the yeast GSK-3 signaling pathway	117
4.2.1 Identification of H7530 and H7667 in the GSK-3 $\beta$ screen: high-temperature sensitive phenotype and sorbitol rescue	117
4.2.2 Both H7530 and H7667 inhibited MCK1 in yeast: high- temperature sensitive phenotype and galactose- sensitive phenotype	120
4.2.3 Differential specificity of H7530 and H7667 towards GSK-3 – Glycerol metabolism	125
4.2.4 Activity of H7530 and H7667 in yeast-based protein phosphatase-1 (PP1) screening	126



4.3	Effects of H7530 and H7667 on mammalian protein kinases of Akt/GSK-3 $\beta$ pathway	131
4.3.1	Crude extract of H7530 did not affect phosphorylation of GSK-3 $\beta$ (Ser-9)	131
4.3.2	Insulin-stimulated p-GSK-3 $\beta$ (Ser-9) was inhibited by crude extract of H7667	132
4.3.3	Crude extracts H7530 and H7667 did not inhibit GSK-3 $\beta$ activity <i>in vitro</i>	134
4.4	Isolation of bioactive fraction that affects GSK-3 in yeast and inhibits Akt/GSK-3 $\beta$ pathway in mammalian cells	137
4.4.1	H7530	137
4.4.1.1	HPLC fractionation of H7530	137
4.4.1.2	H7530 (F6) did not inhibit the level of p-GSK-3 $\beta$ (Ser-9)	140
4.4.2	H7667	142
4.4.2.1	HPLC fractionation of H7667	142
4.4.2.2	Effects of H7667 (F21) against protein kinases of the Akt/GSK-3 $\beta$ pathway	153
4.4.2.3	H7667 (F21) inhibited p-BAD in insulin-stimulated Akt/BAD pathway	156
4.4.2.4	Activity of H7667 (F21) against GSK-3 $\beta$ and Akt <i>in vitro</i>	157

## CHAPTER 5: DISCUSSION

5.1	Plant diversity and isolation of actinomycetes	159
5.2	Yeast-based GSK-3 screening systems	161
5.2.1	GSK-3 $\beta$ screening of strains pre-screened toxic to yeast	161
5.2.2	Specificity of GSK-3 $\beta$ screening and <i>in vivo</i> targets of H7530 and H7667 in yeast	162
5.2.2.1	H7530	162
5.2.2.2	H7667	163
5.3	Isolation of H7530 (F6) and H7667 (F19 and F21)	166
5.3.1	H7530 (F6)	166
5.3.2	H7667 (F19) and H7667 (F21)	166
5.4	Inhibition of insulin-activated Akt/GSK-3 $\beta$ and Akt/BAD pathway by H7667	168



5.4.1	H7667 inhibited Akt/GSK-3 $\beta$ pathway leading to inhibition of p-GSK-3 $\beta$ (Ser-9) and cyclin D1	168
5.4.2	H7667 did not alter phosphorylation of Akt (Thr-308) but inhibited Akt/BAD pathway	170
5.4.2.1	Compound behaving in parallel with H7667 (F21)	171
5.4.2.2	H7667 (F21) inhibited Ser-136 phosphorylation of BAD	171
5.5	H7667 (F21) against Akt and GSK-3 $\beta$ activity <i>in vitro</i>	172
5.6	Potential development of H7667 as inhibitor of Akt signaling pathway	174
<b>CHAPTER 6: CONCLUSION</b>		177
6.1	H7667 (F21) – suggestions for future investigation	177
<b>REFERENCES</b>		180
<b>APPENDICES</b>		196



## LIST OF TABLES

Table No.	Title	Page
<b>Table 2.1</b>	Inhibition of protein kinases by lithium (10mM)	30
<b>Table 2.2</b>	Kinase inhibition selectivity of hymenialdisine	31
<b>Table 2.3</b>	Kinase inhibition selectivity table of indirubin-3'-monoxime	33
<b>Table 2.4</b>	Kinase inhibition selectivity table of few compounds derived from staurosporine	35
<b>Table 2.5</b>	Many CDK inhibitors are good inhibitors of GSK-3 $\beta$	36
<b>Table 2.6</b>	Description of some very specific synthetic GSK-3 $\beta$ inhibitor	37
<b>Table 2.7</b>	Yeast homologs of the Akt/GSK-3 $\beta$ pathway	54
<b>Table 3.1</b>	Screening data and sampling sites of 30 actinomycetes strains pre-screened as toxic to yeast	73
<b>Table 3.2</b>	Description of Mkk1 <sup>P386</sup> , MSG5, Ras/Raf and PP1 screenings	74
<b>Table 3.3</b>	Summary of screening methods used in this study	81
<b>Table 3.4</b>	Description of yeast strains used in this study	82
<b>Table 3.5</b>	Screening systems developed for this study to detect GSK-3 $\beta$ inhibitor	84
<b>Table 4.1</b>	GSK-3 $\beta$ screening of 30 actinomycetes strains that were pre-screened as toxic to yeast	102
<b>Table 4.2</b>	Soil samples of Long Pasia	105
<b>Table 4.3</b>	Actinomycete isolates of Long Pasia	106
<b>Table 4.4</b>	GSK-3 $\beta$ screening data of Long Pasia acetone extracts	106
<b>Table 4.5</b>	Soil samples of Lower Segama	108
<b>Table 4.6</b>	Actinomycete isolates of Lower Segama	109
<b>Table 4.7</b>	GSK-3 $\beta$ screening data of Lower Segama acetone extracts	110
<b>Table 4.8</b>	GSK-3 $\beta$ screening data for H11143, H11144, H11146, H11147 and H11128 at both 25°C and 37°C	110
<b>Table 4.9</b>	GSK-3 $\beta$ screening data for H8934	111
<b>Table 4.10</b>	Soil samples of Melalap	114
<b>Table 4.11</b>	Actinomycete isolates of Melalap	114



Table 4.12	GSK-3 $\beta$ screening data of Melalap acetone extracts	115
Table 4.13	GSK-3 $\beta$ screening data for H11329, H11337, H11339, H11364, H11402 with 100 $\mu$ L of crude acetone extract	115
Table 4.14	Activity of crude acetone extracts of H7530 and H7667 against H10075 (GSK-3 $\beta$ )	117
Table 4.15	Activity of crude acetone extracts of H7530 and H7667 against D-sorbitol - supplemented H10075 (GSK-3 $\beta$ )	118
Table 4.16	Growth inhibition of H10075 (GSK-3 $\beta$ ) only at 37°C by freeze-dried crude acetone extracts of H7530 and H7667	118
Table 4.17	Growth inhibition of H10075 (GSK-3 $\beta$ ) only at 37°C by H7530 and H7667	119
Table 4.18	Activity of crude acetone extracts of H7530 and H7667 against H10079 (MCK1)	121
Table 4.19	Growth inhibition of H10079 (MCK1) only at 37°C by freeze-dried crude acetone extracts of H7530 and H7667	121
Table 4.20	Growth inhibition of H10079 (MCK1) at 37°C by H7530 and H7667	122
Table 4.21	Growth inhibition of H10079 (MCK1) by H7530 and H7667 in galactose medium, SG-Ura	123
Table 4.22	Growth inhibition of H10079 (MCK1) by H7530 and H7667 in SG-Ura, with LiCl as control	123
Table 4.23	Activity of freeze-dried crude acetone extracts of H7530 and H7667 against H10084 (MCK1) and H10085 ( $\Delta mck1$ )	124
Table 4.24	Activity of freeze-dried crude acetone extract of H7530 against H10082 (BUL1) in glycerol (5%) medium	126
Table 4.25	Activity of freeze-dried crude acetone extract of H7667 and H7530 against wild-type yeast (GLC7) and <i>glc7-10</i> mutant	128
Table 4.26	Activity of crude acetone extract of H7520 and H9318 against H10075 (GSK-3 $\beta$ )	130
Table 4.27	Screening summary for H7530 and H7667	130
Table 4.28	Activity of freeze-dried crude extract of H7530 against GSK-3 $\beta$ activity (as determined by Prof. Sir Philip Cohen and Dr. Jennifer Bain of University of Dundee, U.K.)	135
Table 4.29	Activity of freeze-dried crude extract of H7530 (1mg/ml) in the panel of 30 different protein kinases (as determined by Prof. Sir Philip Cohen and Dr. Jennifer Bain of University of Dundee, U.K.)	135



<b>Table 4.30</b>	Activity of freeze-dried crude extract of H7667 and H7530 against GSK-3 $\beta$ and CDK5 (as determined by Dr. Laurent Meijer and Dr. Olivier Lozach of Station Biologique de Roscoff, C.N.R.S.)	136
<b>Table 4.31</b>	Yeast-based GSK-3 $\beta$ screening results for HPLC fractions of H7530	139
<b>Table 4.32</b>	Yeast GSK-3 $\beta$ screening results for HPLC fractions of H7667	144
<b>Table 4.33</b>	Yeast GSK-3 $\beta$ screening result for H7667 (F19) used for Western blotting analysis	144
<b>Table 4.34</b>	Yeast GSK-3 $\beta$ screening result for H7667 (F18-F20) used for Western blotting analysis	145
<b>Table 4.35</b>	Yeast GSK-3 $\beta$ screening result for H7667 (F17-F19) used for Western blotting analysis	146
<b>Table 4.36</b>	Screening results of crude extract of H7667, before and after liquid-liquid extraction	148
<b>Table 4.37</b>	Isolation of bioactive HPLC fraction from crude n-butanol layer of H7667. Twenty-four fractions were collected, from the 10 <sup>th</sup> min to the 25 <sup>th</sup> min, 1.0ml/min	149
<b>Table 4.38</b>	Growth inhibition of H10075 (GSK-3 $\beta$ ) only at 37°C by H7667 (F21), with crude acetone extract of H7667 as control	150
<b>Table 4.39</b>	Inhibition of H10075 (GSK-3 $\beta$ ) by H7667 (F21)	152
<b>Table 4.40</b>	Partial D-sorbitol-rescued inhibition by H7667 (F21)	152
<b>Table 4.41</b>	<i>In vitro</i> GSK- $\beta$ assay of H7667 (F21)	158
<b>Table 4.42</b>	<i>In vitro</i> Akt ( $\Delta$ PH-PKB $\alpha$ -S473D) assay of H7667 (F21)	158



## LIST OF FIGURES

Figure No.	Title	Page
<b>Figure 1.1</b>	Small molecule inhibitors for bioprobing of signaling pathways	2
<b>Figure 1.2</b>	Plant-synthesized small molecule anti-proliferative agents	3
<b>Figure 1.3</b>	Anti-tumor agents produced by actinomycetes with clinical application in cancer chemotherapy	4
<b>Figure 1.4</b>	Structures of staurosporine and its analog, UCN-01	5
<b>Figure 1.5</b>	Clinically approved synthetic tyrosine kinase inhibitors for the treatment of CML (Gleevec) and NSCLC (Iressa)	7
<b>Figure 1.6</b>	Farnesyl transferase inhibitors currently in clinical trials	8
<b>Figure 1.7</b>	Microbial inhibitors of Hsp90, resulting in the inactivation of Raf kinase	8
<b>Figure 1.8</b>	Specific inhibitors of MKK1	9
<b>Figure 1.9</b>	Synthetic inhibitors of Akt being developed by Abbott and Merck	10
<b>Figure 1.10</b>	Molecular structures of GSK-3 $\beta$ inhibitors CHIR98014, CHIR98023 and CHIR99021	11
<b>Figure 1.11</b>	Immunosuppressants isolated from <i>Streptomyces sp.</i>	12
<b>Figure 2.1</b>	Schematic representation of mammalian GSK-3	19
<b>Figure 2.2</b>	Regulation of GSK-3 by the insulin signal-transduction pathway	24
<b>Figure 2.3</b>	Central role of GSK-3 in the Wnt/ $\beta$ -catenin pathway	27
<b>Figure 2.4</b>	Structures of hymenialdisine and indirubin-3'-monoxime	32
<b>Figure 2.5</b>	Structures of GF 109203X (Bis-1) and Ro 318220 (Bis-IX)	34
<b>Figure 2.6</b>	Structures of some specific synthetic GSK-3 $\beta$ inhibitors	39
<b>Figure 2.7</b>	Domain structure of PKB isoforms and variants	41
<b>Figure 2.8</b>	Schematic of signalling through the phosphatidylinositol-3-kinase (PI3K)/AKT pathway	48
<b>Figure 2.9</b>	Structure of deguelin	50
<b>Figure 2.10</b>	Structures of some specific synthetic Akt inhibitors	53

<b>Figure 2.11</b>	Model of functional interaction between GSK-3, Bul1 and Bul2, and Rsp5	57
<b>Figure 2.12</b>	Ca <sup>2+</sup> -induced cell cycle delay, mediated by Mck1 and calcineurin. This model only operates at 28°C	58
<b>Figure 2.13</b>	Working model of feedback control of cell wall integrity signaling by Bcy1 and Zds1	60
<b>Figure 2.14</b>	Conserved longevity-regulatory pathways in <i>S. cerevisiae</i> , <i>C. elegans</i> and possibly human	63
<b>Figure 3.1</b>	Outline of methodology used in this study	69
<b>Figure 4.1</b>	Colony morphology of H7530 on oatmeal agar	103
<b>Figure 4.2</b>	Colony morphology of H7667 on oatmeal agar	103
<b>Figure 4.3</b>	Sporophore morphology of H7530	104
<b>Figure 4.4</b>	Sporophore morphology of H7667	104
<b>Figure 4.5</b>	Topography map showing specific sampling locations in Long Pasia	107
<b>Figure 4.6</b>	Isolation plates (HV agar) prepared using the usual method (left) and with the 1.0% chloramine-T method (right)	109
<b>Figure 4.7</b>	Topography map showing specific sampling locations in Lower Segama	112
<b>Figure 4.8</b>	Topography map showing specific sampling locations in Melalap	116
<b>Figure 4.9</b>	Growth inhibition of H10075 (GSK-3β) only at 37°C in SC-Ura, with and without the addition of 1.2M D-sorbitol	119
<b>Figure 4.10</b>	Growth inhibition of H10079 (MCK1) only at 37°C in SC-Ura, with and without the addition of 1.2M D-sorbitol	122
<b>Figure 4.11</b>	Growth inhibition of H10079 (MCK1) by H7530 and H7667 in SG-Ura with LiCl as control	123
<b>Figure 4.12</b>	Activity of crude acetone extract of H7530 against H10084 (MCK1) and H10085 ( $\Delta mck1$ ), in YPD without addition of CaCl <sub>2</sub>	124
<b>Figure 4.13</b>	Activity of crude acetone extract of H7530 against H10082 (BUL1) in glycerol (5%) medium	126
<b>Figure 4.14</b>	Concentration-dependent inhibition of wild-type (GLC7) and mutant <i>glc7-10</i> at 37°C, by H7667 and H7530	129



<b>Figure 4.15</b>	Western blotting showing level of Ser-9 phosphorylation of GSK-3 $\beta$ in human MCF-7 cells treated with freeze-dried crude acetone extract of H7530	131
<b>Figure 4.16</b>	Non-inhibition of p-GSK-3 $\beta$ (Ser-9) in MCF-7 cells treated with higher concentration of freeze-dried crude acetone extract of H7530	132
<b>Figure 4.17</b>	Inhibitory effects of freeze-dried crude acetone extract of H7667 on the level of p-GSK-3 $\beta$ (Ser-9) in human MCF-7 cells	133
<b>Figure 4.18</b>	Concentration-dependent inhibition of p-GSK-3 $\beta$ (Ser-9) in MCF-7 cells treated with freeze-dried crude acetone extract of H7667	133
<b>Figure 4.19</b>	Freeze-dried crude acetone extract of H7667 inhibited p-GSK-3 $\beta$ (Ser-9) without any inhibition of p-Akt (Thr-308)	134
<b>Figure 4.20</b>	HPLC chromatogram of (A) blank mannitol-peptone medium and (B) Freeze-dried crude acetone extract of H7530 (400mg/ml)	138
<b>Figure 4.21</b>	HPLC chromatogram H7530 (F6) (1mg/ml)	139
<b>Figure 4.22</b>	H7530 (F6) had no effect on the level of p-GSK-3 $\beta$ (Ser-9) in MCF-7 cells	140
<b>Figure 4.23</b>	Higher concentration of H7530 (F6) on the level of p-GSK-3 $\beta$ (Ser-9) and its upstream protein kinases, p-Akt (Thr-308) and PI3K	141
<b>Figure 4.24</b>	Increased concentration of H7530 (F6) and longer treatment duration did not affect the level of Ser-9 phosphorylation of GSK-3 $\beta$	141
<b>Figure 4.25</b>	HPLC chromatogram of (A) freeze-dried crude acetone extract of H7667 and (B) H7667 (F19)	143
<b>Figure 4.26</b>	Effects of H7667 (F19) on p-GSK-3 $\beta$ (Ser-9) and p-Akt (Thr-308) in MCF-7 cells	144
<b>Figure 4.27</b>	Effects of H7667 (F18 – F20) on p-GSK-3 $\beta$ (Ser-9) in MCF-7 cells as predetermined by yeast-based screening results	145
<b>Figure 4.28</b>	Effects of H7667 (F17 – F19) on p-GSK-3 $\beta$ (Ser-9) and p-Akt (Thr-308) as predetermined by yeast-based screening results, with crude extract as positive control	146
<b>Figure 4.29</b>	HPLC chromatogram of (A) Freeze-dried crude acetone extract of H7667, 100mg/ml H <sub>2</sub> O prior to extraction, (B) n-butanol layer after extraction with (A), 1:1 and (C) Aqueous layer after extraction with n-butanol, 1:1	147



<b>Figure 4.30</b>	Isolation of bioactive HPLC fraction of crude n-butanol layer of H7667	149
<b>Figure 4.31</b>	H7667 (F21) inhibited H10075 (GSK-3 $\beta$ ) in dose-dependent manner, rescued by 1.2M D-sorbitol	150
<b>Figure 4.32</b>	HPLC chromatogram of (A) Crude n-butanol layer of H7667 (1:1) (500mg/ml H <sub>2</sub> O prior to extraction), (B) Freeze-dried bioactive fraction, 0.4mg/ml BuOH, 20 $\mu$ l injection and (C) Freeze-dried bioactive fraction, 0.4mg/ml BuOH, 40 $\mu$ l injection	151
<b>Figure 4.33</b>	Inhibition of H10075 (GSK-3 $\beta$ ) by H7667 (F21) (48 $\mu$ g)	152
<b>Figure 4.34</b>	Partial D-sorbitol-rescued inhibition by H7667 (F21) (48 $\mu$ g)	152
<b>Figure 4.35</b>	Inhibitory effects of H7667 (F21) on p-GSK-3 $\beta$ (Ser-9) in increasing concentration	153
<b>Figure 4.36</b>	H7667 (F21) inhibited p-GSK-3 $\beta$ (Ser-9) in a concentration-dependent manner without affecting level of p-Akt (Thr-308)	154
<b>Figure 4.37</b>	Inhibition of p-GSK-3 $\beta$ (Ser-9) by H7667 (F21) in increasing incubation time	154
<b>Figure 4.38</b>	H7667 (F21) inhibited expression of cyclin D1 in increasing concentration	155
<b>Figure 4.39</b>	H7667 (F21) inhibited p-BAD (Ser-136), in a concentration-dependent manner	156
<b>Figure 4.40</b>	Inhibitory effects of H7667 (F21) on p-BAD (Ser-136), in a time-dependent manner	157
<b>Figure 5.1</b>	An illustrated hypothesis of signaling proteins inhibited by H7530 and H7667	165
<b>Figure 5.2</b>	IGF-1-induced nuclear accumulation of cyclin D1 involving Akt/GSK-3 $\beta$ pathway	170



# CHAPTER 1

## INTRODUCTION

This study was designed to screen for inhibitor specific to GSK-3 $\beta$  using yeast-based screening systems. Inhibitor in this context refers to small molecule compound synthesized by actinomycetes which acts on GSK-3 $\beta$  of the insulin-activated Akt/GSK-3 $\beta$  pathway. Yeast (*Saccharomyces cerevisiae*) mutants and transformants were tools to develop systems that screen for this inhibitor.

This chapter will therefore review the functional values of some existing small molecule inhibitors, and the progressing discovery which lead to the current interest of this study.

### 1.1 Small molecule inhibitors of signal transduction: sources and applications

Earlier small molecule inhibitors of signal transduction were predominantly tools in understanding the physiological roles of protein kinases and the molecular mechanisms of protein phosphorylation (Figure 1.1). In the 1980s, research group led by Hidaka utilized synthetic H7 [1-(5-isoquinolinesulphonyl)-2-methylpiperazine], a selective inhibitor of protein kinase C (PKC) to study the role of PKC while its derivatives H88 and H89 were useful probes in determining physiological function of cAMP-dependent protein kinase (PKA) (Naito *et al.*, 1999). Fungal metabolite, wortmannin and plant flavonoid derivative LY294002 that specifically inhibit PI3K (phosphatidylinositol 3-kinase) are often used to dissect PI3K-mediated pathway.



**UMS**  
UNIVERSITI MALAYSIA SABAH

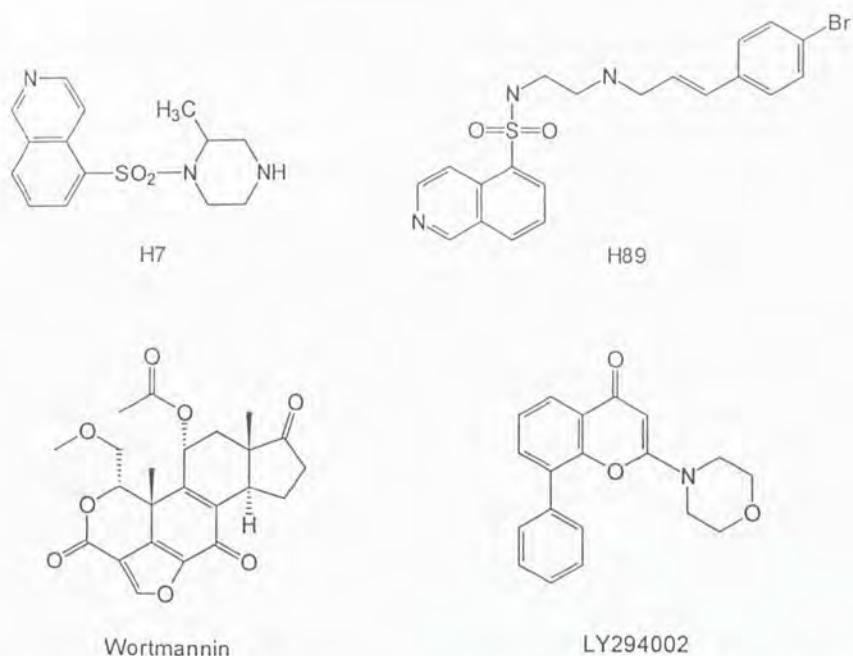


Figure 1.1: Small molecule inhibitors for bioprobing of signaling pathways

Today, apart from being biochemical probes, small molecule inhibitors have become potential therapeutics for molecular diseases such as cancer, diabetes and Alzheimer's, the very important sources of these novel compounds come from nature - terrestrial plants and terrestrial microorganisms. Notable small molecule inhibitors from plants (Figure 1.2) that exhibit antiproliferative properties are (1) flavopiridol, an alkaloid derivative from *Dysoxylum binectariferum* which inhibits CDK1, CDK2 and CDK4 (Elsayed & Sausville, 2001), (2) indirubin-3'-monoxime, originating from indigo-producing plants (*Indigo naturalis*) that inhibits CDK2 and GSK-3 $\beta$  (Bain *et al.*, 2003; Leclerc *et al.*, 2001) and (3) taxol, isolated from *Taxus brevifolia*, which promotes stabilisation of tubulin polymerization into microtubules, and also activates apoptotic signalling cascades such as JNK,3 Raf-1, and Bcl-2 family members (Broxterman & Georgopapadakou, 2005; MacKeigan *et al.*, 2002).

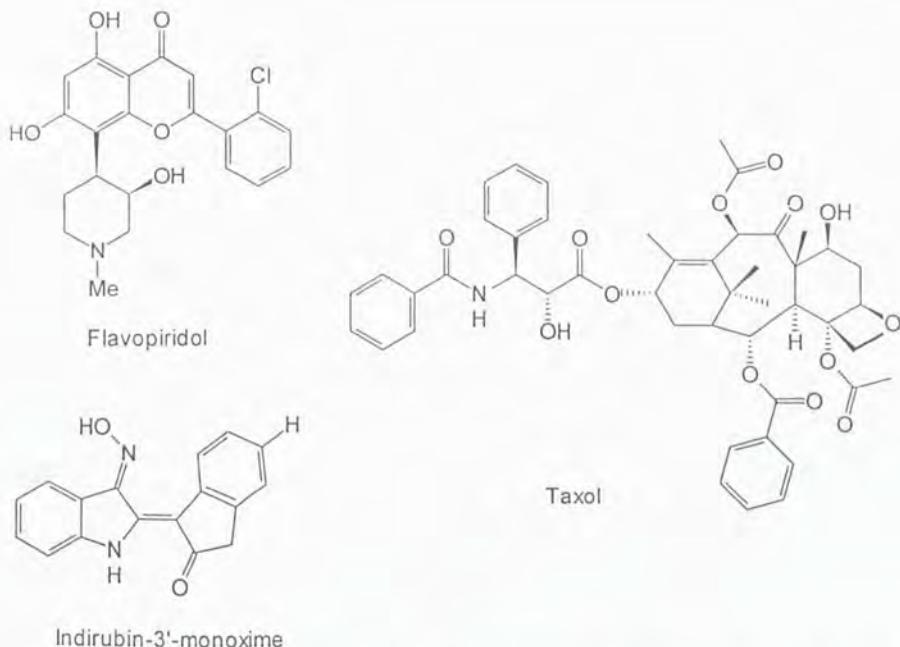


Figure 1.2: Plant-synthesized small molecule anti-proliferative agents

Microbial secondary metabolites represent a large source of compounds endowed with complex structures and potent biological activities. These compounds may not become new drugs, but many are good templates for semisynthetic and total synthetic modification that eventually developed into a drug. Therefore screening of natural products is one of the earliest steps in drug discovery, as to obtain a lead compound.

Active producers of secondary metabolites at the moment are restricted to a few groups of bacterial and eukaryotic microbes. Filamentous actinomycetes, the myxobacteria, the pseudomonads, the cyanobacteria and the eukaryotic filamentous fungi are proven to be producers of many chemically different secondary metabolites, but with the thousands of compounds described in literature, discovering new bioactive metabolites is no easy task (Donadio *et al.*, 2002).

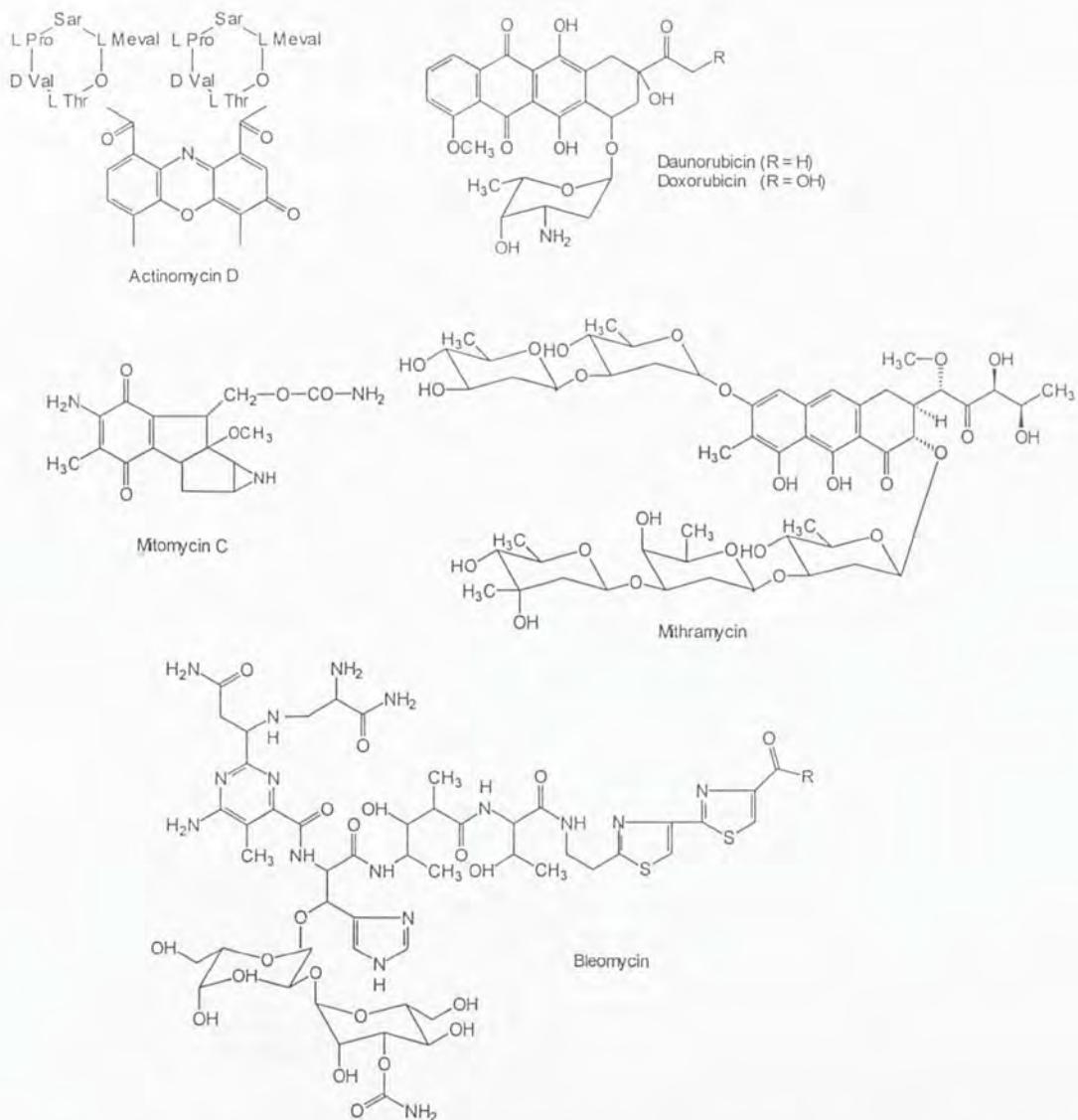


Figure 1.3: Anti-tumor agents produced by actinomycetes with clinical application in cancer chemotherapy

Many of the earlier small molecule anti-tumor agents (Figure 1.3) are synthesized by actinomycetes and these compounds exert cytotoxic action by interfering with DNA function. Actinomycin D and anthracyclines (daunorubicin, doxorubicin) intercalate with DNA, mitomycin C is a crosslinking agent, and bleomycin causes DNA strand breakage. The group of compounds that interacts DNA in a non-intercalative way consists of mithramycin, chromomycin and olivomycin (Salas & Méndez, 1998).

## REFERENCES

- Abraham, R. T. 1998. Mammalian target of rapamycin: immunosuppressive drugs uncover a novel pathway of cytokine receptor signaling. *Current Opinion in Immunology* **10**: 330-336.
- Aikio, S., Vare, H. & Strommer, R. 2000. Soil microbial activity and biomass in the primary succession of a dry heath forest. *Soil Biology & Biochemistry* **32**: 1091-1100.
- Alessi, D. R., Caudwell, F. B., Andjelkovic, M., Hernmings, B. A. & Cohen, P. 1996. Molecular basis for the substrate specificity of protein kinase B; comparison with MAPKAP kinase-1 and p70 S6 kinase. *FEBS Letters* **399**: 333-338.
- Ali, A., Hoeflich, K. P. & Woodgett, J. R. 2000. Glycogen synthase kinase-3: Properties, functions, and regulation. *Chemical Reviews* **101**: 2527 - 2540.
- Altomare, D. A. & Testa, J. R. 2005. Pertubations of AKT signaling pathway in human cancer. *Oncogene* **24**: 7455-7464.
- Anderson, C. & Tatchell, K. 2001. Hyperactive glycogen synthase mutants of *Saccharomyces cerevisiae* suppress the glc7-1 protein phosphatase mutant. *Journal of Bacteriology* **183**: 821-829.
- Andoh, T., Hirata, Y. & Kikuchi, A. 2000. Yeast Glycogen Synthase Kinase 3 Is Involved in Protein Degradation in Cooperation with Bul1, Bul2, and Rsp5. *Molecular and Cellular Biology* **20**: 6712 - 6720.
- Andrews, P. D. & Stark, M. J. R. 2000. Type 1 protein phosphatase is required for maintenance of cell wall integrity, morphogenesis and cell cycle progression in *Saccharomyces cerevisiae*. *Journal of Cell Science* **113**: 507-520.
- Ang, K. L., Shi, D. L., Keong, W. W. & Epstein, R. J. 2005. Upregulated Akt signaling adjacent to gastric cancers: implications for screening and chemoprevention. *Cancer Letters* **225**: 53-59.
- Asai, A., Tsujita, T., Sharma, S. V., Yamashita, Y., Akinaga, S., Funakoshi, M., Kobayashi, H. & Mizukamia, T. 2004. A new structural class of proteasome inhibitors identified by microbial screening using yeast-based assay. *Biochemical Pharmacology* **67**: 227-234.
- Bach, S., Talarek, N., Andrieu, T., Virefond, J.-M., Mettey, Y., Galons, H., Dormont, D., Meijer, L., Cullin, C. & Blondel, M. 2003. Isolation of drugs active against mammalian prions using a yeast-based screening assay. *Nature Biotechnology* **21**: 1075 - 1081.
- Bain, J., McLauchlan, H., Elliot, M. & Cohen, P. 2003. The specificities of protein kinase inhibitors : an update. *Biochemical Journal* **371**: 199 - 204.
- Barbieri, M., Bonafè, M., Franceschi, C. & Paolisso, G. 2003. Insulin/IGF-I-signaling pathway: an evolutionarily conserved mechanism of longevity from yeast to humans. *American Journal of Physiology - Endocrinology and Metabolism* **285**: E1064-E1071.
- Bennecib, M., Gong, C.-X., Grundke-Iqbali, I. & Iqbal, K. 2000. Role of protein phosphatase-2A and -1 in the regulation of GSK-3, cdk5 and cdc2 and the phosphorylation of tau in rat forebrain. *FEBS Letters* **485**: 87-93.

- Bitterman, K. J., Medvedik, O. & Sinclair, D. A. 2003. Longevity regulation in *Saccharomyces cerevisiae*: Linking metabolism, genome stability, and heterochromatin. *Microbiology and Molecular Biology Reviews* **67**: 376–399.
- Bondar, V. M., Sweeney-Gotsch, B., Andreeff, M., Mills, G. B. & McConkey, D. J. 2002. Inhibition of the phosphatidylinositol 3'-kinase-AKT pathway induces apoptosis in pancreatic carcinoma cells *in vitro* and *in vivo*. *Molecular Cancer Therapeutics* **1**: 989–997.
- Bro, C., Regenberg, B., Lagniel, G., Labarre, J., Montero-Lomeli, M. & Nielsen, J. 2003. Transcriptional, proteomic, and metabolic responses to lithium in galactose-grown yeast cells. *Journal of Biological Chemistry* **278**: 32141 - 32149.
- Brown, J. D. & Moon, R. T. 1998. Wnt signaling: why is everything so negative? *Current Opinion in Cell Biology* **10**: 182-187.
- Broxterman, H. J. & Georgopapadakou, N. H. 2005. Anticancer therapeutics: "Addictive" targets, multi-targeted drugs, new drug combinations. *Drug Resistance Updates* **8**: 183 - 197.
- Cade, J. 1949. Lithium salts in the treatment of psychotic excitement. *Medical Journal of Australia* **2**: 349–352.
- Cantrell, D. A. 2001. Phosphoinositide 3-kinase signalling pathways. *Journal of Cell Science* **114**: 1439-1445.
- Cardenas, M. E., Cruz, C., Poeta, M. D., Chung, N., Perfect, J. R. & Heitman, J. 1999. Antifungal activities of antineoplastic agents: *Saccharomyces cerevisiae* as a model system to study drug action. *Clinical Microbiology Reviews* **12**: 583–611.
- Casamayor, A., Torrance, P. D., Kobayashi, T., Thorner, J. & Alessi, D. R. 1999. Functional counterparts of mammalian protein kinases PDK1 and SGK in budding yeast. *Current Biology* **9**: 186 - 197.
- Castillo, S. S., Brognard, J., Petukhov, P. A., Zhang, C., Tsurutani, J., Granville, C. A., Li, M., Jung, M., West, K. A., Gills, J. G., Kozikowski, A. P. & Dennis, P. A. 2004. Preferential inhibition of Akt and killing of Akt-dependent cancer cells by rationally designed phosphatidylinositol ether lipid analogues. *Cancer Research* **64**: 2782-2792.
- Cheah, H.-Y. 2003. *Isolation of actinomycetes from mangroves in Sabah and screening for inhibitors against eukaryotic signal transduction*. Master Thesis. School of Science and Technology, Universiti Malaysia Sabah.
- Chen, G., Bower, K. A., Ma, C., Fang, S., Thiele, C. J. & Luo, J. 2004. Glycogen synthase kinase 3 $\beta$  (GSK3 $\beta$ ) mediates 6-hydroxydopamine-induced neuronal death. *FASEB Journal* **18**: 1162-1164.
- Chen, R.-H., Ding, W. V. & McCormick, F. 2000. Wnt Signaling to b-Catenin Involves Two Interactive Components: Glycogen synthase kinase-3 $\beta$  inhibition and activation of protein kinase C. *Journal of Biological Chemistry* **275**: 17894 - 17899.
- Chen, W. S., Xu, P.-Z., Gottlob, K., Chen, M.-L., Sokol, K., Shiyanova, T., Roninson, I., Weng, W., Suzuki, R., Tobe, K., Kadokawa, T. & Hay, N. 2001. Growth retardation and increased apoptosis in mice with homozygous disruption of the *akt1* gene. *Genes & Development* **15**: 2203 - 2208.

- Cheng, J. Q., Lindsley, C. W., Cheng, G. Z., Yang, H. & Nicosia, S. V. 2005. The Akt/PKB pathway: molecular target for cancer drug discovery. *Oncogene* **24**: 7482 - 7492.
- Chin, Y.-W., Balunas, M. J., Chai, H. B. & Kinghorn, A. D. 2006. Drug discovery from natural sources. *The AAPS Journal* **8**: E239-E253.
- Cho, H., Thorvaldsen, J. L., Chu, Q., Feng, F. & Birnbaum, M. J. 2001. Akt1/PKB $\alpha$  is required for normal growth but dispensable for maintenance of glucose homeostasis in mice. *Journal of Biological Chemistry* **276**: 38349 - 38352.
- Chun, K.-H., Li, J. W. K., Sun, S., Pezzuto, J. M., Lotan, R., Hong, W. K. & Lee, H.-Y. 2003. Effects of deguelin on the Phosphatidylinositol 3-Kinase/Akt pathway and apoptosis in premalignant human bronchial epithelial cells. *J Natl Cancer Inst* **95**: 291-302.
- Cline, G. W., Johnson, K., Regitnig, W., Perret, P., Tozzo, E., Xiao, L., Damico, C. & Shulman, G. I. 2002. Effects of a novel glycogen synthase kinase-3 inhibitor on insulin-stimulated glucose metabolism in Zucker Diabetic Fatty (fa/fa) rats. *Diabetes* **51**: 2903 - 2910.
- Clodfeller-Miller, B., Sarno, P. D., Zmijewska, A. A., Song, L. & Jope, R. S. 2005. Physiological and pathological changes in glucose regulate brain Akt and glycogen synthase kinase-3. *Journal of Biological Chemistry* **280**: 39723-39731.
- Coffer, P. J., Jin, J. & Woodgett, J. R. 1998. Protein kinase B (c-Akt) : a multifunctional mediator of phosphatidylinositol 3-kinase activation. *Biochemical J* **335**: 1 - 13.
- Coghlan, M. P., Culbert, A. A., Cross, D. A., Corcoran, S. L., Yates, J. W., Pearce, N. J., Rausch, O. L., Murphy, G. J., Carter, P. S. & Roxbee Cox, L. 2000. Selective small molecule inhibitors of glycogen synthase kinase-3 modulate glycogen metabolism and gene transcription. *Chemistry & Biology* **7**: 793-803.
- Cohen, P. 2001. The role of protein phosphorylation in human health and disease. *European Journal of Biochemistry* **268**: 5001 - 5010.
- Cohen, P. & Frame, S. 2001. The renaissance of GSK3. *Nature Reviews: Molecular Cell Biology* **2**: 769 - 776.
- Cohen, P., Alessi, D. R. & Cross, D. A. E. 1997. PDK1, one of the missing links in insulin signal transduction? *FEBS Letters* **410**: 3-10.
- Cottier, V., Barberis, A. & Luthi, U. 2006. Novel yeast cell-based assay to screen for inhibitors of human cytomegalovirus protease in a high-throughput format. *Antimicrob. Agents Chemother.* **50**: 565-571.
- Crauwels, M., Donaton, M. C. V., Pernambuco, M. B., Winderickx, J., Winde, J. H. D. & Thevelein, J. M. 1997. The Sch9 protein kinase in the yeast *Saccharomyces cerevisiae* controls cAPK activity and is required for nitrogen activation of the fermentable-growth-medium-induced (FGM) pathway. *Microbiology* **143**: 2627-2637.
- Cross, D. A. E., Culbert, A. A., Chalmers, K. A., Facci, L., Skaper, S. D. & Reith, A. D. 2001. Selective small-molecule inhibitors of glycogen synthase kinase-3 activity protect primary neurones from death. *Journal of Neurochemistry* **77**: 94-102.

- Crowder, R. J. & Freeman, R. S. 2000. Glycogen Synthase Kinase-3 $\beta$  Activity is critical for neuronal death caused by inhibiting phosphatidylinositol 3-kinase or Akt but not for death caused by nerve growth factor withdrawal. *Journal of Biological Chemistry* 275: 34266 - 34271.
- Dajani, R., Fraser, E., Roe, S. M., Young, N., Good, V., Dale, T. C. & Pearl, L. H. 2001. Crystal structure of glycogen synthase kinase 3 $\beta$ : Structural Basis for Phosphate-Primed Substrate Specificity and Autoinhibition. *Cell* 105: 721 - 732.
- Damiens, E., Baratte, B., Marie, D., Eisenbrand, G. & Meijer, L. 2001. Anti-mitotic properties of indirubin-3'-monoxime, a CDK/GSK-3 inhibitor: induction of endoreplication following prophase arrest. *Oncogene* 20: 3786 - 3797.
- Dasmahapatra, G. P., Didolkar, P., Alley, M. C., Ghosh, S., Sausville, E. A. & Roy, K. K. 2004. *In vitro* combination treatment with perifosine and UCN-01 demonstrates synergism against prostate (PC-3) and lung (A549) epithelial adenocarcinoma cell lines. *Clinical Cancer Research* 10: 5242-5252.
- Datta, S. R., Brunet, A. & Greenberg, M. E. 1999. Cellular survival: a play in three Akts. *Genes & Development* 13: 2905 - 2927.
- Datta, S. R., Katsov, A., Hu, L., Petros, A., Fesik, S. W., Yaffe, M. B. & Greenberg, M. E. 2000. 14-3-3 proteins and survival kinases cooperate to inactivate BAD by BH3 domain phosphorylation. *Molecular Cell* 6: 41-51.
- Davies, S. P., Reddy, H., Caivano, M. & Cohen, P. 2000. Specificity and mechanism of action of some commonly used protein kinase inhibitors. *Biochemical Journal* 351: 95 - 105.
- Demain, A. L. 1999. Pharmaceutically active secondary metabolites of microorganisms. *Applied Microbiology and Biotechnology* 52: 455-463.
- Demetri, G. D. 2002. Identification and treatment of chemoresistant inoperable or metastatic GIST: experience with the selective tyrosine kinase inhibitor imatinib mesylate (ST1571). *European Journal of Cancer* 38: S52-S59.
- Ding, H., Zhang, C., Wu, X., Yang, C., Zhang, X., Ding, J. & Xie, Y. 2005. Novel indole a-methylene- $\gamma$ -lactones as potent inhibitors for AKT-mTOR signaling pathway kinases. *Bioorganic & Medicinal Chemistry Letters* 15: 4799-4802.
- Ding, V. W., Chen, R.-H. & McCormick, F. 2000. Differential regulation of glycogen synthase kinase 3 $\beta$  by Insulin and Wnt Signaling. *Journal of Biological Chemistry* 275:
- Doble, B. W. & Woodgett, J. R. 2003. GSK-3: tricks of the trade for a multi-tasking kinase. *Journal of Cell Science* 116: 1175-1186.
- Dominguez, I. & Green, J. B. A. 2001. Missing Links in GSK3 Regulation. *Developmental Biology* 235: 303 - 313.
- Donadio, S., Monciardini, P., Alduina, R., Mazza, P., Chiocchini, C., Cavaletti, L., Sosio, M. & Puglia, A. M. 2002. Microbial technologies for the discovery of novel bioactive metabolites. *Journal of Biotechnology* 99: 187-198.



- Eldar-Finkelman, H., Seger, R., Vandenheede, J. R. & Krebs, E. G. 1995. Inactivation of glycogen synthase kinase-3 by epidermal growth factor is mediated by mitogen-activated protein kinase/p90 ribosomal protein S6 kinase signaling pathway in NIH3T3 cells. *Journal of Biological Chemistry* **270**: 987-990.
- Elsayed, Y. A. & Sausville, E. A. 2001. Selected novel anticancer treatments targeting cell signaling proteins. *The Oncologist* **6**: 517-537.
- Fang, X., Yu, S. X., Lu, Y., Jr, R. C. B., Woodgett, J. R. & Mills, G. B. 2000. Phosphorylation and inactivation of glycogen synthase kinase 3 by protein kinase A. *Proc. Natl. Acad. Sci.* **97**: 11960-11965.
- Fang, X., Yu, S., Tanyi, J. L., Lu, Y., Woodgett, J. R. & Mills, G. B. 2002. Convergence of multiple signaling cascades at glycogen synthase kinase 3: Edg receptor-mediated phosphorylation and inactivation by lysophosphatidic acid through a protein kinase C-dependent intracellular pathway. *Molecular and Cellular Biology* **22**: 2099 - 2110.
- Farkas, I., Hardy, T. A., Depaoli-Roach, A. A. & Roach, P. J. 1990. Isolation of the GSY1 gene encoding yeast glycogen synthase and evidence for the existence of a second gene. *Journal of Biological Chemistry* **265**: 20879-20686.
- Farkas, I., Hardy, T. A., Goebel, M. G. & Roach, P. J. 1991. Two glycogen synthase isoforms in *Saccharomyces cerevisiae* are coded by distinct genes that are differentially controlled. *Journal of Biological Chemistry* **266**: 15602-15607.
- Feng, J., Park, J., Cron, P., Hess, D. & Hemmings, B. A. 2004. Identification of a PKB/Akt Hydrophobic Motif Ser-473 Kinase as DNA-dependent Protein Kinase. *Journal of Biological Chemistry* **279**: 41189-41196.
- Feng, Z., Wilson, S. E., Peng, Z.-Y., Schlender, K. K., Reimann, E. M. & Trumbly, R. J. 1991. The yeast GLC7 gene required for glycogen accumulation encodes a Type 1 Protein Phosphatase. *Journal of Biological Chemistry* **266**: 23796-23801.
- Finlay, D., Patel, S., Dickson, L. M., Shpiro, N., Marquez, R., Rhodes, C. J. & Sutherland, C. 2004. Glycogen Synthase Kinase-3 regulates IGFBP-1 gene transcription through the Thymine-rich Insulin Response Element. *BMC Molecular Biology* **5**: 15-27.
- Frame, S. & Cohen, P. 2001. GSK3 takes centre stage more than 20 years after its discovery. *Biochemical Journal* **359**: 1 - 16.
- Geyskens, I., Kumara, S., Donaton, M., Bergsma, J., Thevelein, J. & Wera, S. 2000. Expression of mammalian PKB complements deletion of the yeast protein kinase Sch9. *NATO ASI Ser. A, Life Sci.* **A316**: 117-126.
- Gomes, K. N., Freitas, S. M. A. C., Pais, T. M., Fietto, J. L. R., Totola, A. H., Arantes, R. M. E., Martins, A., Lucas, C., Schuller, D., Casal, M., Castro, I. M., Fietto, L. G. & Brandao, R. L. 2005. Deficiency of Pkc1 activity affects glycerol metabolism in *Saccharomyces cerevisiae*. *Yeast Research* **5**: 767-776.
- Gradl, D., Kuhl, M. & Wedlich, D. 1999. Keeping a close eye on Wnt-1/wg signaling in *Xenopus*. *Mechanisms of Development* **86**: 3-15.
- Griffioen, G., Swinnen, S. & Thevelein, J. M. 2003. Feedback inhibition on cell wall integrity signaling by Zds1 involves Gsk3 phosphorylation of a cAMP-dependent protein kinase regulatory subunit. *Journal of Biological Chemistry* **278**: 23460-23471.

- Grimes, C. A. & Jope, R. S. 2001. The multifaceted roles of glycogen synthase kinase 3b in cellular signaling. *Progress in Neurobiology* 65: 391-426.
- Gurvich, N. & Klein, P. S. 2002. Lithium and valproic acid: parallels and contrasts in diverse signaling contexts. *Pharmacology & Therapeutics* 96: 45-66.
- Gustin, M. C., Albertyn, J., Alexander, M. & Davenport, K. 1998. MAP Kinase Pathways in the Yeast *Saccharomyces cerevisiae*. *Microbiology and Molecular Biology Reviews* 62: 1264-1300.
- Hajji, K., Clotet, J. & Ariño, J. 1999. Disruption and phenotypic analysis of seven ORFs from the left arm of chromosome XV of *Saccharomyces cerevisiae*. *Yeast* 15: 435-441.
- Hamelers, I. H. L., Schaik, R. F. M. A. V., Sipkema, J., Sussenbach, J. S. & Steenbergh, P. H. 2002. Insulin-like growth factor I triggers nuclear accumulation of cyclin D1 in MCF-7S breast cancer cells. *Journal of Biological Chemistry* 277: 47645-47652.
- Hanada, M., Feng, J. & Hemmings, B. A. 2004. Structure, regulation and function of PKB/AKT--a major therapeutic target. *Biochimica et Biophysica Acta (BBA) - Proteins & Proteomics* 1697: 3-16.
- Hanks, S. K. & Hunter, T. 1995. The eukaryotic protein kinase superfamily: kinase (catalytic) domain structure and classification. *FASEB Journal* 9: 576-596.
- Hardy, T. A., Wu, D. & Roach, P. J. 1995. Novel *Saccharomyces cerevisiae* Gene, MRK1, Encoding a Putative Protein Kinase with Similarity to Mammalian Glycogen Synthase Kinase-3 and *Drosophila* Zeste-White3/Shaggy. *Biochemical and Biophysical Research Communications* 208: 728-734.
- Hartley, A. D., Ward, M. P. & Garrett, S. 1994. The YAK1 protein kinase of *Saccharomyces cerevisiae* moderates thermotolerance and inhibits growth by an Sch9 protein kinase-independent mechanism. *Genetics* 136: 465-474.
- Hayakawa, M. & Nonomura, H. 1987. Humic acid-vitamin agar, a new medium for the selective isolation of soil actinomycetes. *Journal of Fermentation Technology* 65: 501-509.
- Hayakawa, M. & Nonomura, H. 1993. *Selective methods for soil actinomycetes*. Tokyo: Japanese Association of Actinomycetes.
- Hayakawa, M., Iino, H., Takeuchi, S. & Yamazaki, T. 1997. Application of a method incorporating treatment with chloramine-T for the selective isolation of Streptosporangiaceae from soil. *Journal of Fermentation and Bioengineering* 84: 599-602.
- Heng, H. L., Tan, I., Tan, J. S., Ho, W. L., Lee, S. H., Tee, L. K., Gan, S. P., Chan, S. Y. & Ho, C. C. 2003. Searching for inhibitors against MAPK kinase (MKK1), protein phosphatase (MSG5 and GLC7) and isocitrate lyase (ICL) from actinomycetes and filamentous fungi isolated from lower Kinabatangan. In *Lower Kinabatangan scientific expedition 2002*. Mohamed, Takano, Goossens and Indran (Eds.). Kota Kinabalu: Universiti Malaysia Sabah. 89-119.
- Hennessy, B. T., Smith, D. L., Ram, P. T., Lu, Y. & Mills, G. B. 2005. Exploiting the PI3K/Akt pathway for cancer drug discovery. *Nature Reviews: Drug Discovery* 4: 988-1004.

- Herman, P. K. & Emr, S. D. 1990. Characterization of VPS34, a gene required for vacuolar protein sorting and vacuole segregation in *Saccharomyces cerevisiae*. *Molecular and Cellular Biology* 10: 6742-6754.
- Hers, I., Tavare, J. M. & Denton, R. M. 1999. The protein kinase C inhibitors bisindolylmaleimide I (GF 109203x) and IX (Ro 31-8220) are potent inhibitors of glycogen synthase kinase-3 activity. *FEBS Letters* 460: 433-436.
- Hessler, P. E., Larsen, P. E., Constantinou, A. I., Schram, K. H. & Weber, J. M. 1997. Isolation of isoflavones from soy-based fermentations of the erythromycin-producing bacterium *Saccharopolyspora erythraea*. *Appl. Microbiol. Biotechnol.* 47: 398-404.
- Hill, M. M. & Hemmings, B. A. 2002. Inhibition of protein kinase B/Akt: implications for cancer therapy. *Pharmacology & Therapeutics* 93: 243-251.
- Hill, M. M., Andjelkovic, M., Brazil, D. P., Ferrari, S., Fabbro, D. & Hemmings, B. A. 2001. Insulin-stimulated protein Kinase B phosphorylation on Ser-473 is independent of its activity and occurs through a staurosporine-insensitive kinase. *Journal of Biological Chemistry* 276: 25643-25646.
- Hirata, Y., Andoh, T., Asahara, T. & Kikuchi, A. 2003. Yeast glycogen synthase kinase-3 activates Msn2p-dependent transcription of stress responsive genes. *Molecular and Cellular Biology* 14: 302 - 312.
- Ho, C. C. 2003. *Professorial lecture series: Molecular cell biology, biodiversity and biotechnology*. Kota Kinabalu: Universiti Malaysia Sabah.
- Ho, C. C., Tan, G. Y. A., Seow, I., Ajam, N., Tan, E. L., Goodfellow, M., Ward, A. C., Brown, R., Wong, N. K., Lo, C. W., Cheah, H. Y., Lai, N. S. & Suzuki, K. I. 2001. Isolation, characterisation and biological activities of actinomycetes isolated from dipterocarp rain forest soils in Malaysia. In *Microbial diversity in Asia: Technology and Prospects*. Nga, Tan and Suzuki (Eds.). Singapore: World Scientific. 209-228.
- Hoessl, R., Leclerc, S., Endicott, J. A., Nobel, M. E. M., Lawrie, A., Tunnah, P., Leost, M., Damiens, E., Marie, D., Marko, D., Niederberger, E., Tang, W., Eisenbrand, G. & Meijer, L. 1999. Indirubin, the active constituent of a Chinese antileukaemia medicine, inhibits cyclin-dependent kinases. *Nature Cell Biology* 1: 60 - 67.
- Hong, M., Chen, D. C. R., Klein, P. S. & Lee, V. M.-Y. 1997. Lithium reduces tau phosphorylation by inhibition of Glycogen Synthase Kinase-3. *Journal of Biological Chemistry* 40: 25326 - 25332.
- Inagaki, M., Schmelzle, T., Yamaguchi, K., Irie, K., Hall, M. N. & Matsumoto, K. 1999. PDK1 Homologs Activate the Pkc1-Mitogen-Activated Protein Kinase Pathway in Yeast. *Molecular and Cellular Biology* 19: 8344-8352.
- Janmaat, M. L., Kruyt, F. A. E., Rodriguez, J. A. & Giaccone, G. 2003. Response to epidermal growth factor receptor inhibitors in non-small cell lung cancer cells: Limited antiproliferative effects and absence of apoptosis associated with persistent activity of extracellular signal-regulated kinase or Akt kinase pathways. *Clinical Cancer Research* 9: 2316-2326.
- Jin, X., Gossett, D. R., Wang, S., Yang, D., Cao, Y., Chen, J., Guo, R., Reynolds, R. K. & Lin, J. 2004. Inhibition of AKT survival pathway by a small molecule inhibitor in human endometrial cancer cells. *British Journal of Cancer* 91: 1808 – 1812.

- Johnston, S. R. D. 2005. Clinical trials of intracellular signal transductions inhibitors for breast cancer — a strategy to overcome endocrine resistance. *Endocrine-Related Cancer* **12**: S145–S157.
- Jope, R. S. 2003. Lithium and GSK-3: one inhibitor, two inhibitory actions, multiple outcomes. *Trends in Pharmacological Sciences* **24**: 441-443.
- Kanfer, I., Skinner, M. F. & Walker, R. B. 1998. Analysis of macrolide antibiotics. *Journal of Chromatography A* **812**: 255–286.
- Kawauchi, K., Ogasawara, T., Yasuyama, M. & Ohkawa, S.-I. 2003. Involvement of Akt kinase in the action of ST1571 on chronic myelogenous leukemia cells. *Blood Cells, Molecules, and Diseases* **31**: 11-17.
- Ki, S. W., Kasahara, K., Kwon, H. J. & Eishima, J. 1998. Identification of radicicol as an *in vivo* ras/raf interaction with the yeast two-hybrid screening system. *Journal of Antibiotics* **51**: 936-944.
- Kibat, C., Lai, N. S., Puah, S. H., Hew, C. S. & Voo, C. L. Y. 2005. Screening for bioactive compounds from microorganisms isolated from Trus Madi mountain, Sabah. *Journal of Tropical Biology and Conservation* **1**: 9-19.
- Kim, D., Kim, S., Koh, H., Yoon, S.-O., Chung, A.-S., Cho, K. S. & Chung, J. 2001. Akt/PKB promotes cancer cell invasion via increased motility and metalloproteinase production. *FASEB Journal* **15**: 1953 - 1962.
- Klein, P. S. & Melton, D. A. 1996. A molecular mechanism for the effect of lithium on development. *Proc. Natl. Acad. Sci.* **93**: 8455 - 8459.
- Kolch, W. 2000. Meaningful relationships : the regulation of the Ras/Raf/MEK/ERK pathway by protein interactions. *Biochemical Journal* **351**: 289-305.
- Kondapaka, S. B., Singh, S. S., Dasmahapatra, G. P., Sausville, E. A. & Roy, K. K. 2003. Perifosine, a novel alkylphospholipid, inhibits protein kinase B activation. *Molecular Cancer Therapeutics* **2**: 1093–1103.
- Kumar, C. C. & Madison, V. 2005. Akt crystal structure and Akt-specific inhibitors. *Oncogene* **24**: 7493-7501.
- Kunick, C., Lauenroth, K., Leost, M., Meijer, L. & Lemcke, T. 2004. 1-Azakenpaullone is a selective inhibitor of glycogen synthase kinase-3b. *Bioorganic & Medicinal Chemistry Letters* **14**: 413 - 416.
- Lai, N. S. 2003. *Isolation of actinomycetes from Sabah and the screening of inhibitors against eukaryotic signal transduction*. Masters Thesis. School of Science and Technology, Universiti Malaysia Sabah.
- Lechevalier, H. A. & Lechevalier, M. P. 1981. Introduction to the Order Actinomycetales. In *The Prokaryotes: A Handbook on Habitats, Isolation and Identification of Bacteria*. Starr, Stolp, Truper, Balows and Schlegel (Eds.). Berlin Heidelberg: Springer-Verlag. 1915-1922.

- Leclerc, S., Garnier, M., Hoessel, R., Marko, D., Bibb, J. A., Snyder, G. L., Greengard, P., Biernati, J., Wui, Y.-Z., Mandelkow, E.-M., Eisenbrand, G. & Meijer, L. 2001. Indirubins inhibit glycogen synthase kinase-3 $\beta$  and CDK5/p25, two protein kinases involved in abnormal tau phosphorylation in Alzheimer's disease: A property common to most cyclin-dependent kinase inhibitors? *Journal of Biological Chemistry* **276**: 251 - 260.
- Lee, H.-Y. 2004. Molecular mechanisms of deguelin-induced apoptosis in transformed human bronchial epithelial cells. *Biochemical Pharmacology* **68**: 1119-1124.
- Leost, M., Schultz, C., Link, A., Wu, Y.-Z., Biernat, J., Mandelkow, E.-M., Bibb, J. A., Snyder, G. L., Greengard, P., Zaharevitz, D. W., Gussio, R., Senderowicz, A. M., Sausville, E. A., Kunick, C. & Meijer, L. 2000. Paullones are potent inhibitors of glycogen synthase kinase-3 $\beta$  and cyclin-dependent kinase 5/p25. *European Journal of Biochemistry* **267**: 5983 - 5994.
- Li, Q., Li, T., Zhu, G.-D., Gong, J., Claibone, A., Dalton, C., Luo, Y., Johnson, E. F., Shi, Y., Liu, X., Klinghofer, V., Bauch, J. L., Marsh, K. C., Bouska, J. J., Arries, S., Jong, R. D., Oltersdorf, T., Stoll, V. S., Jakob, C. G., Rosenberg, S. H. & Giranda, V. L. 2006a. Discovery of trans-3,4'-bispyridinylethylenes as potent and novel inhibitors of protein kinase B (PKB/Akt) for the treatment of cancer: Synthesis and biological evaluation. *Bioorganic & Medicinal Chemistry Letters* **16**: 1679-1685.
- Li, Q., Woods, K. W., Thomas, S., Zhu, G.-D., Packard, G., Fisher, J., Li, T., Gong, J., Dinges, J., Song, X., Abrams, J., Luo, Y., Johnson, E. F., Shi, Y., Liu, X., Klinghofer, V., Jong, R. D., Oltersdorf, T., Stoll, V. S., Jakob, C. G., Rosenberg, S. H. & Giranda, V. L. 2006b. Synthesis and structure-activity relationship of 3,40-bispyridinylethylenes: Discovery of a potent 3-isoquinolinylpyridine inhibitor of protein kinase B (PKB/Akt) for the treatment of cancer. *Bioorganic & Medicinal Chemistry Letters* **16**: 2000-2007.
- Lim, M.-Y., Dailey, D., Martin, G. S. & Thorner, J. 1993. Yeast *MCK1* protein kinase autophosphorylates at tyrosine and serine but phosphorylates exogenous substrates at serine and threonine. *Journal of Biological Chemistry* **268**: 21155-21164.
- Lindsley, C. W., Zhao, Z., Leister, W. H., Robinson, R. G., Barnett, S. F., Defeo-Jones, D., Jones, R. E., Hartman, G. D., Huff, J. R., Huber, H. E. & Duggan, M. E. 2005. Allosteric Akt (PKB) inhibitors: discovery and SAR of isozyme selective inhibitors. *Bioorganic & Medicinal Chemistry Letters* **15**: 761-764.
- Lo, C. W., Lai, N. S., Cheah, H. Y., Wong, N. K. I. & Ho, C. C. 2002. Actinomycetes isolated from soil samples from Crocker Range Sabah. <http://www.arbec.com.my/pdf/art21julysep02.pdf>
- Lobenhofer, E. K., Huper, G., Iglesias, J. D. & Marks, J. R. 2000. Inhibition of mitogen-activated protein kinase and phosphatidylinositol 3-Kinase activity in MCF-7 cells prevents estrogen-induced mitogenesis. *Cell Growth and Differentiation* **11**: 99-110.
- Loewith, R., Jacinto, E., Wullschleger, S., Lorberg, A., Crespo, J. L., Bonenfant, D. B., Oppenheimer, W., Jenoe, P. & Hall, M. N. 2002. Two TOR complexes, only one of which is rapamycin sensitive, have distinct roles in cell growth control. *Molecular Cell* **10**: 457-468.
- Longo, V. D. 2003. The Ras and Sch9 pathways regulate stress resistance and longevity. *Experimental Gerontology* **38**: 807-811.

- Ma, N., Jin, J., Lu, F., Woodgett, J. & Liu, F.-F. 2001. The role of protein kinase B (PKB) in modulating heat sensitivity in a human breast cancer cell line. *International Journal of Radiation Oncology, Biology and Physics* **50**: 1041-1050.
- Mackeigan, J. P., Taxman, D. J., Hunter, D., Earp, H. S., Iii, Graves, L. M. & Ting, J. P.-Y. 2002. Inactivation of the antiapoptotic phosphatidylinositol 3-kinase-Akt pathway by the combined treatment of Taxol and mitogen-activated protein kinase kinase inhibition. *Clinical Cancer Research* **8**: 2091-2099.
- Malathi, K., Xiao, Y. & Mitchell, A. P. 1997. Interaction of yeast repressor-activator protein Ume6p with glycogen synthase kinase 3 homolog Rim11p. *Molecular and Cellular Biology* **17**: 7230-7236.
- Malathi, K., Xiao, Y. & Mitchell, A. P. 1999. Catalytic roles of yeast GSK3 $\beta$ /Shaggy homolog Rim11p in meiotic activation. *Genetics* **153**: 1145-1152.
- Manley, P. W., Cowan-Jacob, S. W., Buchdunger, E., Fabbro, D., Fendrich, G., Furet, P., Meyer, T. & Zimmermann, J. 2002. Imatinib: a selective tyrosine kinase inhibitor. *European Journal of Cancer* **38**: S19-S27.
- Mcmillan, J. N., Longtine, M. S., Sia, R. A. L., Theesfeld, C. L., Bardes, E. S. G., Pringle, J. R. & Lew, D. J. 1999. The morphogenesis checkpoint in *Saccharomyces cerevisiae*: Cell cycle control of Swe1p degradation by Hsl1p and Hsl7p. *Molecular and Cellular Biology* **19**: 6929-6939.
- Meier, R., Alessi, D. R., Cron, P. & Hemmings, B. A. 1997. Mitogenic activation, phosphorylation, and nuclear translocation of protein kinase B $\beta$ . *Journal of Biological Chemistry* **272**: 30491-30497.
- Meijer, L., Flajolet, M. & Greengard, P. 2004. Pharmacological inhibitors of glycogen synthase kinase 3. *Trends in Pharmacological Sciences* **25**: 471-480.
- Meijer, L., Thunnissen, A.-M., White, A., Garnier, M., Nikolic, M., Tsai, L.-H., Walter, J., Cleverley, K., Salinas, P., Wu, Y.-Z., Biernat, J., Mandelkow, E.-M., Kim, S.-H. & Pettit, G. 2000. Inhibition of cyclin-dependent kinases, GSK-3 $\beta$  and CK1 by hymenialdisine, a marine sponge constituent. *Chemistry & Biology* **7**: 51-63.
- Miyadoh, S. 1997. Frontispiece. In *Atlas of Actinomycetes*. Miyadoh, Hamada, Hotta, Kudo, Seino, Vobis and Yokota (Eds.). Japan: Asakura Publishing. 2-8.
- Mizunuma, M., Hirata, D., Miyaoka, R. & Miyakawa, T. 2001. GSK-3 kinase Mck1 and calcineurin coordinately mediate Hsl1 down-regulation by Ca<sup>2+</sup> in budding yeast. *EMBO J.* **20**: 1074 - 1085.
- Morano, K. A. & Thiele, D. J. 1999. The Sch9 protein kinase regulates Hsp90 chaperone complex signal transduction activity *in vivo*. *EMBO J.* **18**: 5953-5962.
- Morovjan, G., Szakacs, G. & Fekete, J. 1997. Monitoring of selected metabolites and biotransformation products from fermentation broths by high-performance liquid chromatography. *Journal of Chromatography A* **763**: 165-172.
- Mukai, F., Ishiguro, K., Sano, Y. & Fujita, S. C. 2002. Alternative splicing isoform of tau protein kinase I/glycogen synthase kinase 3 $\beta$ . *Journal of Neurochemistry* **81**: 1073-1083.

- Murray, N. R., Davidson, L. A., Chapkin, R. S., Gustafson, W. C., Schattenberg, D. G. & Fields, A. P. 1999. Overexpression of protein kinase C  $\beta$ II induces colonic hyperproliferation and increased sensitivity to colon carcinogenesis. *Journal of Cell Biology* **145**: 699–711.
- Naito, Y., Kobayashi, R. & Hidaka, H. 1999. Analysis of signal transduction pathways using protein kinase inhibitors and activators. In *Protein phosphorylation*. Hames (Ed.). New York: Oxford University Press. 33-67.
- Neigeborn, L. & Mitchell, A. 1991. The yeast MCK1 gene encodes a protein kinase homolog that activates early meiotic gene expression. *Genes & Development* **5**: 533-548.
- Nikoulina, S. E., Ciaraldi, T. P., Mudaliar, S., Carter, L., Johnson, K. & Henry, R. R. 2002. Inhibition of glycogen synthase kinase 3 improves insulin action and glucose metabolism in human skeletal muscle. *Diabetes* **51**: 2190-2198.
- Numata, K. & Nimura, S. 2003. Access to soil actinomycetes in Malaysian tropical rain forests. *Actinomycetol.* **17**: 54-56.
- Ong, S. M., Voo, L. Y. C., Lai, N. S., Stark, M. J. R. & Ho, C. C. Screening and characterization of microbial inhibitors against eukaryotic protein phosphatases (PP1 and PP2A). *Journal of Applied Microbiology* – Published article online 30 Aug 2006
- Patel, S., Doble, B. & Woodgett, J. R. 2004. Glycogen synthase kinase-3 in insulin and Wnt signalling: a double-edged sword. *Biochemical Society Transactions* **32**: 803-808.
- Peat, A. J., Garrido, D., Boucheron, J. A., Schweiker, S. L., Dickerson, S. H., Wilson, J. R., Wang, T. Y. & Thomson, S. A. 2004a. Novel GSK-3 inhibitors with improved cellular activity. *Bioorganic & Medicinal Chemistry Letters* **14**: 2127-2130.
- Peat, A. J., Boucheron, J. A., Dickerson, S. H., Garrido, D., Mills, W., Peckham, J., Preugschat, F., Smalley, T., Schweiker, S. L. & Wilson, J. R. 2004b. Novel pyrazolopyrimidine derivatives as GSK-3 inhibitors. *Bioorganic & Medicinal Chemistry Letters* **14**: 2121-2125.
- Pederson, B. A., Cheng, C., Wilson, W. A. & Roach, P. J. 2000. Regulation of glycogen synthase: Identification of residues involved in regulation by the allosteric ligand glucose-6-p and by phosphorylation. *Journal of Biological Chemistry* **275**: 27753–27761.
- Polakis, P. 2000. Wnt signaling and cancer. *Genes & Development* **14**: 1837 - 1851.
- Puziss, J. W., Hardy, T. A., Johnson, R. B., Roach, P. J. & Hieter, P. 1994. MDS1, a Dosage Suppressor of an mck1 Mutant, Encodes a Putative Yeast Homolog of Glycogen Synthase Kinase 3. *Molecular and Cellular Biology* **14**: 831-839.
- Ramaswamy, N. T., Li, L., Khalil, M. & Cannon, J. F. 1998. Regulation of yeast glycogen metabolism and sporulation by Glc7p protein phosphatase. *Genetics* **149**: 57-72.
- Rayner, T. F., Gray, J. V. & Thorner, J. W. 2002. Direct and novel regulation of cAMP-dependent protein kinase by Mck1p, a yeast glycogen synthase kinase-3. *Journal of Biological Chemistry* **277**: 16814–16822.

- Ring, D. B., Johnson, K. W., Henriksen, E. J., Nuss, J. M., Goff, D., Kinnick, T. R., Ma, S. T., Reeder, J. W., Samuels, I., Slabiak, T., Wagman, A. S., Hammond, M.-E. W. & Harrison, S. D. 2003. Selective Glycogen Synthase Kinase 3 inhibitors potentiates insulin activation of glucose transport and utilization *in vitro* and *in vivo*. *Diabetes* **52**: 588 - 595.
- Rode, M. W. 1999. Influence of forest growth on former heathland on nutrient input and its consequences for nutrition and management of heath and forest. *Forest Ecology and Management* **114**: 31-43.
- Roelants, F. M., Torrance, P. D. & Thorner, J. 2004. Differential roles of PDK1- and PDK2-phosphorylation sites in the yeast AGC kinases Ypk1, Pkc1 and Sch9. *Microbiology* **150**: 3289–3304.
- Roelants, F. M., Torrance, P. D., Bezman, N. & Thorner, J. 2002. Pkh1 and Pkh2 differentially phosphorylate and activate Ypk1 and Ykr2 and define protein kinase modules required for maintenance of cell wall integrity. *Molecular Biology of the Cell* **13**: 3005–3028.
- Row, P. E., Reaves, B. J., Domin, J., Luzio, J. P. & Davidson, H. W. 2001. Overexpression of a rat kinase-deficient phosphoinositide 3-kinase, Vps34p, inhibits cathepsin D maturation. *Biochemical Journal*. **353**: 655-661.
- Salas, J. A. & Méndez, C. 1998. Genetic manipulation of antitumor-agent biosynthesis to produce novel drugs. *TIBTECH* **16**: 475-482.
- Schenk, P. W. & Snaar-Jagalska, B. E. 1999a. Signal perception and transduction: the role of protein kinases. *Biochimica et Biophysica Acta* **1449**: 1 - 24.
- Schenk, P. W. & Snaar-Jagalska, B. E. 1999b. Signal perception and transduction: the role of protein kinases. *Biochimica et Biophysica Acta (BBA) - Molecular Cell Research* **1449**: 1-24.
- Schmelzle, T. & Hall, M. N. 2000. TOR, a central controller of cell growth. *Cell* **103**: 253–262.
- Shankar, S. L., Krupski, M., Parashar, B., Okwuaka, C., O'Gui, K., Mani, S. & Shafit-Zagardo, B. 2004. UCN-01 alters phosphorylation of Akt and GSK3 $\beta$  and induces apoptosis in six independent human neuroblastoma cell lines. *Journal of Neurochemistry* **90**: 702 - 711.
- Sharma, V. & Tepe, J. J. 2004. Potent inhibition of checkpoint kinase activity by a hymenialdisine-derived indoloazepine. *Bioorganic & Medicinal Chemistry Letters* **14**: 4319 - 4321.
- Shaw, R. J. & Cantley, L. C. 2006. Ras, PI(3)K and mTOR signaling controls tumour cell growth. *Nature* **441**: 424-430.
- Shen, L., Proulx, C., Conway, B. R., Westover, L., Xu, J. Z., Look, R. A., Chen, X., Beavers, M. P., Roberts, J., Murray, W. V., Demarest, K. T. & Kuo, G.-H. 2004. Synthesis and biological evaluation of novel macrocyclic bis-7-azaindolylmaleimides as potent and highly selective glycogen synthase kinase-3 $\beta$  (GSK-3 $\beta$ ) inhibitors. *Bioorganic & Medicinal Chemistry* **12**: 1239 - 1255.

- Shero, J. & Hieter, P. 1991. A suppressor of a centromere DNA mutation encodes a putative protein kinase (MCK1). *Genes & Development* **5**: 549-560.
- Shimizu, T., Okayama, A., Inoue, T. & Takeda, K. 2005. Analysis of gene expression during staurosporine-induced neuronal differentiation of human prostate cancer cells. *Oncology Reports* **14**: 441-448.
- Shirling, E. B. & Gottlieb, D. 1966. Methods for characterization of *Streptomyces* species. *International Journal Systematic Bacteriology* **16**: 313-340.
- Silakowski, B., Kunze, B. & Mullera, R. 2001. Multiple hybrid polyketide synthase/non-ribosomal peptide synthetase gene clusters in the myxobacterium *Stigmatella aurantiaca*. *Gene* **275**: 233-240.
- Skoko, N., Vujovi, J., Savic, M., Papic, N., Vasiljevic, B. & Ljubijankic, G. 2005. Construction of *Saccharomyces cerevisiae* strain FAV20 useful in detection of immunosuppressants produced by soil actinomycetes. *Journal of Microbiological Methods* **61**: 137-140.
- Smith, D. G., Buffet, M., Fenwick, A. E., Haigh, D., Ife, R. J., Saunders, M., Slingsby, B. P., Stacey, R. & Ward, R. W. 2001. 3-Anilino-4-arylmaleimides: potent and selective inhibitors of glycogen synthase kinase-3 (GSK-3). *Bioorganic & Medicinal Chemistry Letters* **11**: 635-639.
- Stack, J. H. & Emr, S. D. 1994. Vps34p required for yeast vacuolar protein sorting is a multiple specificity kinase that exhibits both protein kinase and phosphatidylinositol-specific PI 3-Kinase activities. *Journal of Biological Chemistry* **269**: 31552-31562.
- Stadler, M., Henkel, T., Muller, H., Weber, K. & Schlecker, H. 1998. Identification of alkaloids and polyketides in an Actinomycete by high-performance liquid chromatography with mass spectrometric and UV-Visible detection. *Journal of Chromatography A* **818**: 187-195.
- Strobel, G. & Daisy, B. 2003. Bioprospecting for microbial endophytes and their natural products. *Microbiology and Molecular Biology Reviews* **67**: 491-502.
- Sugita, K. & Ohtani, M. 1997. Inhibitors of ras-transformation. *Current pharmaceutical design* **3**: 323-334.
- Tanaka, K., Nakafuku, M., Tamanoi, F., Kaziro, Y., Matsumoto, K. & Toh-E, A. 1990. IRA2, a second gene of *Saccharomyces cerevisiae* that encodes a protein with a domain homologous to mammalian ras GTPase-activating protein. *Molecular and Cellular Biology* **10**: 4303-4313.
- Tang, H.-J., Jin, X., Wang, S., Yang, D., Cao, Y., Chen, J., Gossett, D. R. & Lin, J. 2006. A small molecule compound inhibits AKT pathway in ovarian cancer cell lines. *Gynecologic Oncology* **100**: 308-317.
- Teraishi, F., Kagawa, S., Watanabe, T., Tango, Y., Kawashima, T., Umeoka, T., Nisizaki, M., Tanaka, N. & Fujiwara, T. 2005. ZD1839 (Gefitinib, 'Iressa'), an epidermal growth factor receptor-tyrosine kinase inhibitor, enhances the anti-cancer effects of TRAIL in human esophageal squamous cell carcinoma. *FEBS Letters* **579**: 4069-4075.
- Tremont-Lukats, I. W. & Gilbert, M. R. 2003. Advances in molecular therapies in patients with brain tumors. *Cancer Control* **10**: 125-137.

- Ummersen, L. V., Binger, K., Volkman, J., Marnocha, R., Tutsch, K., Kolesar, J., Arzoomanian, R., Alberti, D. & Wilding, G. 2004. A phase I trial of perifosine (NSC 639966) on a loading dose/maintenance dose schedule in patients with advanced cancer. *Clinical Cancer Research* **10**: 7450–7456.
- Unnikrishnan, I., Miller, S., Meinke, M. & Laporte, D. C. 2003. Multiple positive and negative elements involved in the regulation of expression of GSY1 in *Saccharomyces cerevisiae*. *Journal of Biological Chemistry* **278**: 26450–26457.
- Vandromme, M., Rochat, A., Meier, R., Carnac, G., Besser, D., Hemmings, B. A., Fernandez, A. & Lamb, N. J. C. 2001. Protein Kinase B beta /Akt2 Plays a Specific Role in Muscle Differentiation. *Journal of Biological Chemistry* **276**: 8173–8179.
- Vink, S. R., Schellens, J. H. M., Blitterswijk, W. J. V. & Verheij, M. 2005. Tumor and normal tissue pharmacokinetics of perifosine, an oral anti-cancer alkylphospholipid. *Investigational New Drugs* **23**: 279–286.
- Vobis, G. 1997. Morphology of actinomycetes. In *Atlas of Actinomycetes*. Miyadoh, Hamada, Hotta, Kudo, Seino, Vobis and Yokota (Eds.). Japan: Asakura Publishing. 180–191.
- Voo, L. Y., Lai, N. S., Daim, S., Lo, C. W., Chung, Y. T., Moh, M. H. & Ho, C. C. 2003. Screening for inhibitors against eukaryotic signal transduction from actinomycetes and fungi from limestone hills, Tabin and caves, Sapulut. In *Tabin limestone scientific expedition 2000*. Mohamed, Schilthuizen and Andau (Eds.). Kota Kinabalu: Universiti Malaysia Sabah.
- Wang, J., Ito, T., Ueda, N., Okudela, K., Yazawa, T. & Kitamura, H. 2005. PI3K-AKT pathway mediates growth and survival signals during development of fetal mouse lung. *Tissue and Cell* **37**: 25–35.
- Watanabe, Y., Irie, K. & Matsumoto, K. 1995. Yeast RLM1 encodes a serum response factor-like protein that may function downstream of the Mpk1 (Slc2) Mitogen-Activated Protein Kinase pathway. *Molecular and Cellular Biology* **15**: 5740–5749.
- Weinstein-Oppenheimer, C. R., Blalock, W. L., Steelman, L. S., Chang, F. & McCubrey, J. A. 2000. The Raf signal transduction cascade as a target for chemotherapeutic intervention in growth factor-responsive tumors. *Pharmacology & Therapeutics* **88**: 229–279.
- Wera, S., Bergsma, J. C. T. & Thevelein, J. M. 2001. Phosphoinositides in yeast: genetically tractable signalling. *Yeast Research* **1**: 9–13.
- Witherington, J., Bordas, V., Gaiba, A., Naylor, A., Rawlings, A. D., Slingsby, B. P., Smith, D. G., Takle, A. K. & Ward, R. W. 2003a. 6-Heteroaryl-pyrazolo[3,4-b]pyridines: potent and selective inhibitors of glycogen synthase kinase-3 (GSK-3). *Bioorganic & Medicinal Chemistry Letters* **13**: 3059–3062.
- Witherington, J., Bordas, V., Garland, S. L., Hickey, D. M. B., Ife, R. J., Liddle, J., Saunders, M., Smith, D. G. & Ward, R. W. 2003b. 5-Aryl-pyrazolo[3,4-b]pyridines: potent inhibitors of glycogen synthase kinase-3 (GSK-3). *Bioorganic & Medicinal Chemistry Letters* **13**: 1577–1580.

- Witherington, J., Bordas, V., Haigh, D., Hickey, D. M. B., Ife, R. J., Rawlings, A. D., Slingsby, B. P., Smith, D. G. & Ward, R. W. 2003c. 5-Aryl-pyrazolo[3,4-b]pyridazines: potent inhibitors of glycogen synthase kinase-3 (GSK-3). *Bioorganic & Medicinal Chemistry Letters* **13**: 1581-1584.
- Witherington, J., Bordas, V., Gaiba, A., Garton, N. S., Naylor, A., Rawlings, A. D., Slingsby, B. P., Smith, D. G., Takle, A. K. & Ward, R. W. 2003d. 6-Aryl-pyrazolo[3,4-b]pyridines: potent inhibitors of glycogen synthase kinase-3 (GSK-3). *Bioorganic & Medicinal Chemistry Letters* **13**: 3055-3057.
- Yang, L., Dan, H. C., Sun, M., Liu, Q., Sun, X.-M., Feldman, R. I., Hamilton, A. D., Polokoff, M., Nicosia, S. V., Herlyn, M., Sebti, S. M. & Cheng, J. Q. 2004. Akt/Protein Kinase B signaling inhibitor-2, a selective small molecule inhibitor of Akt signaling with antitumor activity in cancer cells overexpressing Akt. *Cancer Research* **64**: 4394-4399.
- Yang, R., Chun, K. T. & Wek, R. C. 1998. Mitochondrial respiratory mutants in yeast Inhibit glycogen accumulation by blocking activation of glycogen synthase. *Journal of Biological Chemistry* **273**: 31337-31344.
- Yang, Z.-Z., Tschopp, O., Hemmings-Miesczak, M., Feng, J., Brodbeck, D., Perentes, E. & Hemmings, B. A. 2003. Protein kinase Ba/Akt1 regulates placental development and fetal growth. *Journal of Biological Chemistry* **278**: 32124-32131.
- Yashiroda, H., Kaida, D., Toh-E, A. & Kikuchi, Y. 1998. The PY-motif of Bul1 protein is essential for growth of *Saccharomyces cerevisiae* under various stress conditions. *Gene* **225**: 39-46.
- Yashiroda, H., Oguchi, T., Yasuda, Y., Toh-E, A. & Kikuchi, Y. 1996. Bul1, a new protein that binds to the Rsp5 ubiquitin ligase in *Saccharomyces cerevisiae*. *Molecular and Cellular Biology* **16**: 3255-3263.
- Yuan, Z., Agarwal-Mawal, A. & Paudel, H. K. 2004. 14-3-3 binds to and mediates phosphorylation of microtubule-associated tau protein by Ser<sup>9</sup>-phosphorylated glycogen synthase kinase 3b in the brain. *Journal of Biological Chemistry* **279**: 26105-26114.
- Zhang, F., Phiel, C. J., Spece, L., Gurvich, N. & Klein, P. S. 2003. Inhibitory phosphorylation of glycogen synthase kinase-3 (GSK-3) in response to lithium: Evidence for autoregulation of GSK-3. *Journal of Biological Chemistry* **278**: 33067 - 33077.
- Zhang, H.-C., Ye, H., Conway, B. R., Derian, C. K., Addo, M. F., Kuo, G.-H., Hecker, L. R., Croll, D. R., Li, J. & Westover, L. 2004. 3-(7-Azaindolyl)-4-arylmaleimides as potent, selective inhibitors of glycogen synthase kinase-3. *Bioorganic & Medicinal Chemistry Letters* **14**: 3245-3250.
- Zhang, H.-C., White, K. B., Ye, H., McComsey, D. F., Derian, C. K., Addo, M. F., Andrade-Gordon, P., Eckardt, A. J., Conway, B. R., Westover, L., Xu, J. Z., Look, R., Demarest, K. T., Emanuelb, S. & Maryano, B. E. 2003. Macrocyclic bisindolylmaleimides as inhibitors of protein kinase C and glycogen synthase kinase-3. *Bioorganic & Medicinal Chemistry Letters* **13**: 3049 - 3053.
- Zhao, Z., Leister, W. H., Robinson, R. G., Barnett, S. F., Defeo-Jones, D., Jones, R. E., Hartman, G. D., Huff, J. R., Huber, H. E., Duggan, M. E. & Lindsley, C. W. 2005. Discovery of 2,3,5-trisubstituted pyridine derivatives as potent Akt1 and Akt2 dual inhibitors. *Bioorganic & Medicinal Chemistry Letters* **15**: 905-909.



- Zolnierowicz, S. 2000. Type 2A protein phosphatase, the complex regulator of numerous signaling pathways. *Biochemical Pharmacology* **60**: 1225–1235.