

**ISOLATION, IDENTIFICATION AND
CHARACTERIZATION OF MICROORGANISMS
ASSOCIATED WITH *BUDU* FERMENTATION**

SIM KHENG YUEN



PERPUSTAKAAN
UNIVERSITI MALAYSIA SABAH

UMS

**THESIS SUBMITTED IN FULFILLMENT FOR
THE DEGREE OF MASTER OF SCIENCE**

**BIOTECHNOLOGY RESEARCH INSTITUTE
UNIVERSITI MALAYSIA SABAH
2010**

UNIVERSITI MALAYSIA SABAH

BORANG PENGESAHAN STATUS TESIS

JUDUL: ISOLATION, IDENTIFICATION AND CHARACTERIZATION OF
MICROORGANISMS ASSOCIATED WITH *BUDU* FERMENTATION

IJAZAH: SARJANA SAINS (BIOTEKNOLOGI MAKANAN)

SESI PENGAJIAN: 2005-2010


Saya, Sim Kheng Yuen mengaku membenarkan tesis sarjana ini disimpan di perpustakaan Universiti Malaysia Sabah dengan syarat-syarat kegunaan seperti berikut:

- 1 Tesis adalah hak milik Universiti Malaysia Sabah.
- 2 Perpustakaan Universiti Malaysia Sabah dibenarkan membuat salinan untuk tujuan pengajian sahaja.
- 3 Perpustakaan dibenarkan membuat salinan tesis ini sebagai bahan pertukaran Antara institusi pengajian tinggi
- 4 TIDAK TERHAD



Penulis: SIM KHENG YUEN

Disahkan oleh

 ANITA BINTI ARSAD
PUSTAKAWAN KANAN
UNIVERSITI MALAYSIA SABAH

TANDATANGAN PUSTAKAWAN



Penyelia: Prof. Madya Dr. Chye Fook Yee



Penyelia bersama:

Prof. Datin Seri Panglima Dr Ann Anton

Tarikh: 2010

CERTIFICATION

NAME : SIM KHENG YUEN
MATRIC NO : PS05-013-003
TITLE : ISOLATION, IDENTIFICATION AND
CHARACTERIZATION OF MICROORGANISMS
ASSOCIATED WITH *BUDU* FERMENTATION
DEGREE : MASTER OF SCIENCE
VIVA DATE : 25 NOVEMBER 2010

DECLARED BY

Signature

1. **SUPERVISOR**
(ASSOC. PROF. DR CHYE FOOK YEE)



2. **CO-SUPERVISOR**
(PROF. DATIN SERI PANGLIMA DR ANN ANTON)



UMS
UNIVERSITI MALAYSIA SABAH

DECLARATION

I hereby declare that the material in this thesis is my own except for quotations, excerpts, equations, summaries and references, which have been duly acknowledged.

November 2010



Sim Kheng Yuen
PS05-013-003



UMS
UNIVERSITI MALAYSIA SABAH

ACKNOWLEDGEMENTS

I wish to express my deepest gratitude and appreciation to my supervisors, Associate Professor Dr Chye Fook Yee of the School of Food Science and Nutrition, Universiti Malaysia Sabah, and Professor Datin Seri Panglima Dr Ann Anton of the Biotechnology Research Institute, Universiti Malaysia Sabah who have been patient enough to advice, guide and supervise me throughout the past few years. Their continuous encouragement provided me the necessary impetus to complete the research and publish this thesis.

I would like to thank to Mr. Taipin, the lab assistant of the Food Microbiology lab who assisted me in order to complete my study. A special thank to Ms. Shanty who arranged for any field trip during the study is much appreciated. They had showed their patience and guidance me throughout the study. Without their assistance, this study can't be done smoothly.

Meanwhile, I wish to thank for the Ministry of Science, Technology and Innovation (MOSTI) for sponsoring this study under the IRPA grant (NBD TOPDOWN IRPA01-02-10-002 BTK/ER/39). With the kind financial support by the MOSTI, many significant findings were discovered and benefit respective community including industry.

Last but not least, I would like to express my gratitude to all of the *budu* producers located in Kelantan (Mr. Ismail, Mr. Roslee and workers) who kindly provided me *budu* samples for my research study.

ABSTRACT

ISOLATION, IDENTIFICATION AND CHARACTERIZATION OF MICROORGANISMS INVOLVED IN *BUDU* FERMENTATION

This study was carried out to determine the chemical and microbiological changes during *budu* fermentation (A1, A2 and A3 located in Tumpat, Kelantan). Samples from different producers were allowed to ferment at room temperature similar to that applied by producers. Samples were taken on monthly basis for the chemical (pH, acidity, total soluble solid, salt and soluble protein content), proximate compositions (moisture, crude fat, crude protein and fibre content) and microbiological analyses. Changes in microbial flora such as halophilic, proteolytic, lactic acid bacteria (LAB), yeasts and enterobacteriaceae were monitored and all the isolates were phenotypically identified by Biolog Microlog Database System before further characterized for hydrolytic (pectinolytic, lipolytic, proteolytic and amylolytic), enzymatic activity and probiotic properties. The moisture, protein and ash content of all samples were increased, while the fat content decreased significantly ($p < 0.05$) during fermentation. Sample from producer A2 (Orkid) recorded the highest increase in total soluble protein, from the initial of 10.92 ± 0.30 to 40.72 ± 0.02 mg/ml after 12 months of fermentation. However, sample of producer A3 (Roslee) exhibited greatest decrease in fat content compared to other samples. The initial microbial load for all samples decreased significantly ($p < 0.05$) during fermentation. The total proteolytic count for producer A1 (Ketereh) and A2 (Orkid) increased in the first few months before decrease at the end of fermentation. A total of 150 isolates were identified, with majority are bacteria (77%), followed by yeasts (12%) and 11% of unconfirmed identity. Only two species of *Micrococcus*, namely *Micrococcus luteus* and *Micrococcus luteus* ATCC 9341, while four species of *Staphylococcus* were identified as *Staphylococcus arlettae*, *Staphylococcus cohnii*, *Staphylococcus carnosus* and *Staphylococcus xylosus* were identified. *Saccharomyces cerevisiae* and *Candida famata* were the major yeast species in all *budu* samples. The *M. luteus* ML2 was predominant strain to initiate *budu* fermentation before *Staphylococcus arlettae* SA strain took over the role. API ZYM test revealed that the *S. arlettae* SA1 and *L. plantarum* LP1 and LP2 were only strains to have strong lipolytic and proteolytic activity that associated with *budu* fermentation. However, none of the tested strains showed pectinolytic activity. *Lactobacillus plantarum* LP1 and LP2, *Staphylococcus arlettae* SA, *Saccharomyces cerevisiae* SC3, *Candida glabrata* CG2 strains were identified as potential probiotics as they were tolerant to acid and bile salt as well as exhibited antimicrobial activity against selected foodborne pathogens. In conclusion, a consortium of microorganisms was involved in *budu* fermentation and led to the changes of sensory attributes of the *budu* during fermentation. Further studies are necessary to evaluate the feasibility of the selected strains as starter cultures in pilot processing for controllable *budu* fermentation.

ABSTRAK

Kajian ini dibuat bagi mengkaji perubahan kimia dan mikrobiologi semasa berlangsungnya fermentasi budu. Sampel budu dari pengusaha A1 (Ketereh), A2 (Orkid) dan A3 (Roslee) terletak di sekitar daerah Tumpat, Kelantan telah diperam dalam suhu persekitaran seumpama dengan tempat pemprosesan asal. Analisis bulanan terhadap kandungan kimia (pH, keasidan, jumlah bahan terlarut, garam dan protin terlarut), proksimat (kelembapan, lemak, protin dan serat) serta mikrobiologi bagi sampel berlainan pengusaha telah dilakukan sepanjang fermentasi budu. Perubahan flora mikroorganisma jenis halofilik, proteolitik, laktik asid bakteria, kulat dan enterobakteria telah dikaji sepanjang fermentasi. Mikroorganisma yang terpicil telah dikenalpasti berdasarkan sistem pangkalan data Biolog Microlog dan pencirian lanjutan terhadap mikroflora tersebut turut dilakukan berdasarkan ciri-ciri hidrolitik (pektinolitik, lipolitik, proteolitik, amylolitik), aktiviti enzim serta ciri-ciri probiotik. Kandungan kelembapan, protin dan abu bagi semua sampel telah meningkat secara signifikan ($p < 0.05$), manakala kandungan lemak mencatat penurunan signifikan ($p < 0.05$) di sepanjang fermentasi budu. Sampel A2 mencatat peningkatan tertinggi bagi kandungan protin terlarut berbanding sampel yang lain, iaitu dari 10.92 ± 0.30 kepada 40.72 ± 0.02 mg/ml selepas 12 bulan, manakala sampel A3 mencatat penurunan kandungan lemak yang terbanyak. Jumlah kiraan mikroorganisma bagi sampel budu dari semua pengusaha menurun secara mendadak ($p < 0.05$) sepanjang fermentasi. Namun, jumlah kiraan proteolitik bagi sampel dari pengusaha A1 dan A2 meningkat pada awal fermentasi sebelum ia menurun secara mendadak pada akhir fermentasi. Sejumlah 150 picilan telah dikenalpasti dan majoritinya adalah bakteria (77%), diikuti oleh kulat (12%) dan sebanyak 11% isolat gagal dikenalpasti. Hanya dua spesis *Micrococcus* (*Micrococcus luteus*, *Micrococcus luteus* ATCC 9341) dan empat spesis *Staphylococcus* (*Staphylococcus arlettae*, *Staphylococcus cohnii*, *Staphylococcus carnosus* dan *Staphylococcus xylosum*) telah dikenalpasti dalam kajian ini. Sehubungan itu, *Saccharomyces cerevisiae* dan *Candida famata* merupakan spesis yis yang kerap muncul dalam semua sampel yang dikaji. Keputusan juga menunjukkan bahawa *Micrococcus luteus* ML2 merupakan strain yang memulakan proses fermentasi sebelum diganti oleh *Staphylococcus arlettae* SA bagi melanjutkan fermentasi budu pada peringkat akhir. Keputusan API ZYM menunjukkan bahawa *S. arlettae* SA dan *L. plantarum* LP1 dan LP2 mempunyai aktiviti lipolitik dan proteolitik yang kuat lalu memainkan peranan dalam fermentasi budu. Namun begitu, tiada sebarang strain yang menunjukkan aktiviti pektinolitik. Cuma *Lactobacillus plantarum* LP1, LP2, *Staphylococcus arlettae* SA, *Saccharomyces cerevisiae* SC3 dan *Candida glabrata* CG2 berpotensi sebagai strain probiotik memandangkan mereka adalah rintang kepada asid, garam hempedu serta menunjukkan aktiviti antimikrobial terhadap beberapa patogen bawaan makanan yang terpilih dalam kajian ini. Kesimpulannya, mikroorganisma yang dikenalpasti dalam kajian ini terlibat dalam penguraian budu yang membawa kepada perubahan sepanjang fermentasi berlangsung.

TABLE OF CONTENTS

Page

TITLE	i
DECLARATION	ii
CERTIFICATION	iii
ACKNOWLEDGEMENT	iv
ABSTRACT	v
<i>ABSTRAK</i>	vi
TABLE OF CONTENTS	vii
LIST OF TABLES	xi
LIST OF FIGURES	xiii
LIST OF APPENDIX	xiv
CHAPTER 1: INTRODUCTION	1
CHAPTER 2: LITERATURE REVIEW	5
2.1 The <i>Budu</i> Industry	5
2.1.1 Small and Medium Enterprises	5
2.1.2 <i>Budu</i> Industry in Malaysia	5
2.2 Fermentation	6
2.2.1 Industrial Fermentation	7
a. Production of Organic Acid	8
b. Enzyme Production	9
c. Production of Bioactive Products	12
d. Production of Biofuels	18
2.2.2 Food Fermentation	19
a. Classification of Fermented Food	23
b. Traditional Fermented Foods	24
c. Indigenous Fermented Foods in Malaysia	25
2.2.3 Fish Sauce	26

a.	Fish Sauce Manufacturing	27
b.	Chemical and Biochemical Composition of Fish Sauce	30
c.	Microbiology of Fish Sauce	33
2.2.4	<i>Budu</i> Fermentation	34
a.	Traditional <i>budu</i> Processing in Malaysia	35
b.	Parameters Affecting Quality of <i>budu</i>	36
2.3	Microbial Interaction in Fermented Foods	38
2.3.1	Bacterial and Yeast Interaction	39
2.3.2	Yeast and Yeast Interaction	41
2.3.3	Yeast and Filamentous Fungi Interaction	43
2.4	Safety Issues of Fermented Foods	44
2.4.1	Presence of Biogenic Amines in Fermented Foods	44
2.4.2	Presence of Food Poisoning Microorganisms in Fermented Foods	47
2.5	Biotechnology Application in Food Fermentation	49
2.5.1	Biotechnological Importance of Microorganisms	50
a.	Production of Nutraceuticals	51
b.	Production of Food Additives	53
2.5.2	Development of Starter Culture Technology	56
a.	Spontaneous Fermentation and Back Slopping Technique in Food Fermentation	56
b.	Approaches and Applications of Starter Cultures in Small-Scale Fermentation	58
c.	Selection Criteria for Starter Cultures Development	59
d.	Disadvantages of Using Starter Cultures	61
CHAPTER 3: MATERIALS AND METHODS		63
3.1	Sample Preparation	63
3.2	Proximate Analysis	64
3.2.1	Moisture and Ash Content	64
3.2.2	Crude Protein Content	64
3.2.3	Fat Content	65
3.2.4	Crude Fiber Content	65
3.2.5	Carbohydrate Content	66
3.3	Other Chemical Analysis	66
3.3.1	Total Soluble Solids	66
3.3.2	pH and Titratable Acid	66
3.3.3	Salt Content	66
3.3.4	Total Soluble Protein Content	67

3.4	Microbiological Analysis	67
3.4.1	Sample Preparation	67
3.4.2	Enumeration and Isolation	68
3.4.3	Phenotypic Characterization and Identification of Isolates	68
a.	Characterization of Non-LAB Microorganisms	69
b.	Characterization of LAB Microorganisms	69
c.	Characterization of Yeast	70
d.	Characterization of Mould	70
3.5	Identification of Microbial Groups Using the Biolog Microplates (GP2, GN2, AN, YT, FF)	70
3.6	Technological Characteristics of Isolate	71
3.6.1	Screening of hydrolytic activity	71
a.	Proteolytic Activity	71
b.	Lipolytic Activity	72
c.	Amylolytic Activity	72
d.	Pectinolytic Activity	72
3.6.2	Enzymatic Activities of Isolates By the API ZYM Method	73
3.6.3	Screening of Probiotic Activities	73
3.7	Statistical Analysis	74
CHAPTER 4 RESULT AND DISCUSSION		75
4.1	Chemical and Proximate Changes during <i>Budu</i> Fermentation	75
4.2	Microbiological Changes during <i>Budu</i> Fermentation	81
4.3	Phenotypic Characterization and Identification of Gram Positive and Catalase Positive Bacteria	86
4.4	Phenotypic Characterization and Identification of Gram Negative Bacteria	91
4.5	Phenotypic Characterization and Identification of LAB	92
4.6	Phenotypic Characterization and Identification of yeast	95
4.7	Cluster analysis of Identified strains	99
4.8	Microbial Succession Involved in Fermentation	103
4.9	Technological Properties of the Strains	114
4.9.1	Screening of hydrolytic activity	114
4.9.2	Screening of probiotic activity	125

CHAPTER 5 CONCLUSIONS AND FUTURE STUDY

133

REFERENCES

135

APPENDICES

175



UMS
UNIVERSITI MALAYSIA SABAH

LIST OF TABLES

		Page
Table 2.1	Production of enzymes through fermentation	11
Table 2.2	List of bioactive peptide released from milk proteins by various microorganisms and microbial enzymes	15
Table 2.3	Classification of fermented foods	22
Table 2.4	Classification of fermentation	23
Table 2.5	Example of selected fermented foods around the world	24
Table 2.6	Indigenous fermented foods in Malaysia	26
Table 2.7	Fish sauce processing methods and different species of fish used in various countries	28
Table 2.8	Amino acid composition (mg/100mL) of various fish sauce	32
Table 2.9	Occurrence of biogenic amines in several fermented foods	46
Table 2.10	Incident of reported foodborne disease outbreak associated with fermented products	48
Table 2.11	Research priorities required for development of starter cultures	59
Table 4.1	Chemical and proximate analysis of sample from producer A1 (Keterreh) during <i>budu</i> fermentation	78
Table 4.2	Chemical and proximate analysis of sample from producer A2 (Orkid) during <i>budu</i> fermentation	79
Table 4.3	Chemical and proximate analysis of sample from producer A3 (Roslee) during <i>budu</i> fermentation	80
Table 4.4	Microbial load of sample from producer A1 (Keterreh) during <i>budu</i> fermentation	81
Table 4.5	Microbial load of sample from producer A2 (Orkid) during <i>budu</i> fermentation	84
Table 4.6	Microbial load of sample from producer A3 (Roslee) during <i>budu</i> fermentation	85
Table 4.7	Physiological characteristics of Gram positive bacteria Isolated from <i>budu</i>	87

Table 4.8	Gram positive bacteria identified from <i>budu</i> samples	90
Table 4.9	Physiological characteristics of Gram negative bacteria Isolated from <i>budu</i>	91
Table 4.10	Identification of Gram negative bacteria during fermentation	92
Table 4.11	Physiological characteristics of LAB found in <i>budu</i> fermentation	93
Table 4.12	LAB identified	94
Table 4.13	Physiological and biochemical characteristics of yeasts found in <i>budu</i> fermentation	97
Table 4.14	Identification of yeast during <i>budu</i> fermentation	97
Table 4.15	Microbial composition involved in <i>budu</i> fermentation	105
Table 4.16	Distribution of identified microbial strains during <i>budu</i> Fermentation	108
Table 4.17	Hydrolytic activities of the tested strains	117
Table 4.18	Enzymatic activities (API ZYM) of the corresponding strains	122
Table 4.19	Screening on the probiotic properties of the investigated strains	128

LIST OF FIGURES

	Page
Figure 2.1: Schematic flow sheet for the conversion of biomass to ethanol	19
Figure 2.2: Production scheme of nampla (Thai fish sauce)	29
Figure 2.3: Schematic <i>budu</i> processing in Malaysia	36
Figure 4.1 Dendrogram of the Gram positive bacteria isolated from <i>budu</i> fermentation	100
Figure 4.2 Dendrogram of the LAB isolated from <i>budu</i> fermentation	101
Figure 4.3 Dendrogram of the yeasts isolated from <i>budu</i> fermentation	102



UMS
UNIVERSITI MALAYSIA SABAH

LIST OF APPENDIX

	Page
Appendix A: Traditional <i>budu</i> processing	175
Appendix B: Identification of isolates via Biolog Microlog System	176
Appendix C: Substrate utilization of <i>Micrococcus luteus</i> in Biolog GP2 plate	177
Appendix D: Substrate utilization of <i>M. luteus</i> ML1 in Biolog GP2 plate	178
Appendix E: Substrate utilization of <i>M. luteus</i> ML2 in Biolog GP2 plate	179
Appendix F: Substrate utilization of <i>M. luteus</i> ML3 in Biolog GP2 plate	180
Appendix G: Substrate utilization of <i>M. luteus</i> ML4 in Biolog GP2 plate	181
Appendix H: Substrate utilization of <i>M. luteus</i> ATCC9341 MLATCC in Biolog GP2 plate	182
Appendix I: Substrate utilization of <i>M. luteus</i> ATCC9341 MLATCC1 in Biolog GP2 plate	183
Appendix J: Substrate utilization of <i>M. luteus</i> ATCC9341 MLATCC2 in Biolog GP2 plate	184
Appendix K: Substrate utilization of <i>Staphylococcus arlettae</i> in Biolog GP2 plate	185
Appendix L: Substrate utilization of <i>S. carnosus</i> in Biolog GP2 plate	186
Appendix M: Substrate utilization of <i>S. carnosus</i> SCN1 in Biolog GP2 plate	187
Appendix N: Substrate utilization of <i>S. carnosus</i> SCN2 in Biolog GP2 plate	188
Appendix O: Substrate utilization of <i>S. carnosus</i> SCN3 in Biolog GP2	189
Appendix P: Substrate utilization of <i>S. cohnii</i> in Biolog GP2 plate	190
Appendix Q: Substrate utilization of <i>S. cohnii</i> SCH1 in Biolog GP2	191

Appendix R:	Substrate utilization of <i>S. cohnii</i> SCH2 in Biolog GP2 plate	192
Appendix S:	Substrate utilization of <i>S. cohnii</i> SCH3 in Biolog GP2 plate	193
Appendix T:	Substrate utilization of <i>S. xylosus</i> in Biolog GP2 plate	194
Appendix U:	Substrate utilization of <i>S. xylosus</i> SX1 in Biolog GP2 plate	195
Appendix V:	Substrate utilization of <i>S. xylosus</i> SX2 in Biolog GP2 plate	196
Appendix W:	Substrate utilization of <i>Corynebacterium afermentans ss lipophilum</i> in Biolog GP2 plate	197
Appendix X:	Substrate utilization of <i>Rahnella aquatilis</i> in Biolog GN2	198
Appendix Y:	Substrate utilization of <i>Enterobacter agglomerans</i> bgp7 in Biolog GN2 plate	199
Appendix Z:	Biochemical and technological characterization of isolates	200



UMS
UNIVERSITI MALAYSIA SABAH

CHAPTER 1

INTRODUCTION

Food fermentation is one of the oldest forms of food biotechnology which provide 20-40% of our food supply. Typically, about one third of our food intakes are comprised of fermented foods such as yoghurts, cheese, yoghurt drinks, soy and fish sauce, soymilk, wines, breads, fermented sausages or related meat products and fermented soy bean products. Fermented foods that derived from plant or animal materials are usually acceptable and become essential part of the diet in most parts of the world. This is because a wide variety of raw materials are used and the technology of producing it basically ranging from the primitive to the most advanced in order to achieve an outstanding range of sensory and textural qualities of the final products (Paul-Ross *et al.*, 2002).

In some developing countries, food fermentation is still carried out as household or cottage industry level without any application of modern and scientific principle. The process normally initiated spontaneously and thus, retains the unique flavour and aroma characteristics of the end products compared to those fermented foods produced by controlled fermentation. However, the microbiological and biochemical aspects of this traditional process are complicated and not fully elucidated. In general, the process is carried out by means of minimum technological know-how with a low yields, and with variable quality, as in the production of *Hussuwa*, a traditional African fermented sorghum food (Yousif *et al.*, 2010). Furthermore, the poor hygienic conditions and improper handling during fermentation render the products susceptible to contamination. The inadequate knowledge on proper packaging or post fermentation treatments denies the fate of end product quality and eventually limits their acceptance by consumers. On contrary, controlled fermentation involving the use of starter cultures is already becoming a trend for most fermentation processes. It is believed that a large production scale can be met through the use of starter

cultures. Hence, the safety and quality control of fermented products are also well promised (Holzapfel, 1997).

The main reason for food fermentation can be ascribed to preservation. In addition, the nutritional properties of the fermented products can be improved through fermentation. In most developing countries, malnutrition become a major problems as the raw materials are so limited (Paredes-Lopez and Harry, 1988). Therefore, people from those regions depend on fermented vegetal starches or fermented root crops such as cassava to sustain their daily diets. Fermentation of a substrate have been diversified to the production of secondary metabolites such as biocatalysts (Yang *et al.*, 2005), starter cultures (Hugas and Monfort, 1997; Papamaloni *et al.*, 2002) and potential pharmaceutical metabolites (Aymerich *et al.*, 2000; Messi *et al.*, 2001). This indicates that the understanding of the fermentation process not only confined to the production of fermented products, but it also significantly contributes to the scientific knowledge and economic of human.

Indigenous fermented foods in Malaysia are mostly produced in small and backyard scales. The know-how in producing fermented foods is delivered from generation to generation within the locality and only small quantity of the product is distributed beyond a geographical area. Microbiological and biochemical changes during fermentation are partially understood, therefore the end products will always have low product yields with variable quality (Rolle and Satin, 2002). The processes is normally resulting from the competitive activities of a variety of contaminating microorganisms and those best adapted to the food substrate and to technical control parameters, eventually dominate the process (Holzapfel, 2002). This shows that there is a possibility the local producers may use the back-slopping technique to produce fermented foods. However, the fermentation is still confined to small-scale production due to inadequate financial supports as well as scientific understanding of the process.

Fish sauce is one of the fermented products that prepared by adding certain amount of salt to the fish species and allowed to ferment for certain periods of time. The final product will be in solubilized liquid forms due to

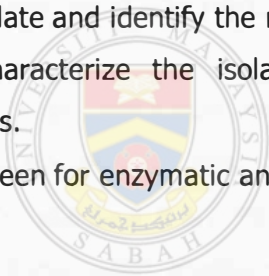
hydrolysis that aided by microorganisms. During fermentation, microbial succession encourages complete solubilization of the fish protein into free amino acids and peptides for the development of its unique characteristic. Many different species of microorganisms are isolated from fish sauce produced in various regions including nampla (Tanasupawat and Komagata, 2001), shotturu (Mura *et al.* 2000) and bakasang (Ijong and Ohta 1996). An understanding on the types of dominant microorganisms involved in fish fermentation is vital. According to Lopetcharat *et al.* (2001), the dominant species of microorganisms that have been isolated in fish sauce usually are those produce proteolytic enzymes and tolerate to high salt content, such as *Bacillus*, *Pseudomonas*, *Micrococcus*, *Staphylococcus*, *Halobacterium* and *Halococcus sp.* In another study by Lopetcharat and Park (2002), they found that *Staphylococcus*, *Bacillus* and *Micrococcus* were the predominant bacteria involved in fish sauce made from pacific whiting (*Merluccius productus*).

The complexity of the fish sauce fermentation is a challenge to scientists who interested to study the traditional process with the aim to improve the fish sauce quality through controlled fermentation process. Fukami *et al.* (2004b) found that the genotypic characterized through the DNA-DNA hybridization analysis of *Staphylococcus nepalensis* can actually be employed to improve the unpleasant odor of fish sauce. However, Jiang *et al.* (2007) claimed that yu-lu (Chinese fish sauce) is mixed cultures fermentation and the process favors the growth of halotolerant and halophiles only. The fermentation seems to increase the total soluble nitrogen, trichloroacetic acid (TCA) soluble peptides and free amino acids that eventually improve the nutritional value of the fermented product. Lipid oxidation that occurred during fermentation may cause bad taste and aroma to fish sauce that made from sardines (Kilinc *et al.*, 2005). According to Yongsawatdigul *et al.* (2007) even pointed out that the *nam-pla* (Thailand fish sauce) fermentation can be accelerated by using proteinases and bacterial starter cultures.

Budu, one of the Malaysian indigenous fermented food which is cloudy in color due to the sediment of fish bones and hydrolyzed fish meat, is popular in the east coast of peninsular Malaysia (Kelantan, Terengganu and Pahang). The

production of *budu* actually is a time consuming process as it required at least 8-12 months to ensure full solubilization of the fish mixture. There is less technological input in this traditional fermentation process as it is developed through trial and errors and the technique of production may depend on skill of producers. Thus, this uncontrolled process usually ends up with a small production scale and low quality products compared to fermented fishery products from neighboring countries like Thailand (nam-pla) and Vietnam (Nuoc-mam). In order to overcome inconsistent in the *budu* quality, the use of starter cultures derived from this spontaneous fermentation is appreciated. Even though *budu* has been consumed by local consumers due to its unique meaty flavours and aroma, but the microbial diversity as well as the chemical changes involved in *budu* fermentation are not well elucidated. Hence, this study is undertaken with the objectives:

1. To elucidate the microbiological and biochemical changes during fermentation.
2. To isolate and identify the microorganisms involved in *budu* fermentation.
3. To characterize the isolated microorganisms based on their biochemical profiles.
4. To screen for enzymatic and probiotic properties of isolates.



UNIVERSITI MALAYSIA SABAH

CHAPTER 2

LITERATURE REVIEW

2.1 The *Budu* Industry

2.1.1 Small-and Medium Scale Food Processing Enterprises (SMEs)

Small and medium-scale enterprises (SMEs) played an important role in the Malaysian economy for the past few decades. A total of 548,307 or 99.2% were defined as SMEs which can be further classified into service sector based enterprise (86.6%), managing sector (7.2%) and agriculture sector (6.2%). However, statistic released by Department of Statistic, Malaysia (2005) showed that the largest number of establishments of SMEs is the textile and apparel sector which accounts for 23.4% of the total manufacturing sector. This is followed by food and beverages (15.0%), metal and metal products (13.0%) and eventually wood and wood products (14.1%).

There are more than 9000 food processing factories in Malaysia of which 95% are classified as small scale with the annual sale turnover is between RM250, 000 and less than RM10 million with a full time employees between 5 and 50 persons; while medium scale enterprise is the one with the annual sale turnover between RM 10 and RM25 million with a full time employees between 51 and 150 person. According to Chee (1986), small scale food processing enterprises may evolve if the enterprises expand which will eventually lead to the formation of a limited company. Another criteria that distinguish the small scale food enterprises from the large scale enterprises is the organizational structure which operated by a manager-owner assisted by a few workers. The products are generally cheap and rather low quality and the marketing strategy is done directly or through agents.

2.1.2 *Budu* Industry in Malaysia

Budu is one of fermented fishery products which are quite common among the local people in the East Coast of Peninsular Malaysia (Kelantan, Terengganu and Pahang). The production scale is usually very small with a minimum quality control

throughout the process, less technological inputs are implemented and most importantly the method of producing is a heritage from one generation to the next. Hence, the *budu* industry is considered as a small scale processing enterprise due to limited annual sale turnover and products are sold within local markets only.

The *budu* manufacturers in Kelantan are usually operating in small scale production to meet local market demands. This industry is not well expanded due to low consumers' acceptance on the unique flavour and aroma of *budu* as well as its appearances. On the other hand, the emergence of imported clarified fish sauce products from neighboring countries which is more acceptable in sensory attributes or appearance also affect the growth of the local industry. Besides, *budu* industry is also facing several problems, such as inconsistency in price as some of the producer sell their product directly to local customers based on the landing price of anchovies in different seasons, for instance, the price of *budu* during monsoon season is high compared to other non-monsoon seasons. Due to low quality of the end products cause the *budu* incompatible with other imported fish sauces. Therefore, a strategic planning for improvement of *budu* production is needed in order to improve demand for the traditional fermented food.

2.2 Fermentation

Fermentation is defined as an energy yielding anaerobic metabolic breakdown of sugar containing compounds such as glucose into smaller compounds and eventually to yield desirable fermentation end products including acid, alcohol (ethanol) and other simple products without net oxidation. Fermentation can also be defined as a process of the bioconversion of organic substances by microorganisms and/or by enzymes of microbial, plant or animal origin into secondary metabolites. Fermentation is able to produce material in a way that would be difficult or very costly if ordinary chemical methods are chosen to synthesis it. For example, the production of enzymes that harvested from microorganisms through a solid state or submerged fermentation technology. Fermentation technology is currently applied in various fields such as food and

animal feed production, production of biopesticides, biofuel, pharmaceuticals and even waste treatments system.

Both submerged (SmF) and solid-state (SSF) fermentation have been applied in many fermentation process such as brewing, bread making industry, enzymes synthesis, biopulping industry, biodegradation of biomass from waste materials and production of pharmaceuticals or functional foods. Submerged (SmF) fermentation actually involves the cultivation of microorganisms in a liquid nutrient broth to produce expected end products, such as the production of enzymes (Ito *et al.*, 2001) and amino acids (Gomes and Kumar, 2005). However, solid-state (SSF) fermentation involving solids in absence (or near absence) of free water, and the substrate used must possess enough moisture to support the growth and metabolism of microorganisms (Pandey *et al.*, 2001). Therefore, in a SSF system, it actually stimulates the growth of microorganisms in nature on moist solids with a low energy requirement, produce lesser waste water and more environmentally friendly as they resolve the problem of solid waste disposal. Thus, there has been an increasing interest on SSF which has been applied in most bioprocesses such as bioleaching, biopulping, bioremediation and biobenefication (Classen *et al.*, 2000; Ogonna *et al.*, 2001; Han and Rombouts, 2001; Haddadin *et al.*, 2001)

2.2.1 Industrial Fermentation

Industrial fermentation has great impact on human beings as most of the important primary metabolites such as amino acids, nucleotides, vitamins, enzymes, solvents, organic acids and vaccines have been produced through various fermentation processes. Microorganisms are efficiently utilized and optimized for the production of the above metabolites via large-scale industrial fermentation technology (Suryanarayan, 2003; Anderson, 2009). There are some advantages to apply microbial fermentation technique for the above purposes as compared to chemically synthesized process. At first, high rates of metabolism and biosynthesis can be achieved due to the high ratio of surface area to volume which facilitates the uptake of nutrients. Secondly, a tremendous variety of reactions which microorganisms are capable of carrying out also show the importance of their existence in most fermentation process. Then, the ability of the

microorganisms to adapt well in different environments as well as their capability to grow on most inexpensive carbon and nitrogen sources actually lower down the production cost in most fermentation industry. Apart from that, the ability to undergo genetic manipulation on the microorganisms either in vivo or in vitro to increase the production of the required products again show that the feasibility and benefits of manipulating microorganisms in any fermentation processes (Demain, 2000).

Fermentation usually consider as a low cost but high yield biological process because the raw materials such as corn starch, sugar cane waste, molasses or other carbohydrate rich substrates are utilized efficiently by microorganisms in order to produce essential primary and secondary metabolites as compared to chemical synthesis process (Schuster *et al.*, 2002; Rodriguez-Couto, 2008). The application of microorganisms in fermentation industry can also be much appreciated by the fact that even simple molecules such as L-glutamic acid and L-lysine are produced through fermentation rather than by chemical synthesis. Apart from that, most natural products able to undergo fermentation process since they are very complex and contain many centers of asymmetry that will never be made commercially by chemical synthesis. Below are several end products that being produced through industrial fermentation technology.

a. Production of Organic Acids

The production of organic acids seems vital as there is a high demands in food, pharmaceutical, leather, textile and feed stocks industries. The great expanding of food industry also sees the increased demand for the organic acids especially lactic and citric acids as more than 50% lactic acid produced is used to emulsify bakery products (Litchfield, 1996). Then, lactic acid also used as acidulant / flavouring agent as well as inhibitors to bacterial spoilage in processed foods such as candy, breads and bakery products, soft drinks, soups, dairy products, beers and processed eggs. Furthermore lactic acid is also used in cosmetic formulations, oilments and lotions due to their high water retaining capacity.