

**SCREENING FOR MICROBIAL INHIBITORS  
AGAINST SIGNAL TRANSDUCTION IN  
EUKARYOTES AND *Mycobacterium***

**PUAH SEOK HWA**

PERPUSTAKAAN  
UNIVERSITI MALAYSIA SABAH

**SCHOOL OF SCIENCE AND TECHNOLOGY  
UNIVERSITI MALAYSIA SABAH  
2006**



**UMS**  
UNIVERSITI MALAYSIA SABAH

UNIVERSITI MALAYSIA SABAH

BORANG PENGESAHAN STATUS TESIS @

JUDUL: SCREENING FOR MICROBIAL INHIBITORS AGAINST SIGNAL  
TRANSDUCTION IN EUKARYOTES AND *Mycobacterium*

IJAZAH: SARJANA SAINS (BIOTEKNOLOGI)

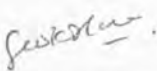
SESI PENGAJIAN: 2002-2006

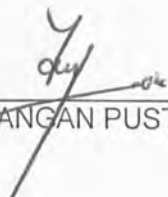
Saya, PUAH SEOK HWA mengaku membenarkan tesis Sarjana ini disimpan di Perpustakaan Universiti Malaysia Sabah dengan syarat-syarat kegunaan seperti berikut:

1. Tesis adalah hakmilik Universiti Malaysia Sabah.
2. Perpustakaan Universiti Malaysia Sabah dibenarkan membuat salinan untuk tujuan pengajian saya.
3. Perpustakaan dibenarkan membuat salinan tesis ini sebagai bahan pertukaran antara institusi pengajian tinggi.
4. TIDAK TERHAD.

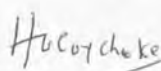
Disahkan oleh

PERPUSTAKAAN  
UNIVERSITI MALAYSIA SABAH

  
(Penulis: PUAH SEOK HWA)

  
(TANDATANGAN PUSTAKAWAN)

Alamat:  
Sekolah Sains dan Teknologi  
Universiti Malaysia Sabah  
Beg Berkunci 2073  
88999 Kota Kinabalu  
Sabah, MALAYSIA.

  
(Penyelia: Prof. Dr. Ho Coy Choke)

**CATATAN:** @ Tesis dimaksudkan sebagai tesis ijazah Doktor Falsafah dan Sarjana secara penyelidikan atau disertasi bagi pengajian secara kerja kursus dan penyelidikan, atau laporan Projek Sarjana Muda (LPSM).



UMS  
UNIVERSITI MALAYSIA SABAH

## DECLARATION

I hereby declare that the materials in this thesis are original except for quotations, excerpts, summaries and references, which have been duly acknowledged.

PUAH SEOK HWA  
PS2002-001-515  
OCTOBER 2006

PERPUSTAKAAN  
UNIVERSITI MALAYSIA SABAH



**UMS**  
UNIVERSITI MALAYSIA SABAH

## ACKNOWLEDGEMENT

I am very grateful to my supervisor Prof. Ho Coy Choke for his guidance and patience along this project.

I am very indebted to Miss Cheah Hwen-Yee for providing H7372 and establishing the yeast two-hybrid screening in this lab.

Many thanks to Prof. Maryati Mohamed and her groups in the Institute for Tropical Biology and Conservation (ITBC), UMS for including and facilitating our participation in the field explorations in Lower Segama and Melalap. Thanks to Mr. Johnny Gisil for his help in plant identifications.

Thanks to WWF Malaysia and the Lundayeh community of Long Pasia for the Ulu Padas trip from which we collected our first batch of field samples.

I would like to thank Prof. Seow Heng Fong of Universiti Putra Malaysia for her kindness in allowing us to use her lab and for her advises in the western blot analysis part of this thesis. My deep appreciation also goes to Pn. Masriana Hassan who has helped in every possible ways.

I would also like to thank Sir Philip Cohen and Dr. Jennifer Bain of the MRC Protein Phosphorylation Unit, University of Dundee, Scotland, U.K. for testing the extract in the *in vitro* kinase assay.

I am thankful to Dr. Peter Csemerly and Dr. Csaba Soti from the Department of Medical Chemistry, Semmelweis University, Hungary of the for performing the surface plasmon resonance and nucleotide affinity binding assay.

Thanks to Dr. Lee Ping-Chin for her advises especially regarding yeast handling.

I appreciate the help of Miss Joan Pereira and Mr. Dewol Sundaling of the Forest Research Centre (FRC) in Sandakan, for aiding us in plant identifications.

Thanks to Mr. Chong Tong Seng, Mr. Lum Mok Sum and Miss Tan Su Hui for their kindness and help in HPLC and many other technical aspects.

Great thanks to fellow friends Hew Chaw Sen, Foo Sek Hin, Ong Si Mon and Ho Wei Loon for all their supports in many ways.

And lastly, thanks to my parents and family.





## ABSTRACT

This study involves the search for microbial inhibitors that disrupt Ras/Raf-1 interaction in the yeast two-hybrid system by the expression of *HIS3* and *lacZ* reporter genes. It is a continuation of the previous study where a *Streptomyces*, H7372 was identified as a possible inhibitor of Ras/Raf-1 protein-protein interaction. H7372 caused larger inhibition zone on histidine minus plates and reduce  $\beta$ -galactosidase activity. Radicicol was also positive in this screening. Radicicol and geldanamycin are two inhibitors of Ras/Raf protein-protein interaction by acting on HSP90 causing the degradation of Raf. H7372 also inhibited the growth of *Mycobacterium smegmatis*. Raf-1 is a serine-threonine kinase. *Mycobacterium* also possessed eukaryotic-like serine-threonine kinases. This creates the opportunity to investigate the possibility that the inhibitor of Raf-1 also inhibit the serine-threonine kinases of *Mycobacterium*. The first part of this study concerned isolating more actinomycetes for screening and the second part was to study H7372 in detail. Eighty actinomycete isolates were obtained by two selective isolation methods using soil samples from three forests in Sabah. No Ras/Raf-1 protein-protein interaction inhibitor was found. One false positive H11337 was identified. The extract was positive in the *HIS3* reporter gene but negative in the *lacZ* reporter gene. Crude freeze-dried extract of H7372 reduced p-ERK1/2 level in MCF-7 cells under insulin stimulation at 250 $\mu$ g/ml and 500 $\mu$ g/ml. A purified fraction, H7372PRE was isolated by cold precipitation method and produced one single peak at 26.03 min retention time by reversed-phase HPLC. H7372PRE inhibited both *HIS3* and *lacZ* reporter expression. H7372PRE reduced the level of p-ERK1/2 at 75 $\mu$ g/ml and 100 $\mu$ g/ml and increased the level of p-MEK1/2 at 75 $\mu$ g/ml and 100 $\mu$ g/ml. HPLC fraction of H7372PRE reduced the level of p-ERK1/2 at 50 $\mu$ g/ml. Crude freeze-dried extract of H7372 decreased JNK/SAPK1c, MAPKAP1a and PKCa kinase activity to 54-63% of the original activity. The binding to HSP90 was not detected by surface plasmon resonance. The crude extract and fraction of H7372 consistently reduced p-ERK level, indicating its intervention to the MAPK pathway, possibly through disruption of Ras/Raf-1 interaction through HSP90. H7372PRE did not inhibit the growth of *M. smegmatis*. H9318 (*Penilicium sp.*) did not inhibit *M. smegmatis* but it's semi-purified fraction, H9318\_S1 that inhibited PP2A and PP1 in vitro inhibited *M. smegmatis*, indicating that the inhibition was possibly through the eukaryotic-like phosphatase of *Mycobacterium*. This study demonstrated that the actinomycetes from forests in Sabah possess interesting compounds that affected the signal transduction pathways in eukaryotes and *Mycobacterium*.



## ABSTRAK

### PENYARINGAN PERENCAT MIKROBIAL TERHADAP TRANSDUKSI ISYARAT DALAM EUKARIOT DAN *Mycobacterium*

Kajian ini melibatkan pencarian perencat mikrobial yang merencat interaksi protein-protein Ras/Raf-1 dalam sistem dua-hibrid yis melalui pengepresan gen pelapor HIS3 dan lacZ. Ia merupakan lanjutan daripada kajian sebelumnya dimana *Streptomyces*, H7372 telah dikenalpasti berkemungkinan besar merencat interaksi protein-protein Ras/Raf-1. H7372 menyebabkan zon rencatan pertumbuhan yang lebih besar dalam piring tanpa histidine dan merendahkan aktiviti  $\beta$ -galaktosidase. Radicicol juga menunjukkan keputusan positif dalam penyaringan ini. Radicicol dan geldanamycin merupakan dua perencat interaksi protein-protein Ras/Raf melalui HSP90 dengan menyebabkan degradasi Raf. H7372 juga merencat pertumbuhan *Mycobacterium smegmatis*. Raf-1 merupakan kinase serine-threonine. *Mycobacterium* juga mempunyai kinase serine-threonine seperti eukariot. Ini menghasilkan peluang untuk menyiasat kemungkinan perencat Ras/Raf-1 juga merencat kinase serine-threonine dalam *Mycobacterium*. Bahagian pertama kajian ini adalah berkenaan mengasingkan aktinomiset untuk penyaringan dan bahagian kedua adalah untuk mengkaji H7372 dengan lebih lanjut. Lapan puluh aktinomiset telah diperoleh dengan kaedah pemencilan selektif menggunakan sampel tanah dari tiga hutan di Sabah. Tiada perencat interaksi-protein-protein Ras/Raf-1 ditemui. Satu positif palsu H11337 dikenalpasti, ia positif dalam gen pelapor HIS3 tetapi negatif dalam gen pelapor lacZ. Ekstrak sejuk kering H7372 merencat p-ERK1/2 dalam sel MCF-7 di bawah rangsangan insulin pada 250 $\mu$ g/ml dan 500 $\mu$ g/ml. Satu fraksi, H7372PRE yang diasingkan melalui kaedah mendakan, menunjukkan puncak tunggal pada 26.03 min dalam analisis HPLC. Ia merencatkan ekspresi kedua-dua gen pelapor HIS3 dan lacZ. H732PRE merencat p-ERK1/2 pada 75 $\mu$ g/ml dan 100 $\mu$ g/ml dan meningkatkan p-MEK1/2 pada 75 $\mu$ g/ml and 100 $\mu$ g/ml. Fraksi HPLC H7372PRE merencatkan p-ERK1/2 pada 50 $\mu$ g/ml. Ekstrak dan fraksi HPLC H7372 merencat p-ERK, ini menandakan ia mengakibatkan perencatan pada laluan MAPK, kemungkinan besar melalui perencatan interaksi Ras/Raf-1 melalui HSP90. Extract sejuk kering H7372 merendahkan aktiviti kinase JNK/SAPK1c, MAPKAP1a and PKCa kepada 54-63% daripada aktiviti asal. Penggabungan kepada HSP90 tidak dikesan melalui ujian 'surface plasmon resonance'. Ekstrak dan fraksi H7372 merencatkan p-ERK secara konsisten. Ini menunjukkan ia mengakibatkan perencatan pada laluan MAPK, kemungkinan merencat interaksi protein-protein Ras/Raf-1 melalui HSP90. H7372PRE tidak merencat pertumbuhan *M. smegmatis*. H9318 (*Penicillium* sp.) tidak merencat pertumbuhan *M. smegmatis* tetapi fraksi separa tulennya, H9318\_S1 yang merencat PP2A and PP1 secara *in vitro* merencat *M. smegmatis* mc<sup>2</sup>155, ini menunjukkan kemungkinan besar perencatan adalah melalui fosfatase yang menyerupai eukariot dalam *Mycobacterium*. Kajian ini telah menunjukkan bahawa aktinomiset dari hutan di Sabah mempunyai kompon yang menarik yang memberi kesan kepada sistem transduksi isyarat dalam eukariot dan *Mycobacterium*.





## LIST OF ABBREVIATIONS

ADP	Adenosine 5'-diphosphate
ATP	Adenosine 5'-triphosphate
CK2	Casein kinase 2
CR1	Conserved domain 1
CR2	Conserved domain 2
CR3	Conserved domain 3
CRD	Cysteine rich domain
DAP	Diaminopimelic acid
DNA-BD	DNA-binding domain
DOTS	Direct Observed Treatment Short-course
DTT	Dithiothreitol
EDTA	Ethylenediaminetetraacetic acid
EGF	Epidermal growth factor
ERK	Extracellular-signal-regulated kinase
FBS	Fetal bovine serum
FGF	Fibroblast growth factor
FT	Farnesyl transferase
G+C	Guanine plus cytosine
GA	Geldanamycin
GAP	GTPase activating protein
GDP	Guanosine diphosphate
GEF	Guanine nucleotide exchange factor
GTP	Guanosine triphosphate
H7372 FD	Crude freeze dried extract of H7372
H7372 PRE	H7372 extract purified by cold precipitation
HIS3	Histidine biosynthesis gene
HPLC	High performance liquid chromatography
HSP90	Heat shock protein 90
HV	Humic acid B-vitamins
ICL	Isocitrate lyase
<i>lacZ</i>	B-galactosidase gene
MBP	Myelin basic protein
MEK	Mitogen-activated protein kinase
MS	Malate synthase
OA	Oatmeal agar
ONPG	O-nitrophenyl- $\beta$ -D-galactosidase
PAGE	Polyacrylamide gel electrophoresis
PAK	P21-activated kinase
PDGF	Platelet-derived growth factor
PMSF	Phenylmethylsulfonyl flouride
PP2A	Protein phosphatase 2A
RA	Radicicol
RBD	Ras binding domain
rRNA	ribosomal Ribonucleic acid
RTK	Receptor tyrosine kinase
SCN	Starch casein nitrate
SDS	Sodium dodecyl sulfate
SOS	Son of sevenless
SPR	Surface plasmon resonance



Ste11	Yeast homologue of Raf
TAD	Transcription activation domain
TBS	Tris-buffered saline
TEMED	N,N,N,N-tetramethylethylenediamine
VEGF	Vascular endothelia growth factor
WHO	World health organization
WSSP	Whole cell sugar pattern
PP1	Protein phosphatase 1
$\beta$	Beta





## TABLE OF CONTENTS

	Page
RESEARCH TITLE	i
DECLARATION	ii
ACKNOWLEDGEMENT	iii
ABSTRACT	iv
ABSTRACK	v
LIST OF ABBREVIATION	vi
TABLE OF CONTENTS	viii
LIST OF TABLE	xii
LIST OF FIGURE	xiv
 <b>CHAPTER 1 – INTRODUCTION</b>	
1.1 Research background	1
1.2 Objectives of study	5
 <b>CHAPTER 2 - LITERATURE REVIEW</b>	
2.1 Actinomycetes	7
2.1.1 Streptomyces	12
2.1.2 Micromonospora	13
2.2 Ras/Raf/MEK/ERK pathway	15
2.2.1 RAS	16
2.2.2 RAF	18
2.2.3 MEK	19
2.2.4 ERK	20
2.2.5 Heat Shock Protein 90 (HSP90)	22
2.3 The Ras/Raf/MEK/ERK pathway inhibitor	24
2.4 Yeast two-hybrid system	31
2.5 Mycobacteria	35
2.5.1 Tuberculosis	35
2.5.2 Signal transduction in mycobacterium	36



## CHAPTER 3 - MATERIALS AND METHODS

### Part A:

#### Isolation, purification of actinomycetes and microbial extract production

3.1	Collection of soil samples	40
3.2	Isolation of actinomycetes	40
3.2.1	Selective isolation of actinomycetes	40
3.2.1.1	Humic Acid B-Vitamins Agar	41
3.2.1.2	Oatmeal Agar	41
3.2.2	Selective isolation of <i>Micromonospora</i>	42
3.2.2.1	Starch Casein Nitrate (SCN) Agar	42
3.3	Storage of actinomycetes isolates	42
3.3.1	20% Glycerol	43
3.4	Production of microbial extracts	43
3.4.1	Mannitol-peptone broth	43
3.5	Detection of 2,6-Diaminopimelic acid (DAP) in whole-cell hydrolysate of actinomycetes isolates by thin layer chromatography (TLC)	43

### Part B:

#### Screening

3.6	Ras-Raf Interaction inhibitor screening	44
3.6.1	Yeast and plasmids strain number	44
3.6.2	Paper disc screening for histidine biosynthesis	47
3.6.2.1	SD medium	47
3.6.3	$\beta$ -galactosidase assay	48
3.6.3.1	Stock solutions for $\beta$ -galactosidase assay	49
3.7	Screening for inhibitors of Isocitrate lyase (ICL) in <i>Mycobacterium smegmatis</i>	50
3.7.1	Modified M9 medium	50

### Part C:

#### H7372 actinomycete extract, bioactivity and purification

3.8	Determination Hsp90 binding activity by crude H7372	51
3.8.1	Surface plasmon resonance and nucleotide affinity cleavage	51
3.9	Inhibition of protein kinase by crude extract of H7372	53
3.10	Preparation of H7372 extract	66
3.10.1	Shake flask fermentation	66
3.10.2	Fermentation of H7372 extract	66



3.10.3 H7372 precipitate preparation (H7372PRE)	66
3.11 Purification and analysis of extracts by HPLC	67
<b>Part D:</b>	
<b>Western blot</b>	
3.12 Cell culture	69
3.12.1 Culture of cell	69
3.12.1.1 Media and solutions for cell culture	69
3.12.2 Cell lysate preparation and protein concentration determination	70
3.12.2.1 Reagents and buffers for cell lysate preparation	70
3.13 SDS-PAGE	71
3.13.1 Reagents for SDS PAGE	71
3.14 Western Blot	72
3.14.1 10X Transfer buffer	72
3.15 Antibodies probing	73
3.15.1 Buffers	73

## CHAPTER 4 – RESULTS

### Part A:

<b>Isolation, purification of actinomycetes and microbial extract production</b>	74
4.1 Sampling Locations	74
4.2 Actinomycete Isolates	79

### Part B:

<b>Screening for microbial inhibitor</b>	87
4.3 Ras/Raf-1 interactions screen	87
4.4 <i>Mycobacterium</i> screening	88
4.5 H11337	88
4.6 Growth inhibitory effect of crude extracts of potential inhibitors of kinase and phosphatase against <i>Mycobacterium</i>	92
4.7 Radicicol	93

### Part C:

#### H7372: bioactivity and purification

4.8 H7372 isolate	95
-------------------	----





4.9	H7372 crude extract bioactivity	97
4.10	H7372 fractions bioactivity	102
4.11	H7372 purification	106
<b>Part D:</b>		
<b>Western Blot</b>		
4.12	Effect of H7372 on MCF-7 cells	116
<b>Part E:</b>		
<b>H9318</b>		
4.13	H9318 effects on <i>Mycobacterium</i>	124
 <b>CHAPTER 5 – DISCUSSION</b>		
5.1	Soil sampling	126
5.2	Actinomycetes isolates	128
5.3	Screening	129
5.4	H7372	131
5.5	Bioactive compounds from <i>Streptomyces</i>	136
5.6	Western blot	137
5.7	H9318	139
 <b>CHAPTER 6 – CONCLUSION</b>		
6.1	Conclusion	140
6.2	Future recommendations	142
<b>REFERENCES</b>		144



## LIST OF TABLES

	Page
<b>Table 4.1:</b> List of soil samples collected from Long Pasia in Sipitang, Lower Segama in Tabin and Melalap in Crocker Range.	75
<b>Table 4.2:</b> List of actinomycetes, especially <i>Streptomyces sp.</i> isolated from HV agar plated on OA	80
<b>Table 4.3:</b> List of actinomycetes isolated by treating soil sample with 1.5% phenol and plated on SCN agar.	82
<b>Table 4.4:</b> Screening data of actinomycetes extracts against Ras/Raf-1 protein-protein interaction screening	87
<b>Table 4.5:</b> Screening data of actinomycetes extracts against <i>M. smegmatis</i> mc <sup>2</sup> 155 on glucose and acetate plates	88
<b>Table 4.6:</b> Crude freeze-dried extracts of H11337 did not inhibit Ras/Raf-1 protein interaction by <i>HIS3</i> reporter	89
<b>Table 4.7:</b> Crude freeze-dried extract of H11337 did not lower $\beta$ -galactosidase activity of LZ cells	89
<b>Table 4.8:</b> Screening data of H11337 fractions against Ras/Raf-1 and <i>M. smegmatis</i> screening	91
<b>Table 4.9:</b> Screening data of selected acetone extracts against <i>M. smegmatis</i> mc <sup>2</sup> 155 on glucose and acetate plates.	92
<b>Table 4.10:</b> Radicicol disrupted Ras/Raf-1 protein interaction in LZ cells as screened by <i>HIS3</i> reporter	93
<b>Table 4.11:</b> Radicicol disrupted Ras/Raf-1 protein interaction in LZ cells as assessed by lowered $\beta$ -galactosidase activity in radicicol-treated cells	93
<b>Table 4.12:</b> H7372 crude acetone extract inhibited LZ cells more on His- plates and reduced $\beta$ -galactosidase activity. It also inhibited <i>M. smegmatis</i> growth on glucose plate	97
<b>Table 4.13:</b> H7372 crude freeze-dried extracts effects on LZ cells and <i>M. smegmatis</i>	98
<b>Table 4.14:</b> Inhibition of protein kinases in vitro by crude freeze-dried extract of H7372 (1mg/ml)	99
<b>Table 4.15:</b> Screening data of H7372 HPLC fractions against Ras/Raf-1 interactions by <i>HIS3</i> reporter	102
<b>Table 4.16:</b> H7372T6 reduced $\beta$ -galactosidase activity of LZ while H7372T5 showed mild reduction	102
<b>Table 4.17:</b> H7372 HPLC fractions were collected every one minute and screened against Ras/Raf-1 interaction	105
<b>Table 4.18:</b> H7372 T6-5 mildly reduced $\beta$ -galactosidase activity of LZ cells	106



<b>Table 4.19:</b>	Screening data of H7372 crude freeze-dried extracts dissolved in various extraction solvents against Ras/Raf-1 interactions	106
<b>Table 4.20:</b>	Screening data of H7372 extracts against Ras/Raf-1 interactions	107
<b>Table 4.21:</b>	H7372PRE, isolated by cold precipitation showed 4mm size differences between His <sup>+</sup> and His <sup>-</sup> plates while H7372AQ, the supernatant did not show any size difference	108
<b>Table 4.22:</b>	H7372PRE markedly reduced $\beta$ -galactosidase activity of LZ cells as opposed to H7372AQ which did not reduce $\beta$ -galactosidase activity	108
<b>Table 4.23:</b>	H7372PRE showed increased size differential against Ras/Raf-1 interaction screening upon increased concentration	109
<b>Table 4.24:</b>	Screening data of H7372PRE HPLC fractions against Ras/Raf-1 interaction by <i>HIS3</i> reporter	115
<b>Table 4.25:</b>	$\beta$ -galactosidase activity of H7372PRE and HPLC fraction of H7372PRE	115
<b>Table 4.26:</b>	H9318_S1 and H9318_S2 effects on <i>M. smegmatis</i> mc <sup>2</sup> 155	124





## LIST OF FIGURES

	Page
<b>Figure 2.1:</b> Phylogenetic relationship of 90 genera of actinomycetes and actinobacteria based on 16SrRNA sequences (Yokota, 1997b)	11
<b>Figure 2.2:</b> The organization and function of the Ras/Raf/MEK/ERK pathway (Kolch et al., 2002)	14
<b>Figure 2.3:</b> Ras structure (modified from Midgley & Kerr, 2002).	16
<b>Figure 2.4:</b> Raf-1 structure (modified from Wellbrock <i>et al.</i> , 2004)	18
<b>Figure 2.5:</b> Human HSP90 (modified from Nollen & Morimoto, 2002 and Stebbins <i>et al.</i> , 1997)	22
<b>Figure 2.6:</b> STI571 (Gleevec)	25
<b>Figure 2.7:</b> Manumycin	25
<b>Figure 2.8:</b> BAY43-9006	26
<b>Figure 2.9:</b> ZM336372	26
<b>Figure 2.10:</b> PD98059	27
<b>Figure 2.11:</b> U0126	27
<b>Figure 2.12:</b> Radicicol	28
<b>Figure 2.13:</b> KF25706	28
<b>Figure 2.14:</b> Geldanamycin and its derivative 17-N allylamino-17 demethoxy geldanamycin (17AAG)	29
<b>Figure 2.15:</b> 8-amino adenosine	30
<b>Figure 2.16:</b> UCS15A and it's derivative 2c	31
<b>Figure 2.17:</b> Schematic drawing of the yeast two-hybrid system in the detection of inhibitor	24
<b>Figure 2.18:</b> 1-(5-isoquinolinesulphonyl)-2-methylpiperazine (H7)	39
<b>Figure 3.1:</b> Construct of pBTM116 and pVP16 plasmids (Cheah,2003).	46
<b>Figure 3.2:</b> H7372 extracts preparation	68
<b>Figure 4.1:</b> Soil sampling locations in Ulu Padas, Sipitang.	76
<b>Figure 4.2:</b> Soil sampling locations in Melalap, Crocker Range.	77
<b>Figure 4.3:</b> Soil sampling locations in Lower Segama, Tabin.	78
<b>Figure 4.4:</b> Microscopic observation of H11143 and H11159 under light microscope.	84
<b>Figure 4.5:</b> Aerial mycelia and substrate mycelia view of four isolates, showing colour variation	85
<b>Figure 4.6:</b> Separation of DAP isomers of whole cell lysate of actinomycetes by TLC.	86
<b>Figure 4.7:</b> HPLC profile of H11337	90



<b>Figure 4.8:</b>	Radical effect against LZ yeast	94
<b>Figure 4.9:</b>	H7372 isolate	96
<b>Figure 4.10:</b>	Microscopic examination of H7372	96
<b>Figure 4.11:</b>	SPR sensorgram of H7372 crude freeze dried extract showed H7372 at 300µg/ml did not bind purified immobilized Hsp90	100
<b>Figure 4.12:</b>	Detection of Hsp90 N- and C-terminal ATP site binding after treatment with H7372	101
<b>Figure 4.13:</b>	H7372 HPLC fractions T1-T5 screened against Ras/Raf-1	103
<b>Figure 4.14:</b>	H7372 HPLC fractions T6-T10 screened against Ras/Raf-1	104
<b>Figure 4.15:</b>	HPLC profile of H7372FD	105
<b>Figure 4.16:</b>	Screening of H7372PRE and H7372AQ (supernatant) against Ras/Raf-1 protein-protein interaction screening.	109
<b>Figure 4.17:</b>	Clockwise from top - discs 1 to 5 with increasing concentration of H7372 PRE extract	110
<b>Figure 4.18:</b>	HPLC profile of H7372PRE	111
<b>Figure 4.19:</b>	HPLC analysis of H7372PRE.	112
<b>Figure 4.20:</b>	HPLC analysis of H7372AQ.	113
<b>Figure 4.21:</b>	Overlay view of H7372PRE (bold) and H7372AQ	114
<b>Figure 4.22:</b>	HPLC profile of H7372PRE at higher sample volume	114
<b>Figure 4.23:</b>	H7372 crude freeze-dried extract reduced p-ERK level of MCF-7 cells in a concentration dependent manner.	116
<b>Figure 4.24:</b>	H7372 crude freeze dried extract lowered p-ERK level of MCF-7 cells	117
<b>Figure 4.25:</b>	H7372PRE lowered p-ERK level on a concentration dependent manner in MCF-7 cells.	118
<b>Figure 4.26:</b>	H7372PRE HPLC bioactive fraction lowered p-ERK level of MCF-7 cells	119
<b>Figure 4.27:</b>	H7372PRE HPLC bioactive fraction have mild inhibitory effect on Raf-1 of MCF-7 cells	120
<b>Figure 4.28:</b>	Radical lowered p-ERK level and p-MEK level of MCF-7 cells	121
<b>Figure 4.29:</b>	Radical lowered Raf-1 level of MCF-7 cells	122
<b>Figure 4.30:</b>	Geldanamycin lowered p-ERK, p-Akt and p-GSK3β level of MCF-7 cells	123
<b>Figure 4.31:</b>	Inhibitory effect of H9318_S1 on <i>M. smegmatis</i> mc <sup>2</sup> 155	125





# CHAPTER 1

## INTRODUCTION

### 1.1 Research background

Bioactive compounds are on a new level of pursuit. Unlike the conventional ways, screenings are now targeted not only at growth inhibition of pathogenic microorganisms but also against signal transduction pathways believed to be key players regulating vital cellular functions. Actinomycetes, high G+C, Gram positive bacteria play the central role in drug discovery. The *Streptomyces* and *Micromonospora* are abundant actinomycetes in soil. *Streptomyces* are best known as producers of bioactive compounds like streptomycin and doxorubicin and are currently use as anti-infective and anticancer drugs. Gentamicin is known to be produced by *Micromonospora*. The major differences between the two is the absent of aerial mycelia of *Micromonospora*, extremely slow growth of the bacteria and the differences in cell wall constituents (Kudo, 1997; Yokota, 1997).

Chemical novelty associated with natural product is high and metabolites produced by bacteria provide great structural diversity. More than half of known antibiotics are of microbial origin like kanamycin, streptomycin and erythromycin, whereas daunorubicin, doxorubicin and mitomycin are important antitumor agents (Demain, 1999). Regulatory factors like A-factor, implicated in both morphological differentiation and secondary metabolism, is thought to be responding to environmental and physical stresses (Bibb, 2005). Kinase





inhibitors produced by actinomycetes are like staurosporine, a broad range kinase inhibitor and K252a, protein kinase C inhibitor. There are also protein phosphatase inhibitors like tautomycin, fostriecin and FK506 of *Streptomyces sp.*

Borneo is the second largest tropical island in the world harbouring nature's precious biodiversity resources where its potential has yet to be fully explored. The actinomycetes diversity and abundance is high in the tropical forest region (Wang *et al.*, 1999). The search for biologically active compounds has led us to exploit the forest area in the hope for finding higher microbial diversity that provides better chance at obtaining a hit. Geldanamycin, leptomycin, mitomycin, manumycin, luminacins are all produced by *Streptomyces sp.* and have anti tumour activities by interfering with cell signalling.

In this study, actinomycetes were isolated from soil samples collected from just below the leaf litter as the degradation of the leaf litters and other dead organic material were aided by microbes in which many of them are fungi and actinomycetes. These microbes were selectively isolated and the acetone extracts produced by shake flask fermentation of these isolates were screened against Ras/Raf-1 protein-protein interaction screening using the yeast two-hybrid screening, which was designed to screen for inhibitors affecting mammalian Ras/Raf-1 interaction. Potential inhibitor of the screening affects the interaction directly or indirectly, as demonstrated by radicicol (Ki *et al.*, 1998).

The Ras/Raf/MEK/ERK pathway is important in governing cell proliferation, differentiation, transformation and apoptosis. Growth factors bind to their associated receptors (which are receptor tyrosine kinase) on the cell membrane to activate cascades of signals, relaying cellular information from the cell membrane to the nucleus (Kolch, 2000). The activation steps of Ras protein includes localization of Ras protein to the membrane, ras dimerization and



involving guanine nucleotide exchange factor (GEF), which serve to accelerate the exchange of RAS-GDP to RAS-GTP. RAS-GTP. The active Ras attained a conformation enabling it to bind to Raf-1. In order to return to its resting state RAS-GAP interacts with GTPase activating protein (GAP) (Inouye *et al.*, 2000). Activated Ras will then activate Raf, a process involving many other proteins and critically phosphorylation at S338 (Edin & Juliano, 2005) and dephosphorylation at S259 by PP2A which leads to membrane localization, for subsequent activation step (Kubicek *et al.*, 2002), and S471 phosphorylation for MEK binding (Zhu *et al.*, 2005). Raf activates MEK (Lee & McCubrey, 2002) and subsequently ERK (Zheng & Guan, 1993). The RAS-MAPK pathway is a key signalling mechanism that regulates many cellular functions such as cell growth, transformation and apoptosis. Regulation of both Ras and Raf is crucial in the proper maintenance of cell growth as oncogenic mutations in these genes lead to high transforming activity.

Hsp90 is present in the cytoplasm of eubacteria, yeast and multicellular organisms, it plays an important role in the conformational regulation of key signalling molecules including various steroid hormone receptors, kinases including Raf-1 and Ste11 and other proteins (Buchner, 1999). The disruption of Hsp90 by an ansamycin, geldanamycin leads to destabilization of Raf and thereof prevented the association of Ras with Raf (Schulte *et al.*, 1995). Radicicol, a macrolide antibiotic was also affecting Raf-1 through Hsp90 (Schulte *et al.*, 1999). Novobiocin, unlike geldanamycin and radicicol, act on the carboxyl terminal ATP binding site, also disrupting Hsp90 function (Marcu *et al.*, 2000).

Tuberculosis is reemerging world wide at alarming rate. Apart from the multi drug resistance tuberculosis (MDR-TB), latent persistent infection is posing greater fear as the infected individual will not show any symptom until his



immune system becomes compromised. The bacteria reside in the human lung for decades unnoticed. Several evading mechanism of this bacteria is being noticed, the (isocitrate lyase) ICL and malate synthase (MS) of the glyoxylate cycle are two potential targets.

There are 11 eukaryotic-type serine/threonine kinases (PknA, B, D, E, F, G, H, I, J, K and L) and 4 phosphoprotein phosphatases in *Mycobacterium tuberculosis* genome (Cole *et al.*, 1998). Their presumptive roles includes cell division and differentiation regulation, receptor for environmental sensing and most importantly PknG was found to be vital in mediating intracellular survival of mycobacteria (Walburger *et al.*, 2004). Inhibition of these eukaryotic like kinases or phosphatases is therefore an attractive target. H7 (1-(5-isoquinolinesulfonyl)-2-methylpiperazine), a protein kinase C inhibitor was found inhibit *M. smegmatis* and *M. bovis* BCG growth (Drews *et al.*, 2001).

Mycobacterial serine/threonine phosphatase (mstp) role is implicated in cell division regulation. Mstp is a transmembrane protein that dephosphorylates phosphorylated PknA and PknB on serine/threonine residues. The presence of mstp in slow growing mycobacterial species, its transmembrane localization, and ability to dephosphorylate phosphorylated PknA and PknB implicates that Mstp may play a role in regulating cell division in *M. tuberculosis* (Chopra *et al.*, 2003).

H7372 is a *Streptomyces* isolated from soil from root of *Bruguiera sp.*, a plant found in the mangrove swamp ecosystem. It was found to inhibit Ras/Raf-1 protein-protein interaction, H7372, using yeast two-hybrid screening. This isolate was aerobically cultured to produce bioactive compounds. The crude freeze-dried extract was found to reduce p-ERK and p-MEK level in NIH3T3 cells. H7372 extract was also found to inhibit ICL of the *Mycobacterium* (Cheah, 2003). The compound of interest is therefore pursued.





H9318 was a *Penicillium sp.* Isolated from soil. The extract was picked up from yeast based screening against PP1 (protein phosphatase 1). The crude extract was found to positively inhibit PP1 and PP2A (protein phosphatase 2A) by the *in vitro* test (Voo, 2004). Two fractions, H9318\_S1 and H9318\_S2 were isolated by HPLC of ethyl acetate extract of H9318. H9318\_S1 and H9318\_S2 were later identified to have similar bioactive properties by the yeast based screening and the *in vitro* dephosphorylation assay (Ong, 2005).

## 1.2 Objectives of study

- a. To isolate diverse new actinomycete from forest soils of Sabah using selective isolation methods for actinomycetes particularly *Streptomyces* and *Micromonospora*.
- b. To screen for inhibitors of mammalian Ras/Raf-1 protein-protein interaction by the yeast two-hybrid system from the actinomycetes isolates and compare them with geldanamycin or radicicol. Geldanamycin from *Streptomyces hygroscopicus* (actinomycete) and radicicol from *Monosporium bonorden* (fungus) are two inhibitors that affect Raf-1 through HSP90.
- c. H7372 was identified as Ras/Raf-1 protein-protein interaction inhibitor by yeast two-hybrid system in previous study. H7372 protein kinases inhibition and Hsp90 binding was assessed by *in vitro* kinase assay and surface plasmon resonance (SPR) assay in comparison to geldanamycin or radicicol.
- d. To purify bioactive fraction of H7372 actinomycetes extract that effected the mammalian Ras/Raf-1 interaction in the yeast two-hybrid system in both the *HIS3* and *LacZ* reporter.

- e. To examine the effects of H7372 crude extracts and purified fraction towards phosphorylation level of MAPK pathway key proteins in MCF-7 human fibroblast cell line through immunoblotting approach.
- f. To test the inhibitory effects of H7372 purified fraction against *Mycobacterium smegmatis* mc<sup>2</sup>155 through paper disc susceptibility test. H7372 was found to be inhibitor of ICL in *M. smegmatis* mc<sup>2</sup>155 based assay by previous work.
- g. To test the inhibitory effects of H9318\_S1 and H9318\_S2 [HPLC fractions of *Penicillium sp.* extract, identified through yeast based screening and *in vitro* dephosphorylation assay as inhibitors of mammalian protein phosphatases (PP1 and PP2A)] against *Mycobacterium smegmatis* mc<sup>2</sup>155 through paper disc susceptibility test.



## CHAPTER 2

### LITERATURE REVIEW

#### 2.1 Actinomycetes

Actinomycetes are Gram positive, high G+C content bacteria. They are morphologically characterized by the type and stability of the mycelium, the number, type and disposition of the spores and among others formation of sporangia and flagellate elements. The chemical characterizations include the cell wall and whole cells sugars compositions alongside with type of lipids found. Cell wall type I has LL-DAP (LL-2,6-diaminopimelic acid) and glycine with 11 members of different genera like *Streptomyces*, *Streptoveticiillum*, *Nocardioides*, *Microellobosporia* and *Arachnia* they are further differentiated based on morphology and physiological differences. There are a few genera having type II cell wall, all five of them are *Micromonospora*, *Actinoplanes*, *Amorphosporagium*, *Ampullariella* and *Dactylosporangium*. The cell wall contains *meso*-DAP and glycine. Xylose and arabinose are the diagnostic sugars of the whole cell sugar pattern (WSSP). Type III and IV cell wall also contain *meso*-DAP. *Actinomadura* and *Frankia* are members of the Type III group along with 13 others with WSSP with or without madurose and with diverse physical morphology. As for Type IV, the cell wall and WSSP contains arabinose and galactose with *Mycobacterium* and *Nocardia* as the more well known members. There are up to 9 cell wall type with different cell wall compositions and other members of the actinomycetes (Lechevalier & Lechevalier, 1981).





Actinomycetes on a solid medium forms the substrate mycelium or the primary mycelium, which penetrate into the agar. The aerial mycelium, which is permanently in contact with air is formed later (Vobis, 1997). The colonies consistencies varies: raised, flat, hard, soft, leathery, smooth, wrinkled, granular etc. There are a wide spectrum of colours for the colonies including white, yellow, orange, red, purple, blue, green, brown, grey and black. Growth of filamentous actinomycetes is completely different, in the sense both of biomass increase and cell division, from the patterns of cell enlargement and binary fission that occur in unicellular bacteria. Vegetative hyphae grow into long, often multiply branched masses of tangled threads without undergoing cell division. The result is large, essentially multinucleate cells. Separation of cytoplasm and nuclei by septa occurs sporadically and without apparent pattern. Reproduction occurs by a variety of mechanisms of fragmentation of hyphae to form haploid spherical cells in the nonsporeforming genera such as *Nocardia* or by differentiation of the fragments to spores in other genera (Ensign, 1999).

Actinomycetes can be found easily in humus rich soil, the ecological niche of most actinomycetes is probably the aerobic zone of soil whereby they are actively involved in the degradation of leaf litters and other organic substrates. It was estimated that there are more than one million of them per gram of soil (Nonomura & Hayakawa, 1988). Actinomycetes can also be isolated from river mud, lake bottom and also swampy area. Some corynebacteria, mycobacteria and species of *Actinomyces* are normal microbial flora of the human skin and also found in gingival tissue of the mouth.

Secondary metabolism is brought on by exhaustion of a nutrient, biosynthesis or addition of an inducer, and/or by a growth rate decrease. The deep-seated reason for the production of these secondary metabolites that are



### Reference:

- Alessi, D. R., Cuenda, A., Cohen, P., Dudley, D. T. & Saltiel, A. R. 1995. PD 098059 is a specific inhibitor of the activation of mitogen-activated protein kinase kinase *in vitro* and *in vivo*. *The Journal of Biological Chemistry*. **270**:27489-27494.
- Andersen, M. H., Fensterle, J., Ugurel, S., Reker, S., Houben, R., Guldberg, P., Berger, T. G., Schadendorf, D., Trefzer, U., Brocker, E.-B., Straten, P. t., Rapp, U. R. & Becker, J. C. 2004. Immunogenicity of constitutively active <sup>V599E</sup>BRaf. *Cancer Research*. **64**:5456-5460.
- Appels, N. M. G. M., Beijnen, J. H. & Schellens, J. H. M. 2005. Development of farnesyl transferase inhibitors: a review. *Oncologist*. **10**:565-578.
- Auerbach, D., Arnoldo, A., Bogdan, B., Fetchko, M. & Stagljari, I. 2005. Drug discovery using yeast as a model system: a functional genomic and proteomic view. *Current Proteomics*. **2**:1-13.
- Av-Gay, Y. & Everett, M. 2000. The eukaryotic-like Ser/Thr protein kinases of *Mycobacterium tuberculosis*. *Trends in Microbiology*. **8**:238-244.
- Baltz, R. H. 2001. Genetic methods and strategies for secondary metabolite yield improvement in actinomycetes. *Antonie van Leeuwenhoek*. **79**:251-259.
- Bar-Sagi, D. 2001. A ras by any other name. *Molecular and Cellular Biology*. **21**:1441-1443.
- Besant, P. G., Tan, E. & Attwood, P. V. 2003. Mammalian protein histidine kinases. *The International Journal of Biochemistry and Cell Biology*. **35**:297-309.
- Bibb, M. J. 2005. Regulation of secondary metabolism in *Streptomyces*. *Current Opinion in Microbiology*. **8**:208-215.
- Bliska, J. B., Guan, K., Dixon, J. E. & Falkow, S. 1991. Tyrosine phosphate hydrolysis of host proteins by an essential *Yersinia* virulence determinant. *Proceedings of the National Academy of Sciences*. **88**:1187-1191.
- Blum, R. & Kloog, Y. 2005. Tailoring Ras-pathway-Inhibitor combinations for cancer therapy. *Drug Resistance Updates*. **8**:369-380.
- Boitel, B., Ortiz-Lombardía, M., Durán, R., Pompeo, F., Cole, S. T., Cerveñansky, C. & Alzari, P. M. 2003. PknB kinase activity is regulated by phosphorylation in two Thr residues and dephosphorylation by PstP, the cognate phospho Ser/Thr phosphatase, in *Mycobacterium tuberculosis*. *Molecular Microbiology*. **49**:1493-1508.
- Bossio, D. A. & Scow, K. M. 1995. Impact of carbon and flooding on the metabolic diversity of microbial communities in soils. *Applied and Environmental Microbiology*. **61**:4043-4050.





- Brent, R. & Finley Jr, R. L. 1997. Understanding gene and allele function with two-hybrid methods. *Annual Reviews in Genetics*. **31**:663-704.
- Buchner, J. 1999. Hsp90 & Co. - a holding for folding. *Trends in Biochemical Sciences*. **24**:136-141.
- Bull, A. T., Ward, A. C. & Goodfellow, M. 2000. Search and discovery strategies for biotechnology: the paradigm shift. *Microbiology and Molecular Biology Reviews*. **64**:573-606.
- Catling, A., Schaeffer, H., Reuter, C., Reddy, G. & Weber, M. 1995. A proline-rich sequence unique to MEK1 and MEK2 is required for raf binding and regulates MEK function. *Molecular and Cellular Biology*. **15**:5214-5225.
- Chaba, R., Raje, M. & Chakraborti, P. K. 2002. Evidence that a eukaryotic-type serine/threonine protein kinase from *Mycobacterium tuberculosis* regulates morphological changes associated with cell division. *European Journal of Biochemistry*. **269**:1078-1085.
- Chang, D., Panageas, K., Osman, I., Polsky, D., Busam, K. & Chapman, P. 2004. Clinical significance of BRAF mutations in metastatic melanoma. *Journal of Translational Medicine*. **2**:46.
- Cheah, H.-Y. 2003. Isolation of actinomycetes from mangroves in Sabah and screening for inhibitors against eukaryotic signal transduction. Master of Science. Universiti Malaysia Sabah.
- Chien, C.-T., Bartel, P. L., Sternglanz, R. & Fields, S. 1991. The two-hybrid system: a method to identify and clone genes for proteins that interact with a protein of interest. *Proceedings of the National Academy of Sciences*. **88**:9578-9582.
- Chopra, P., Singh, B., Singh, R., Vohra, R., Koul, A., Meena, L. S., Koduri, H., Ghildiyal, M., Deol, P. & Das, T. K. 2003. Phosphoprotein phosphatase of *Mycobacterium tuberculosis* dephosphorylates serine-threonine kinases PknA and PknB. *Biochemical and Biophysical Research Communications*. **311**:112-120.
- Coates, P. & Hall, P. 2003. The yeast two-hybrid system for identifying protein-protein interactions. *Journal of Pathology*. **199**:4-7.
- Cohen, P. 2002. Protein kinases--the major drug targets of the twenty-first century? *Nature Review Drug Discovery*. **1**:309-315.
- Cole, S. T., Brosch, R., Parkhill, J., Garnier, T., Churcher, C., Harris, D., Gordon, S. V., Eiglmeier, K., Gas, S., Barry III, C. E., Tekaia, F., Badcock, K., Basham, D., Brown, D., Chillingworth, T., Connor, R., Davies, R., Devlin, K., Feltwell, T., Gentles, S., Hamlin, N., Holroyd, S., Hornsby, T., Jagels, K., Krogh, A., McLean, J., Moule, S., Murphy, L., Oliver, K., Osborne, J., Quail, M. A., Rajandream, M.-A., Rogers, J., Rutter, S., Seeger, K., Skelton, J., Squares, R., Squares, S., Sulston, J. E., Taylor, K., Whitehead, S. & Barrell, B. G. 1998. Deciphering the biology of *Mycobacterium tuberculosis* from the complete genome sequence. *Nature*. **393**:537-544.





- Cowley, S., Ko, M., Pick, N., Chow, R., Downing, K. J., Gordhan, B. G., Betts, J. C., Mizrahi, V., Smith, D. A., Stokes, R. W. & Av-Gay, Y. 2004. The *Mycobacterium tuberculosis* protein serine/ threonine kinase PknG is linked to cellular glutamate/ glutamine levels and is important for growth in vivo. *Molecular Microbiology*. **52**:1691-1702.
- Davies, S. P., Reddy, H., Caivano, M. & Cohen, P. 2000. Specificity and mechanism of action of some commonly used protein kinase inhibitors. *Biochemistry Journal*. **351**:95-105.
- Demain, A. L. 1998. Induction of microbial secondary metabolism. *International Microbiology*. **1**:259-264.
- Demain, A. L. 1999. Pharmaceutically active secondary metabolites of microorganisms. *Applied Microbiology and Biotechnology*. **52**:455-463.
- Deol, P., Vohra, R., Saini, A. K., Singh, A., Chandra, H., Chopra, P., Das, T. K., Tyagi, A. K. & Singh, Y. 2005. Role of *Mycobacterium tuberculosis* Ser/Thr kinase PknF: implications in glucose transport and cell division. *Journal of Bacteriology*. **187**:3415-3420.
- Dhillon, A. S., Meikle, S., Yazici, Z., Eulitz, M. & Kolch, W. 2002. Regulation of Raf-1 activation and signalling by dephosphorylation. *Embo J*. **21**:64-71.
- Diaz, B., Barnard, D., Filson, A., MacDonald, S., King, A. & Marshall, M. 1997. Phosphorylation of Raf-1 serine 338-serine 339 is an essential regulatory event for Ras-dependent activation and biological signaling. *Molecular and Cellular Biology*. **17**:4509-4516.
- Drews, S. J., Hung, F. & Av-Gay, Y. 2001. A protein kinase inhibitor as an antimycobacterial agent. *FEMS Microbiology Letters*. **205**:369-374.
- Dudley, D., Pang, L., Decker, S., Bridges, A. & Saltiel, A. 1995. A synthetic inhibitor of the mitogen-activated protein kinase cascade. *Proceedings of the National Academy of Sciences*. **92**:7686-7689.
- Dumaz, N. & Marais, R. 2005. Integrating signals between cAMP and the RAS/RAF/MEK/ERK signalling pathways: Based on The Anniversary Prize of the Gesellschaft fur Biochemie und Molekularbiologie Lecture delivered on 5 July 2003 at the Special FEBS Meeting in Brussels. *FEBS J*. **272**:3491-3504.
- Edin, M. L. & Juliano, R. L. 2005. Raf-1 serine 338 phosphorylation plays a key role in adhesion-dependent activation of extracellular signal-regulated kinase by epidermal growth factor. *Molecular and Cellular Biology*. **25**:4466-4475.
- Ensign, J. C. 1999. Introduction to the Actinomycetes. <http://link.springer-ny.com/link/service/books/10125/>
- Eppstein, A. C., Sandoval, J. A., Klein, P. J., Woodruff, H. A., Grosfeld, J. L., Hickey, R. J., Malkas, L. H. & Schmidt, C. M. 2006. Differential sensitivity of chemoresistant neuroblastoma subtypes to MAPK-targeted treatment correlates with



- ERK, p53 expression, and signaling response to U0126. *Journal of Pediatric Surgery*. **41**:252-259.
- Fabian, J. R., Daar, I. O. & Morrison, D. K. 1993. Critical tyrosine residues regulate the enzymatic and biological activity of Raf-1 kinase. *Molecular and Cellular Biology*. **13**:7170-7179.
- Fontan, P. A., Walters, S. & Smith, I. 2004. Cellular signaling pathways and transcriptional regulation in *Mycobacterium tuberculosis*: Stress control and virulence. *Current Science*. **86**:122-134.
- Ghiasi, K., Ma, C., Gandhi, V., Plataniias, L. C., Krett, N. L. & Rosen, N. 2005. 8-amino-adenosine induces loss of phosphorylation of p38 mitogen-activated protein kinase, extracellular signal-regulated kinase 1/2, and Akt kinase: Role in induction of apoptosis in multiple myeloma. *Mol Cancer Ther*. **4**:569-577.
- Gietz, R. D., Triggs-Raine, B., Robbins, A., Graham, K. C. & Woods, R. A. 1997. Identification of proteins that interact with a protein of interest: Applications of the yeast two hybrid system. *Molecular and Cellular Biochemistry*. **172**:67-79.
- Gil, J. A. & Martin, J. F. 1997. Polyene antibiotics. In *Biotechnology of antibiotics*. Strohl, W.R., editor. Marcel Dekker Inc, New York. 551-575.
- Gonzalez, L., Phalip, V. & Zhang, C.-C. 2001. Characterization of PknC, a Ser/Thr kinase with broad substrate specificity from the cyanobacterium *Anabaena* sp. strain PCC 7120. *Eur. J. Biochem*. **268**:1869-1875.
- Grenert, J. P., Sullivan, W. P., Fadden, P., Haystead, T. A. J., Clark, J., Mimnaugh, E., Krutzsch, H., Ochel, H.-J., Schulte, T. W., Sausville, E., Neckers, L. M. & Toft, D. O. 1997. The amino-terminal domain of heat shock protein 90 (hsp90) that binds geldanamycin is an ATP/ADP switch domain that regulates hsp90 conformation. *The Journal of Biological Chemistry*. **272**:23843-23850.
- Hall-Jackson, C. A., Eyers, P. A., Cohen, P., Goedert, M., Tom Boyle, F., Hewitt, N., Plant, H. & Hedge, P. 1999. Paradoxical activation of Raf by a novel Raf inhibitor. *Chemistry and Biology*. **6**:559-568.
- Hara, M., Akasaka, K., Akinaga, S., Okabe, M., Nakano, H., Gomez, R., Wood, D., Uh, M. & Tamanoi, F. 1993. Identification of Ras farnesyltransferase inhibitors by microbial screening. *Proc. Natl. Acad. Sci. USA*. **90**:2281-2285.
- Hayakawa, M. & Nonomura, H. 1987. Humic acid-vitamin agar, a new medium for the selective isolation of soil actinomycetes. *Journal of Fermentation Technology*. **65**:501-509.
- Hayakawa, M., Sadakata, T., Kajiura, T. & Nonomura, H. 1991. New methods for the high selective isolation of *Micromonospora* and *Microbispora* from soil. *Journal of Fermentation and Bioengineering*. **72**:320-326.





- Hayakawa, M., Yoshida, Y. & Imura, Y. 2004. Selective isolation of bioactive soil actinomycetes belonging to the *Streptomyces violaceusniger* phenotypic cluster. *Journal of Applied Microbiology*. **96**:973-981.
- Ho, C. C. 2003. Molecular cell biology, biodiversity and biotechnology. In Professorial lecture series. Universiti Malaysia Sabah, Kota Kinabalu. 61.
- Hutchinson, C. R. 1997. Antibiotics from genetically engineered microorganism. In *Biotechnology of antibiotics*. Strohl, W.R., editor. Marcel Dekker, New York. 683-702.
- Inouye, K., Mizutani, S., Koide, H. & Kaziro, Y. 2000. Formation of the Ras dimer is essential for Raf-1 activation. *The Journal of Biological Chemistry*. **275**:3737-3740.
- Jain, R. & Inouye, S. 1998. Inhibition of development of *Myxococcus xanthus* by eukaryotic protein kinase inhibitors. *Journal of Bacteriology*. **180**:6544-6550.
- Janssen, R. A. J., Veenstra, K. G., Jonasch, P., Jonasch, E. & Mier, J. W. 1998. Ras- and Raf-induced down-modulation of non-muscle tropomyosin are MEK-independent. *The Journal of Biological Chemistry*. **273**:32182-32186.
- Jelinek, T., Catling, A. D., Reuter, C. W. M., Moodie, S. A., Wolfman, A. & Weber, M. J. 1994. RAS and RAF-1 form a signalling complex with MEK-1 but not MEK-2. *Molecular and Cellular Biology*. **14**:8212-8218.
- Juris, S. J., Rudolph, A. E., Huddler, D., Orth, K. & E., D. J. 2000. A distinctive role for the *Yersinia* protein kinase: Actin binding, kinase activation, and cytoskeleton disruption. *Proceedings of the National Academy of Sciences*. **97**:9431-9436.
- Kampfer, P. 2006. The Family Streptomycetaceae, Part I: Taxonomy. <http://link.springer-ny.com/link/service/books/10125/>
- Kato-Stankiewicz, J., Hakimi, I., Zhi, G., Zhang, J., Serebriiskii, I., Guo, L., Edamatsu, H., Koide, H., Menon, S., Eckl, R., Sakamuri, S., Lu, Y., Chen, Q.-Z., Agarwal, S., Baumbach, W. R., Golemis, E. A., Tamanoi, F. & Khazak, V. 2002. Inhibitors of Ras/Raf-1 interaction identified by two-hybrid screening revert Ras-dependent transformation phenotypes in human cancer cells. *Proceedings of the National Academy of Sciences*. **99**:14398-14403.
- Ki, S. W., Kasahara, K., Kwon, H. J., Eishima, J., Takesako, K., Cooper, J. A., Yoshida, M. & Horinouchi, S. 1998. Identification of radicicol as an inhibitor of *in vivo* Ras/Raf interaction with the yeast two-hybrid screening system. *The Journal of Antibiotics*. **51**:936-944.
- Kim, S. B. & Goodfellow, M. 1999. Reclassification of *Amycolatopsis rugosa* Lechevalier et al. 1986 as *Prauserella rugosa* gen. nov., comb. nov. *International Journal of Systematic Bacteriology*. **49**:507-512.
- Kim, S. B., Lonsdale, J., Seong, C.-N. & Goodfellow, M. 2003. Streptacidiphilus gen. nov., acidophilic actinomycetes with wall chemotype I and emendation of the family Streptomycetaceae. *Antonie van Leeuwenhoek*. **83**:107-116.





- Kolch, W. 2000. Meaningful relationships: the regulation of the Ras/Raf/MEK/ERK pathway by protein interactions. *Biochemistry Journal*. **351**:289-305.
- Kolch, W., Kotwaliwale, A., Vass, K. & Janosch, P. 2002. The role of Raf kinases in malignant transformation. *Expert Reviews in Molecular Medicine*. **2002**:1-18.
- Koul, A., Choidas, A., Treder, M., Tyagi, A. K., Drlica, K., Singh, Y. & Ullrich, A. 2000. Cloning and characterization of secretory tyrosine phosphatases of *Mycobacterium tuberculosis*. *Journal of Bacteriology*. **182**:5425-5432.
- Koul, A., Choidas, A., Tyagi, A. K., Drlica, K., Singh, Y. & Ullrich, A. 2001. Serine/threonine protein kinases PknF and PknG of *Mycobacterium tuberculosis*: characterization and localization. *Microbiology*. **147**:2307-2314.
- Koul, A., Herget, T., Klebl, B. & Ullrich, A. 2004. Interplay between mycobacteria and host signaling pathways. *Nature Reviews Microbiology*. **2**:189-202.
- Krett, N. L., Davies, K. M., Ayres, M., Ma, C., Nabhan, C., Gandhi, V. & Rosen, S. T. 2004. 8-Amino-adenosine is a potential therapeutic agent for multiple myeloma. *Molecular Cancer Therapeutics*. **3**:1411-1419.
- Kubicek, M., Pacher, M., Abraham, D., Podar, K., Eulitz, M. & Baccarini, M. 2002. Dephosphorylation of Ser-259 regulates Raf-1 membrane association. *The Journal of Biological Chemistry*. **277**:7913-7919.
- Kudo, T. 1997. Family *Streptomycetaceae*. In Atlas of actinomycetes. Miyadoh, S., Hamada, M., Hotta, K., Kudo, T., Seino, A., Vobis, G., & Yokota, A., editors. The Society for Actinomycetes Japan.
- Lai, N.-S. 2003. Isolation of actinomycetes from Sabah and the screening for inhibitors against eukaryotic signal transduction. Master of Science. Universiti Malaysia Sabah, Kota Kinabalu.
- Lazzarini, A., Cavaletti, L., Toppo, G. & Marinelli, F. 2000. Rare genera of actinomycetes as potential producers of new antibiotics. *Antonie van Leeuwenhoek*. **78**:399-405.
- Lechevalier, H. A. & Lechevalier, M. P. 1981. Introduction to the Order Actinomycetales. In The Prokaryotes. Vol. 2. Starr, M.P., Stolp, H., Truper, H.G., Balows, A., & Schlegel, H.G., editors. Springer-Verlag, New York. 1915-1922.
- Lee, J. T., Jr. & McCubrey, J. A. 2002. The Raf/MEK/ERK signal transduction cascade as a target for chemotherapeutic intervention in leukemia. *Leukemia*. **16**:486-507.
- Lee, S. D., Kang, S.-O. & Hah, Y. C. 2000. Hongia gen. nov., a new genus of the order Actinomycetales. *International Journal of Systematic and Evolutionary Microbiology*. **50**:191-199.
- Louvion, J.-F., Abbas-Terki, T. & Picard, D. 1998. Hsp90 is required for pheromone signaling in yeast. *Molecular Biology of the Cell*. **9**:3071-3083.



- Lyons, J. F., Wilhelm, S., Hibner, B. & Bollag, G. 2001. Discovery of a novel Raf kinase inhibitor. *Endocrine Related Cancer*. **8**:219-225.
- Malumbres, M. & Barbacid, M. 2003. RAS oncogenes: the first 30 years. *Nature Reviews Cancer*. **3**:459-465.
- Marcu, M. G., Chadli, A., Bouhouche, I., Catelli, M. & Neckers, L. M. 2000. The heat shock protein 90 antagonist novobiocin interacts with a previously unrecognized ATP-binding domain in the carboxyl terminus of the chaperone. *The Journal of Biological Chemistry*. **275**:37181-37186.
- Mason, C. S., Springer, C. J., Cooper, R. G., Superti-Furga, G., Marshall, C. J. & Marais, R. 1999. Serine and tyrosine phosphorylations cooperate in Raf-1, but not B-Raf activation. *European Molecular Biology Organization Journal*. **18**:2137-2148.
- McKinney, J. D., zu Bentrup, K. H., Munoz-Elias, E. J., Miczak, A., Chen, B., Chan, W. T., Swenson, D., Sacchettini, J. C., Jacobs Jr, W. R. & Russell, D. G. 2000. Persistence of *Mycobacterium tuberculosis* in macrophages and mice requires the glyoxylate shunt enzyme isocitrate lyase. *Nature*. **406**:735-738.
- Midgley, R. S. & Kerr, D. J. 2002. Ras as a target in cancer therapy. *Critical Reviews in Oncology/Hematology*. **44**:109-120.
- Mincer, T. J., Jensen, P. R., Kauffman, C. A. & Fenical, W. 2002. Widespread and persistent populations of a major new marine actinomycete taxon in ocean sediments. *Applied and Environmental Microbiology*. **68**:5005-5011.
- Miyata, Y. 2005. Hsp90 inhibitor geldanamycin and its derivatives as novel cancer chemotherapeutic agents. *Current Pharmaceutical Design*. **11**:1131-1138.
- Miyata, Y. & Nishida, E. 2004. CK2 controls multiple protein kinases by phosphorylating a kinase-targeting molecular chaperone, Cdc37. *Molecular and Cellular Biology*. **24**:4065-4074.
- Morrison, D. K., Heidecker, G., Rapp, U. R. & Copeland, T. D. 1993. Identification of the major phosphorylation sites of the Raf-1 kinase. *The Journal of Biological Chemistry*. **268**:17309-17316.
- Morrison, D. K., Kaplan, D. R., Rapp, U. & Roberts, T. M. 1988. Signal transduction from membrane to cytoplasm: growth factors and membrane-bound oncogene products increase Raf-1 phosphorylation and associated protein kinase activity. *Proceedings of the National Academy of Sciences*. **85**:8855-8859.
- Nollen, E. A. A. & Morimoto, R. I. 2002. Chaperoning signaling pathways: molecular chaperones as stress-sensing 'heat shock' proteins. *Journal of Cell Science*. **115**:2809-2816.
- Nonomura, H. & Hayakawa, M. 1988. New methods for the selective isolation of soil actinomycetes. In *Biology of actinomycetes '88*. Okami, Y., Beppu, T., & Ogawara, H., editors. Japan Scientific Societies Press, Tokyo. 288-293.



- Ohta, E., Ohta, S., Kubota, N. K., Suzuki, M., Ogawa, T., Yamasaki, A. & Ikegami, S. 2001. Micromonosporide A, a new macrolide from *Micromonospora* sp. *Tetrahedron Letters*. **42**:4179-4181.
- Omholt, K., Karsberg, S., Platz, A., Kanter, L., Ringborg, U. & Hansson, J. 2002. Screening of N-ras codon 61 mutations in paired primary and metastatic cutaneous melanomas: mutations occur early and persist throughout tumor progression. *Clinical Cancer Research*. **8**:3468-3474.
- Omura, S. 1992. Trends in the search for bioactive microbial metabolites. *Journal of Industrial Microbiology*. **10**:135-156.
- Oneyama, C., Agatsuma, T., Kanda, Y., Nakano, H., Sharma, S. V., Nakano, S., Narazaki, F. & Tatsuta, K. 2003. Synthetic Inhibitors of Proline-Rich Ligand-Mediated Protein-Protein Interaction: Potent Analogs of UCS15A. *Chemistry & Biology*. **10**:443-451.
- Ong, S. M. 2005. Screening and characterization of microbial inhibitors against eukaryotic protein phosphatases (PP1 and PP2A). Master of Science. Universiti Malaysia Sabah, Kota Kinabalu.
- Ory, S., Zhou, M., Conrads, T. P., Veenstra, T. D. & Morrison, D. K. 2003. Protein phosphatase 2A positively regulates Ras signaling by dephosphorylating KSR1 and Raf-1 on critical 14-3-3 binding sites. *Current Biology*. **13**:1356-1364.
- Parish, T., Smith, D. A., Kendall, S., Casali, N., Bancroft, G. J. & Stoker, N. G. 2003. Deletion of two-component regulatory systems increases the virulence of *Mycobacterium tuberculosis*. *Infection and Immunity*. **71**:1134-1140.
- Pearson, G., Robinson, F., Beers Gibson, T., Xu, B.-e., Karandikar, M., Berman, K. & Cobb, M. H. 2001. Mitogen-activated protein (MAP) kinase pathways: regulation and physiological functions. *Endocrine Reviews*. **22**:153-183.
- Perez, E., Samper, S., Bordas, Y., Guilhot, C., Gicquel, B. & Martin, C. 2001. An essential role for phoP in *Mycobacterium tuberculosis* virulence. *Molecular Microbiology*. **41**:179-187.
- Prior, I. & Hancock, J. 2001. Compartmentalization of Ras proteins. *Journal of Cell Science*. **114**:1603-1608.
- Raabe, T. & Rapp, U. R. 2003. Ras signaling: PP2A puts Ksr and Raf in the right place. *Current Biology*. **13**:635-637.
- Schneider, T., Somogyi, J., Csermely, P. & Szamel, M. 2000. The Hsp90-specific inhibitor geldanamycin selectively disrupts kinase-mediated signaling events of T-lymphocyte activation. *Cell Stress and Chaperones*. **5**:52-61.
- Schulte, T. W., Akinaga, S., Murakata, T., Agatsuma, T., Sugimoto, S., Nakano, H., Lee, Y. S., Simen, B. B., Argon, Y., Felts, S., Toft, D. O., Neckers, L. M. & Sharma, S. V. 1999. Interaction of radicicol with members of the heat shock protein 90 family of molecular chaperones. *Molecular Endocrinology*. **13**:1435-1448.





- Vobis, G. 1997. Morphology of actinomycetes. In Atlas of actinomycetes. Miyadoh, S., Hamada, M., Hotta, K., Kudo, T., Seino, A., Vobis, G., & Yokota, A., editors. The Society for Actinomycetes Japan. 180-191.
- Vobis, G. 1999. The Genus *Actinoplanes* and Related Genera. <http://link.springer-ny.com/link/service/books/10125/>
- von Lintig, F. C., Huvar, I., Law, P., Diccianni, M. B., Yu, A. L. & Boss, G. R. 2000. Ras activation in normal white blood cells and childhood acute lymphoblastic leukemia. *Clinical Cancer Research*. **6**:1804-1810.
- Voo, L.-Y. C. 2004. Isolation of fungi from Sabah that produce bioactive compounds which affect signal transduction. Master of Science. Universiti Malaysia Sabah, Kota Kinabalu.
- Wakabayashi, T., Kageyama-Kawase, R., Naruse, N., Funahashi, Y. & Yoshimatsu, K. 2000. Luminacins: A family of capillary tube formation inhibitors from *Streptomyces* sp. II. Biological activities. *Journal of Antibiotics*. **53**:591-596.
- Walburger, A., Koul, A., Ferrari, G., Nguyen, L., Prescianotto-Baschong, C., Huygen, K., Klebl, B., Thompson, C., Bacher, G. & Pieters, J. 2004. Protein kinase G from pathogenic mycobacteria promotes survival within macrophages. *Science*. **304**:1800-1804.
- Wang, Y., Zhang, Z. S., Ruan, J. S., Wang, Y. M. & Ali, S. M. 1999. Investigation of actinomycete diversity in the tropical rainforests of Singapore. *Journal of Industrial Microbiology and Biotechnology*. **23**:178-187.
- Weber, T., Welzel, K., Pelzer, S., Vente, A. & Wohlleben, W. 2003. Exploiting the genetic potential of polyketide producing streptomycetes. *Journal of Biotechnology*. **106**:221-232.
- Whitesell, L., Mimnaugh, E., Costa, B., Myers, C. & Neckers, L. 1994. Inhibition of heat shock protein HSP90-pp60v-src heteroprotein complex formation by benzoquinone ansamycins: essential role for stress proteins in oncogenic transformation. *Proceedings of the National Academy of Sciences*. **91**:8324-8328.
- WHO. 2005. Global tuberculosis control: surveillance, planning, financing.
- Winkler, D. G., Cutler, R. E., Jr., Drugan, J. K., Campbell, S., Morrison, D. K. & Cooper, J. A. 1998. Identification of residues in the cysteine-rich domain of Raf-1 that control Ras binding and Raf-1 activity. *The Journal of Biological Chemistry*. **273**:21578-21584.
- Wu, X., Noh, S. J., Zhou, G., Dixon, J. E. & Guan, K.-L. 1996. Selective activation of MEK1 but not MEK2 by A-Raf from epidermal growth factor-stimulated Hela cells. *The Journal of Biological Chemistry*. **271**:3265-3271.
- Xu, Y. & Lindquist, S. 1993. Heat-shock protein hsp90 governs the activity of pp60v-src kinase. *Proceedings of the National Academy of Sciences*. **90**:7074-7078.



- Yokota, A. 1997a. Family *Micromonosporaceae*. In Atlas of actinomycetes. Miyadoh, S., Hamada, M., Hotta, K., Kudo, T., Seino, A., Vobis, G., & Yokota, A., editors. The Society for Actinomycetes Japan.
- Yokota, A. 1997b. Phylogenetic relationship of actinomycetes. In Atlas of actinomycetes. Miyadoh, S., Hamada, M., Hotta, K., Kudo, T., Seino, A., Vobis, G., & Yokota, A., editors. The Society for Actinomycetes Japan.
- Zheng, C. & Guan, K. 1993a. Cloning and characterization of two distinct human extracellular signal-regulated kinase activator kinases, MEK1 and MEK2. *The Journal of Biological Chemistry*. **268**:11435-11439.
- Zheng, C. F. & Guan, K. L. 1993b. Properties of MEKs, the kinases that phosphorylate and activate the extracellular signal-regulated kinases. *The Journal of Biological Chemistry*. **268**:23933-9.
- Zheng, C. F. & Guan, K. L. 1994. Activation of MEK family kinases requires phosphorylation of two conserved Ser/Thr residues. *European Molecular Biology Organization Journal*. **13**:1123-1131.
- Zhou, J. M., Zhu, X. F., Pan, Q. C., Liao, D. F., Li, Z. M. & Liu, Z. C. 2003. Manumycin inhibits cell proliferation and the Ras signal transduction pathway in human hepatocellular carcinoma cells. *International Journal of Molecular Medicine*. **11**:767-771.
- Zhu, J., Balan, V., Bronisz, A., Balan, K., Sun, H., Leicht, D. T., Luo, Z., Qin, J., Avruch, J. & Tzivion, G. 2005. Identification of Raf-1 S471 as a novel phosphorylation site critical for Raf-1 and B-Raf kinase activities and for MEK binding. *Molecular and Cellular Biology*. **16**:4733-44.

