# EFFECTS OF DIFFERENT FEEDS FOR THE SURVIVAL POTENTIAL ON EARLY LARVAL STAGE OF WHITE BLOTCH SNAPPER, LUTJANUS RIVULATUS

## **CHING FUI FUI**

PERPUSTAKAAN UNIVERSITI MALAYSIA SABAH

Sila dapatkan Video / CD / Kaset / State / 14926 di bahagian Media

## BORNEO MARINE RESEARCH INSTITUTE UNIVERSITI MALAYSIA SABAH



### UNIVERSITI MALAYSIA SABAH

### BORANG PENGESAHAN STATUS TESIS®

EFFECTS OF DIFFERENT FEEDS FOR THE SURVIVAL POTENTIAL JUDUL : ON EARLY LARVAL STAGE OF WHITE BLOTCH SNAPPER, LUTJANUS RIVULATUS

SARJANA SAINS (AKUAKULTUR) IJAZAH:

### SESI PENGAJIAN: 2004-2007

Saya, Ching Fui Fui mengaku membenarkan tesis sarjana ini disimpan di perpustakaan Universiti Malaysia Sabah dengan syarat-syarat kegunaan seperti berikut:

- Tesis adalah hak milik Universiti Malaysia Sabah. 1.
- Perpustakaan Universiti Malaysia Sabah dibenarkan membuat salinan untuk tujuan 2. pengajian saya.
- 3. Perpustakaan dibenarkan membuat salinan tesis ini sebagai bahan pertukaran antara institusi pengajian tinggi.

Disahkan oleh

TIDAK TERHAD. 4.

Penulis: CHIN G FUIT FUIT

PERPUSTAKAAN UNIVERSITI MALAYSIA SARA'TANDATANGAN PUSTAKAWAN

Alamat: House No. 2, Lorong 6-3A, Taman Bukit Sepanggar, 88450 Kota Kinabalu, Sabah.

> Penyelia: Prof. Dr. Shigeharu Senoo Tarikh: 10/7/2008

JAMIUN MICHEAL

PI

TAKAWAN

RPUSTAKAAN UNIVERSITI MALAYSIA SABAH

Tarikh: 2008

CATATAN: "Tesis dimaksudkan sebagai tesis Ijazah Doktor Falsafah dan Sarjana secara penyelidikan atau disertassi bagi pengajian secara kerja kursus dan penyelidikan, atau laporan Projek Sarjana Muda (LPSM).



## CERTIFICATION : CHING FUI FUI NAME MATRIC NO. : PS04-004-010 : EFFECTS OF DIFFERENT FEEDS FOR THE SURVIVAL TITLE POTENTIAL ON EARLY LARVAL STAGE OF WHITE BLOTCH SNAPPER, LUTJANUS RIVULATUS : MASTER OF SCIENCE DEGREE : 6 SEPTEMBER 2007 VIVA DATE **DECLARED BY** (Signature) SUPERVISOR 1. (Prof. Dr. Shigeharu Senoo)



### DECLARATION

The materials in this thesis are original except for quotations, excerpts, summaries and references, which have been duly acknowledged.

6 SEPTEMBER 2007

CHING PS004-040-010



I would like to express my sincere indebted acknowledgement to my supervisor, Prof. Dr. Shigeharu Senoo from Borneo Marine Research Institute, for his invaluable suggestions, guidance and encouragement throughout study period.

I thank Universiti Malaysia Sabah for the endless assistantship. I would like to express my gratitude to Prof. Saleem Mustafa, Director of Borneo Marine Research Institute, Prof. Dr. Ridzwan Abdul Rahman, Director of Centre for Research Management and Conference (Former Director of Borneo Marine Research Institute), Prof. Datin Dr. Maryati Mohamed, Dean of Centre for Postgraduate Studies and Prof. Datuk Dr. Mohd. Noh Dalimin, Vice-chancellor, Universiti Malaysia Sabah, for their encouragement and assistance within the completion of this thesis.

My sincere appreciation is also extended to Dr. Yoshizumi Nakagawa, postdoctoral researcher of Kinki University for supporting, giving meaningful comments and advice, encouragement and assistance to complete my study.

My special thank to National Environment Fellowship (NEF), Japan for granted me a NAGAO scholarship as a study financial aid. I extended my appreciation to UMS NAGAO committee for all the direct or indirect assistantship.

My heartfelt thanks to Dr. Shahbudin Saad, Dr. Sitti Raehana Muhd. Shaleh, Dr. Yukinori Mukai, Mr. Montinius Guntalib, Ms. Siti Badra, Mrs. Rosliah Lawrence, and all the staff of Borneo Marine Research Institute for their co-operation. I wish to heartily express my indebtedness to Mr. Norazmi Osman, Mr. Henry Lipan, Mr. Herman Musanna, Mr. Ha Hou Chew, Ms. Sow Sok Hui, Ms. Nguang Siew Ing, Ms. Mok Wen Jye, Ms. Chun Kia Huey, Ms. Esther Michelle Gunben, , and all the staffs of UMS Hatchery, Borneo Marine Research Institute for their assistance, and well-wishers for their valuable inputs directly or indirectly for the successful completion of the thesis.

Special note of thanks to my parents, Mr Michael Ching Eng Huat and Mdm. Melia @ Angelina Binti Molondoi for their love, patience, support, understanding and encouragement for me to complete my study. My indebted thank to my siblings for their endless support, love and care in completing my study. Last but not least, I would like to express my gratitude to my fiancé, Mr. Redzwan Japri and family for their love, encouragement, understanding and support to make my study possible



#### ABSTRACT

### EFFECTS OF DIFFERENT FEEDS ON THE SURVIVAL POTENTIAL OF EARLY LARVAL STAGE OF HOI TAI KAI, LUTJANUS RIVULATUS

The high mortality in the early larval stage of Hoi Tai Kai, Lutianus rivulatus had caused to the failure in seed production. Three factors were responsible for the mass mortality: Small body size at hatching (2.19 mm), small mouth size at first feeding (80 µm) and short nutrition transition period (18 hours). The difficulty of larvae to consume rotifer, Branchionus sp. had restricted the success first feeding. The available rotifer (150-230 um) is considered too large for the small mouth size larvae to consume. Five artificial feeds and two live feeds were tested as starter feed in this study: boiled eag volk (BE), cod oil juice (COJ), raw egg yolk (RE) and combination feeds of BE and COJ (BE+COJ), combination feeds of BE and RE (BE+RE), rotifer and natural plankton. The effect of these different feeds to the survival potential of larvae was indicated. Finding shows larvae were only able to consume artificial feed based on the gut content observation. BE was found able to contribute the highest survival rate (87.7±3.3) and larvae succeed to survive in 9 dAH. The factors cause BE to become the most effective starter feed for larvae. Several factors responsible for the success use of BE: consumable by larvae, offer wide range of size (10-180 µm), highly preferred to be consume, Chesson index a (0.99), short gut passage time at the fist feeding day (30 minutes), resembles an attractive feed for the visual feeder larvae to detect feed in the rearing water as well as able to being nutritious feed for larvae.

Keywords: *Lutjanus rivulatus*, first feeding, survival potential, selectivity of feed size, selectivity of feed type.



### ABSTRAK

Kadar kematian yang tinggi pada Hoi Tai Kai, Lutjanus rivulatus pada peringkat awal lavi telah membantutkan usaha pengeluaran benih ikan ini. Tiga faktor telah dikenalpasti menyebabkan kematian lavi: menetas pada saiz badan yang kecil (2.19 mm), saiz mulut yang kecil pada hari makan pertama (80 µm) dan masa penukaran nutrisi yang pendek (18 jam). Ketiga-tiga faktor ini telah menyebabkan kesukaran lavi.untuk makan pada hari makan pertama. Rotifer, (150-230 µm) makanan asas pada lavi adalah telalu besar untuk lavi memakannya. Justeru ini, lima jenis makanam tiruan: rebusan kuning telur ayam (BE), jus minyak kod (COJ), telur ayam mentah (RE), kombinasa BE dan COJ (BE+COJ) dan kombinasi BE dan RE (BE+RE) dan 2 jenis makanan hidup: rotifer, Brachionus sp. dan plankton telah diuji bagi mendapatkan makanan yang sesuai bagi lavi dan sekaligus mampu memberi kadar kehidupan yang tinggi. Hasil kajian menunjukan lavi berupaya makan semua makanan yang diuji kecuali makanan hidup. Ini dapat dibuktikan melalui kandungan perut lavi. BE merupakan makanan yang mampu memberi kesan kehidupan yang paling tinggi kepada lavi (87.7±3.3) berbanding lavi yang diberi makanan lain. Lavi ini berupaya untuk hidup selama 9 hari selepas menetas (dAH). BE dirumuskan sebagai makanan terbaik pada lavi dalam kajian ini berdasarkan kebolehannya untuk: hadir dalm pelbagai saiz (10-180 µm), menjadi makanan yang digemari, Chesson indeks a (0.99), masa lalu usus yang pendek (30 minit), mempunyai nilai nutrisi yang baik, dan juga merupakan makanan yang mempunyai ciri-ciri yang mampu menarik minat lavi.



### LIST OF CONTENTS

		PAGE
TITLE		1
DECLARATION		ii
ACKNOWLEDG	EMENTS	III
ABSTRAK		iv
ABSTRACT		V
LIST OF CONTE	INTS	vi
LIST OF TABLE	S	x
LIST OF FIGUR	ES	×ii
LIST OF PHOTO	DS	xiv
LIST OF ABBRE	EVIATIONS	xix
CHAPTER 1:	INTRODUCTION	1
1.1	Hoi Tai Kai, <i>Lutjanus rivulatus</i>	1
1.2	L. rivulatus Seed Production in UMS Hatchery	2
1.3	Objectives	5
CHAPTER 2:	LITERATURE REVIEW	7
2.1	Background of Hoi Tai Kai	7
2.2	Marine Fish Larvae Survival	8
2.3	Live and Artificial Feeds	10
2.4	First Feeding	13
2.5	The Nutritional Transition Period	16
2.6	Feed Size Selectivity	18
2.7	Feed Selectivity	19
2.8	Gut Passage Time	22
CHAPTER 3:	MATERIALS AND METHODS	24
3.1	Preparation of Different Feeds for the Larval Rearing	24
	3.1.1 Boiled Egg Yolk (BE)	26
	3.1.2 Cod Oil Juice (COI)	26



	3.1.3 Raw Egg Yolk (RE), Combination Feeds of BE and COJ (BE+COJ) and Combination Feeds of BE and	27
	RE (BE+RE) 3.1.4 Natural Plankton	28
	3.1.5 Rotifer, <i>Brachiounus</i> sp.	30
3.2	Preparation for the Larval Rearing System	32
	3.2.1 Broodfish Management and Egg Collection System	32
	3.2.2 Larval Rearing	34
	a. Tank Preparation	34
	b. Rearing Water and Aeration Preparation	34
	c. Daily Management	35
	i. Bottom Cleaning	35
	ii. Feeding	37
	iii. Water Quality	38
3.3	Observations of the Reared Larvae Fed with Different Feeds	38
	3.3.1 Survival Rate	38
	3.3.2 Appearance of Larval Gut Content	38
	3.3.3 Selectivity of Feed Size	39
	3.3.4 Selectivity of Feed Type (Chesson, 1978)	41
	3.3.5 Gut Passage Time	43
3.4	Nutritional Analysis of BE and COJ	43
3.5	Statistical Analysis	43
CHAPTER 4:	RESULTS	44
4.1	Survival Rate	44
4.2	Appearance of Larval Gut Content	48
4.3	Selectivity of Feed Size	53
4.4	Selectivity of Feed Type	55
4.5	Gut Passage Time	58
4.6	Feed Nutritional Values	62



CHAPTER 5:	DISCUSSION	64
5.1	Larval Survival	64
	5.1.1 Factors Contributed to the Highest Survival Rate of	
	L. rivulatus	64
		~
	a. Acceptable to be Consumed	64
	b. Ability to Offer Wide Range of Size	67 68
	<ul><li>c. Highly Preferred to be Consumed</li><li>d. Short Gut Passage time</li></ul>	69
	5.1.2 Factors Contributed to the Lowest Survival Rate of	
	L. rivulatus	71
	a. Starvation	71
	b. Delayed Feeding	72
c. Rotifer Size		73
d. Low Natural Plankton Density		74
	e. Difficult to Mass Produce	75
	f. Surface Death	76
	5.1.3 Factors Contributed to the Moderate Survival Rate of	
	L. rivulatus	76
	a. Less Preference	76
	b. Feed Characteristics	77
CHAPTER 6:	CONCLUSION	
6.1	Conclusion	78
6.2	Recommendation and Future Study	78
REFERENCE		80



viii

APPENDICES	A : Feed Characterization	94
	B : SPSS Statistic Analysis	96
	C: Raw Data of Survival Rate	101
	D : Water Quality	103
	E : Publication	105



### LIST OF TABLES

		PAGE
Table 1.1:	Names of Lutjanus rivulatus in different countries (Allen, 1985)	
		1
Table 2.1:	The nutritional composition of chicken egg on a dry basis (Adapted from Chow, 1980)	12
Table 2.2:	First feeding time for larvae of various marine fish species (Rao, 2003)	15
Table 2.3:	The nutrition transition period of several marine fish larvae (Kohno, 1998)	17
Table 3.1:	Nutritional content and composition of Seven Seas cod liver oil per 100mg (Seven Seas Pure Cod Liver Oil, England)	26
	per roomg (oeven beas rare est eiver on england)	20
Table 3.2:	Volume and density of feed given to L. rivulatus	37
Table 4.1:	The mean equilibrium spherical diameter (ESD) in the gut of <i>L.</i> <i>rivulatus</i> fed with boiled egg yolk (BE) on each gut passage time at different feeding day	61
Table 4.2:	The mean equilibrium spherical diameter (ESD) in the gut of <i>L.</i> <i>rivulatus</i> fed with cod oil juice (COJ) on each gut passage time at different feeding day	61
Table 4.3:	The free fatty acid, acid value and fatty acid compositions of boiled egg yolk (BE) and cod oil juice (COJ) as a feeds for <i>L</i> . <i>rivulatus</i>	
Table 5.1:	The Comparison between the recommended artificial feed	



characteristics for fish larvae (adapted by Southgate and Partridge, 1998) and boiled egg yolk (BE)



### LIST OF FIGURES

- Figure 4.1: The changes on survival rate of *Lutjanus rivulatus* larvae fed with different feeds from 1-9 dAH. A. boiled egg yolk (BE), B. cod oil juice (COJ), C.combination feeds of BE and COJ (BE+COJ), D. raw egg (RE), E. combination feeds of BE and RE (BE+RE), F. rotifer, *Brachionus* sp., G. natural plankton, H. no feed were given. All data are presented in percentage of (survival±SD)
- Figure 4.2: Patterns of survival rates in the early larval stage of *Lutjanus* rivulatus larvae fed with different feeds. A, The lowest survival rate showed by larvae fed with rotifer, natural plankton and no feed, B. the moderate survival rate showed by larvae fed with cod oil juice (COJ), raw egg (RE), combination feeds of BE and RE (BE+RE) and combination feeds of BE and COJ (BE+COJ),
  C, the highest survival rate showed by larvae fed with boiled egg yolk (BE). Vertical axis shows the survival rate in percentage (%), horizontal axis shows the age of larvae in day after hatching (dAH)
- Figure 4.3: Changes of the BE and COJ size selectivity on 3,5,7 and 9 dAH *Lutjanus rivulatus* and the sizes of BE and COJ available in the rearing water
- Figure 4.4: Changes of feed type selectivity by of 3, 5, 7 and 9 dAH *L. rivulatus* larvae fed with BE and COJ by different feeding day through Chesson's a-index. Data are showed in mean ± standard deviation. The centre black line shows the preference index at 0.5, the horizontal axis shows the age of larvae in day after hatching (dAH) and the vertical axis shows the preference index of larvae





47

53

Figure 4.5: Changes of the mean gut passage time on 3, 5, 7, and 9 dAH *L. rivulatus* larvae fed with boiled egg yolk (BE) and Cod oil juice (COJ). Vertical bar shows the gut passage time in minutes. Horizontal bar shows the age of larvae in day after hatching (dAH)



### LIST OF PHOTOS

- Photo 3.1: Five artificial feeds were tested as alternative starter feed for *L. rivulatus* larvae. **A**, Raw egg yolk (RE); **B**, cod oil juice (COJ); **C**, boiled egg yolk (BE); **D**, combination feed of boiled egg yolk and cod oil juice (BE+COJ); **E**, combination feed of raw egg yolk and Cod oil juice (RE+COJ). Each feed was well homogenized with seawater by using homogenizer (Ultra Turax, T25 Basic Electrical Homogenizer, IKA Labortechnik, Malaysia). All feeds were stored in refrigerator under 4<sup>o</sup>C.
- Photo 3.2: Boiled egg yolk (BE) feed characterization. A, Processed BE was sampled; B, BE characterization was done under an optical microscope (Nikon, Eclipse E600, Japan); C, the BE size was measured by ocular microscale, the shape was characterized by capturing the image displayed by BE using digital camera, the colour was identified through the observation under the optical microscope; D, BE at 100X magnifying under optical microscope, scale bar, 100 μm.
- Photo 3.3: The raw ingredients and equipment used to prepare the artificial feed. **A**, Boiled egg yolk, 19.50±0.94 g; **B**, raw egg yolk, 46.50±0.94 g; **C**, cod liver oil, 70 ml for each feed prepared; **D**, raw ingredient was homogenized well with 700 mL seawater at 13,000 r.p.m in five minutes to obtain fine texture of feed. Processed feed was stored in refrigerator under 4 °C.
- Photo 3.4: Method of harvesting natural plankton. **A**, Submersible pump (KP Basic 300 A, 0.3 HP, 7 m, Grundfus Pumps Sdn. Bhd,



xiv

PAGE

27

25

Malaysia) was used to collect the scatter natural plankton from the sea near UMS Hatchery; **B**, A 250  $\mu$ m collection net was set up inside 2 tonnes (1.2 x 2.4 x 0.75 m) of FRP (Fibre Reinforce Plastic) tank to keep the natural plankton, **C**, 500  $\mu$ m plankton net was used to separate detritus and minerals, which attached with to the natural plankton; **D**, collected natural plankton were sieved to obtain desirable size.

- Photo 3.5: Harvesting rotifer, *Branchionus* sp. method. A, 60 μm collection net was tied at the outlet pipe of the rotifer mass culture tank in UMS Hatchery; B, the collection net was gently pressed to avoid it to burst due to continuous water pressure;
  C, rotifer were sieved by using 250 μm plankton net to eliminate detritus, 100 μm plankton net was used to obtain approximate 80 μm size of rotifer for larvae, approximately 80 μm; D, rotifer were rinsed by using seawater before being ready to be fed to larvae.
- Photo 3.6: L. rivulatus broodfish management and egg collection system in UMS Hatchery; A, Collected L. rivulatus adult were placed in 150 tonnes (3 m depth and 8 m diameter) of HDPE (High Density Poly Ethylene) circular tank, B, L. rivulatus adults were reared for 12 months and became broodfish that are able to spawn naturally in the tank; C, 250 µm egg collection net was set up at the water outlet pipe of biological filter tank to collect L. rivulatus eggs through the water circulation system; D, L. rivulatus eggs were collected gently from the egg collection net and rinsed before transferred into the incubation tank.
- Photo 3.7: Tank and rearing water preparations for *L. rivulatus* larval rearing. **A**, 16 of 80 L FRP tanks were used to carried out the *L. rivulatus* larval rearing; duplicate were prepared for each



29

31

32

XV

treatment, microalgae, *Nannochloropsis* sp. were added to the rearing water and aerated gently; **B**, larvae were transferred into the rearing tank with the larval stocking density at 30 individuals/L.

- Photo 3.8: Water and aeration systems for *L. rivulatus* larval rearing. A, Seawater was filtered by 60 µm plankton net before flowing into tank; B, microalgae, *Nannochloropsis* sp. of was added into each tank at 5x10<sup>6</sup> cell/ml density; C, air stone was used to aerate the rearing water; moderate aeration was applied into tank D, rearing water was ready for the larval rearing.
- Photo 3.9: The bottom cleaning process for the *L. rivulatus* larval rearing tank. A, The design of bottom cleaner by the UMS Hatchery; a hosepipe was connected to the siphon pipe; B, a horizontal cut was made at the end of siphon pipe and scour pad was attached to it, which act as vacuum cleaner; C, bottom cleaner was placed at the bottom of the tank and the accumulated sediments, which sank at the bottom was gently siphoned out; D, the outlet of the hosepipe was placed in 100 µm plankton net to prevent larvae to be escaped.
- Photo 3.10: Two different gut content observation methods. **A**, First observation method: the physical appearance of larvae gut were observed by using naked eyes and the presence of feed is indicated by the coloured gut; **B**, second observation method: the type of feed presented in the larvae gut were identified by observation under an optical microscope, and the images displayed by larvae gut were captured by digital camera.

Photo 3.11: Feed size selectivity measurement method. A, Sampled larvae



xvi

34

35

36

were transferred and separated in different compartments in multi-disc; **B**, larvae were observed under an optical microscope and the feed size available in the larvae gut were measured by using the optical microscope.

- Photo 3.12: Feed selectivity of larvae were indicated through selectivity index, Chesson's a (Chesson, 1978). A, Sampled larvae from each rearing tank were transferred and separated in different compartments of multi-disc; B, larvae were placed under optical microscope and the ratio and number of different feed found in the gut was counted.
- Photo 3.13: The gut passage time of larvae fed with BE and COJ. **A**, Sampled larvae from each rearing tank were transferred and separated in different compartments of multi-disc; **B**, anaesthesia was added into in order to immobilize larvae.
- Photo 4.1: Feed dark brownish color with uneven shape, 5-180 μm; B, Cod oil juice (COJ); transparent oil-alike shape with wide range of size, 5-180 μm; C, raw egg yolk (RE) brownish water based feed; D, rotifer, *Brachionus* sp.; semi spherical and cylindrical shapes, 150-230 μm; E, Natural plankton, ellipsoidal shape with 90-160 μm, scale bar, 100 μm.
- Photo 4.2: The gut contents of *L. rivulatus* larvae fed with different feeds. **A**, BE was appeared as dark brownish and uneven shape in *L. rivulatus* gut; **B**, COJ was found as transparent oilalike feed with various sizes; C, RE was found as brown water based feed in the *L. rivulatus* gut; **D**, BE+COJ was observed being similar as the combination characteristics of BE and COJ in the *L. rivulatus* gut; **E**, BE+RE was appeared as the combination characteristics of BE and RE in the *L. rivulatus*



xvii

40

41

42

gut, scale bar, 100 µm.

- Photo 4.3: The gut contents of *L. rivulatus* fed with rotifer and natural plankton, and larvae with no feeds were given. **A**, Gut of *L. rivulatus* fed with rotifer was empty, no rotifer were found in the gut; **B**, *L. rivulatus* fed with natural plankton was empty, no natural plankton were found in the gut; **C**, microalgae, *Nannochloropsis* sp. were found in the gut, scale bar, 100 µm.
- Photo 4.4: The feed selectivity of boiled egg yolk and Cod oil juice by *L. rivulatus* larvae; **A**, BE was found in various sizes, shape, number and different location in the larvae gut under an optical microsope; **B**, COJ was also found in various sizes, number and different location in the larvae gut. The ratio and number of both feeds that presented in the larvae gut were used to determine the larval preference feed for consumption, sclae bar 100 μm.
- Photo 4.5: Boiled egg yolk (BE) was found in the excretion gut of 3 dAH Lutjanus rivulatus larvae. A, BE was found in the stomach with larger size and gradually moved to the excretion gut; B, BE was found in the excretion gut and with smaller size of BE. scale bar, 100 μm

62

55



xviii

50

### LIST OF ABBREVIATIONS

%	percentage
&	and
μm	micrometer
AH	after hatching
BE	boiled egg yolk
BL	body length
BW	body weight
COJ	cod oil juice
DHA	docosahexaenoic acid
DO	dissolved oxygen
EPA	eicosapentaenoic acid
ESD	equilibrium spherical diameter
FAO	Food and Agriculture Organisation
FFA	free fatty acid
FRP	fibreglass reinforced plastic
g	gram
h	hour
hAH	hour after hatching
HDPE	high density poly ethylene
HP	house power
IU	international unit
kg	kilogram
kj	kilojoule
1	litre
L	length
m	number of tested feed
mg	milligram
mL	mililiter
mm	millimetre



п	number
°C	degree celsius
рН	potantra of hydogeni (power of
	hydrogen)
ppt	part per thousand
r	r/ propotion
r.p.m	revolution per minute
RE	raw egg
V	volume
W	width
a	alpha
Л	pi



### CHAPTER 1

### INTRODUCTION

### 1.1 HOI TAI KAI, Lutjanus rivulatus

*Lutjanus rivulatus* is classified in the genus of Lutjanus, class of Actinopterygii (rayfinned fishes), order of Perciformes (perch like), family of Lutjanidae (snappers); subfamily of Lutjaninae (Allen, 1985). The common names are White blotch snapper and Blubberlips (Allen, 1985). In Sabah, Malaysia, *L. rivulatus* is known by various names such as Ketambak, Konai, Nai Wong, Hoi Tai Kai, Hai Ti Chi and Siak Bong (Chin, 1998).

Many different names are given to *L. rivulatus* (Table 1.1) due to the wide distribution. *L. rivulatus* can be found in most tropical countries, particularly in Southeast Asia (Lee, 1998), such as Malaysia, Indonesia and Philippines, and other countries, namely Japan, Australia, Africa, United States of America and United Kingdom.

Country	Names
Africa	Speckled snapper
America	Blubberlips snapper
Australia	Maori-seaperch
China	Hai Ti Chi
Indonesia	Gaga
Japan	Nami-fuedai
Malaysia	Hoi Tai Kai
Philipines	Agak-agak

**Table 1.1** Names of *Lutjanus rivulatus* in different countries (Allen, 1985)



*L. rivulatus* is one of the most outstanding fish in the seafood industry (Polovina & Ralston, 1987; Department of Fisheries Sabah, 2004). In recent years, *L. rivulatus* has become one of the most expensive marine food fish (Senoo *et al.*, 2002; Department of Fisheries Sabah, 2004). In Kota Kinabalu, Sabah, *L. rivulatus* is sold for RM 120 -160 per kg in seafood restaurants (Own survey in July 2006). The meat quality of *L. rivulatus* is superior. This makes *L. rivulatus* often being the favorite food fish in both local and overseas markets.

Due to the meat quality, the demand for *L. rivulatus* in the seafood industry has gradually increased, while, the outstanding price has generated interest among seafood operators to commercialize *L. rivulatus* in order to gain higher profit (Chin, 1998). However, *L. rivulatus* is widely harvested from the wild to meet the increasing market demand. The continuous harvest of *L. rivulatus* has caused the population to decrease in the wild (Andrade, 2003). As a result, the supply of *L. rivulatus* has become insufficient, causing an imbalance in the market demand and supply (Andrade, 2003).

### 1.2 L. rivulatus SEED PRODUCTION IN UMS HATCHERY

The inconsistent supply of *L. rivulatus* in the market has led to the need of the seed production. It is aimed to produce sufficient supply of *L. rivulatus* to the market. In 2004, the Marine hatchery of Borneo Marine Research Institute, Universiti Malaysia Sabah (UMS hatchery) has started the effort to produce *L. rivulatus* seeds through aquaculture. Broodfish of *L. rivulatus* were collected from the wild and reared in the UMS hatchery. Reared broodfish are consistently produce eggs under the artificial condition.



### REFERENCES

- Andrade, F.M. 2003. Comparison of Life History and Ecological Aspects among Snappers, *LUTJANIDAE*. Department of Oceanography and Coastal Sciences: Louisiana State University.
- Allen, G.R. 1985. Snappers of the World, an Annotated and Illustrated Catalogue of Lutjanidae Species Known to Date. Food and Agriculture Organization (FAO). 1985.
- Arimoro, F.O. 2006. Culture of Freshwater Rotifer, Brachionus calyciflours and its Application in Fish Larviculture Technology. African Journal of Biotechnology. 5: 536-541.
- Azzaydi, M., Martinez, F.J., Zamora, F.J. & Madrid, J.A. 2000. The Influence Nocturnal and Diurnal Feeding under Winter Conditions on Growth and Feed Conversion of European Seabass, *Dicentratchus labrax* Larvae. *Aquaculture*. **182**: 329-338.
- Bagarinao, T. 1986. Yolk Resoption, Onset of Feeding and Survival Potential of Larvae of Three Tropical Marine Fish Species Reared in Hatchery. *Marine Biology*. **91**: 449-459.
- Battaglene, S., Purser, J., Hart, P. & Morehead, D. 2000. Priority for Live Feed Production and Research in Tasmania. *Proceeding of Hatchery Feeds*. March 9-10, 2000. Cairns, Australia.
- Bell, J.G., McEvoy, L.A., Estevez, A., Shield, R.J. & Sargent, J.R. 2003. Optimizing Lipid Nutrition in First Feeding of Flatfish Larvae. Aquaculture. 227: 211-220.
- Bisbal, G.A. & Bengston, D.V. 1995. Effects of Delayed Feeding on Survival and Growth of Summer Flounder, *Paralichthys dentatus. Marine Ecology Progress Series*. 121: 301-306.
- Bunnell, D.B., Maria, J.G. & Roy, A.S. 2003. Zooplankton Biomass Enhances Growth, but Not Survival, of First Feeding *Pomoxis* spp. *Canadian Journal Fisheries and Aquatic Sciences.* 60: 1314-1323.
- Buskey, E. J. 2005. Behavioral Characteristics of Copepods that Effects Their Suitability as Food for Larval Fishes. In Lee, C.S., O'Bryen, P.J. & Marcus, N.H. (eds.). Copepodes in Aquaculture. 91-106. Oxford: Blackwell Publishing.
- Cahu, C.L. & Zambonino, J.L. 1994. Early Weaning of Sea Bass, *Dicentrarchus labtrax* Larvae with a Compound Diet: Effect on Digestive Enzymes. *Biochemistry Physiology*. **109**: 213-322.
- Cahu, C. & Infante. J.Z. 2001. Substitute of Live Food by Formulated Diets in Marine Fish Larvae. Aquaculture. 200: 161-180.



- Canino, D.V.P. & Bailey, K.M. 1995. Gut Evacuation of Walleye Pollock Larvae in Response to Feeding Condition. *Journal of Fish Biology*. **46**: 389-403.
- Chapman, F.A., Colle, D.E., Rotmann, R.W. & Shireman, J.V. 1997. Controlled Spawning of the Neon Tetra. *The Progressive Fish-Culturist*. **60**: 32-37.
- Chesney, E. J. 2005. Copepodes as Live Prey: A Review of Factors that Influencing the Feeding Success of Marine Fish Larvae. *In* Lee, C.S., O'Bryen, P.J. & Marcus, N.H. (eds.). *Copepodes in Aquaculture*. 133-150. Oxford: Blackwell Publishing.

Chesson, J. 1978. Measuring Preference in Selective Predation. Ecology. 59: 211-215.

- Chin, P.K. 1998. *Marine Food Fishes and Fisheries of Sabah*. Kota Kinabalu: Natural History Publications.
- Chow, K.W. 1980. *Microencapsulated Egg Diets for Fish Larvae*. Food and Agriculture Organization (FAO). 1980.
- Cunha, I. & Planas, M. 1999. Optimal Prey Size for Early Turbot Larvae *Scophthalmus maximus* Based on Mouth and Ingested Prey Size. *Aquaculture*. **175**: 103–110.
- Curnow, J., King, J., Partridge, G. & Kolkovski, S. 2006. Effects of Two commercial Microdiet on Growth and Survival of Barramundi, *Lates calcarifer* Larvae within Various Early Weaning Protocols. *Aquaculture Nutrition*. **12**: 247-255.
- Cuvier & Valenciennes. 1828. *Histoire Naturelle Des Poissons*. 1-490. Paris: History Natural Poission.
- D'Abramo, 2002. Challenges in Developing Successful Formulated Feed for Culture of Larval fFsh and Crustaceans. *In* Cruz-Suarez, L.E., Rique-Marie, D., Tapia, M.S., Gaxiola-Coprtes, M.G. and Simoes, N. (eds.). *Proceeding of Avances en Nutricion Acuícola VI. Memoria de VI Symposium Internacional de Nutricion Acuícola*. 3-6 Cancún, Quintana Roo, Mexico
- Dabrowski, K. & Bardega, R. 1984. Mouth Size and Predicted Feed Size Preference of Larvae of Three Cyprinid Fish Species. *Aquaculture*. **40**: 41-46.
- Dagg, M.J & Walser, W.E. 1987. Ingestion, Gut Passage and Egestion by the Copepod Neocalanus plumchrus in the Laboratory and in the Subartic Pacific Ocean. Limnology and Oceanography. 32: 178-188.
- Das, S.K & Kalita, N. 2003. Captive Breeding of Peacok Eel, *Macrognatus aculeatus*. Aquaculture Asia. 3: 17-21.
- Deng, D.F., Koshio, S., Yokoyama, S., Bai, S.C., Shao., Q., Cui., Yobo. & Hung, S.O. 2003. Effects of Feeding Rate on Growth Performance of White Sturgeon, Acipenser transmointanus Larvae. Aquaculture. 217: 589-598.



- Doi, M., Toledo, J.D., Golez, M.S.N., Santos, M.D.A. & Ohno, A. 1997. Preliminary Investigation of Feeding Performance of Larvae of Early Red Spotted Grouper, *Epinephelus coioides*, Reared with Mixed-zooplankton. *Hydrobiology*. 358: 259-263.
- Doi, M. & Sunghagaraiwan, T. 1993. *Biology and Culture of the Red Snapper, Lutjanus agrentimaculatus.* The Eastern Marine Fisheries Development Centre: Thailand.
- Dou, S., Masuda, R., Tanaka, M. & Tsukamoto, K. 2005. Effects of Temperature and Delayed Initial Feeding of Japanese Flounder Larvae. *Journal of Fish Biology*. 66: 362-377.
- Dou, S., Masuda, R., Tanaka, M. and Tsukamoto, K. 2002. Feeding Resumption, Morphological Changes and Mortality During Starvation in Japanese Flounder Larvae. *Journal of Fish Biology*. 60: 1363-1380.
- Drass, M. D. 2000. Larval Development of Red Snapper, *Lutjanus campachanus*, and Comparisons with Co-occurring Snapper Species – Statistical Data Included. *Fisheries Bulletin*, 98:3.
- El-Dakar, A.Y., Shalaby, S.M., Hassanein, G.D. & Ghoneim, S.I. 2001. Use of Rotifers Cultured on Different Microalgal Species in Larval Feeding of Seabass, *Dicentrarchus labrax. Asian Fisheries Science.* 14: 43-52.
- Eldin, M.S., De Silva., S.S. & Ingram, B.A. 1997. Effects of Diets and Feeding Rate on the Survival and Growth of Macquarie Perch, *Macquaria austmlasica* Larvae, a Threatened Australia Native Fish. *Aquaculture*. **157**: 35-50.
- Fernandez., D.C., Pascual., E. & Yufera, M. 2006. Feeding Behavior and Prey Selection of Gilthead Seabaream, Sparus aurata, Larvae Fed on Inert and Live Food. Marine Biology. 118: 323-328.
- Fielder, D.S., Bardsley, W.J., Allan, G.L. & Pankhurst, P.M. 2005. The Effects of Salinity and Temperature on Growth and Survival of Australia Snapper, *Pagrus auratus* Larvae. *Aquaculture*. **250**: 201-214.
- Fleeger, J.W. 2005. The Potential to Mass Culture Harpacticoid Copepods for Use as Food for Larval Fish. In Lee, C.S., O'Bryen, P.J. & Marcus, N.H. (eds.). Copepodes in Aquaculture. 11-24. Oxford: Blackwell Publishing.
- Garcia, L.M.B., Garcia, C.M.H., Pineda, A.F.S., Gammad, E.A., Canta, J., Simon, S.P.D., Hilomen-Garcia, G.V., Gonzal, A.C. & Santiago, C.B. 1999. Survival and Growth of Bighead Carp Fry Exposed to Low Salinities. *Aquaculture International.* 7: 241-250.
- Gawlicka, A., Parent, B., horn, M.H., Ross, N., Opstad, I. & Torrissen. 2000. Activity of Digestive Enzymes in Yolk-Sac Larvae of Atlantic Halibut , *Hippoglossus hippoglossus*: Indication of Readiness for First Feeding. *Aquaculture*. **184**: 303– 314.



- Gisbert, E., Conklin, D.B. & Piedrahita, R.H. 2004. Effects of Delayed First Feeding on the Nutritional Condition and Mortality of California Halibut Larvae, *Paralichthys californicous. Journal of Fish Biology*. **116**:116-132.
- Hagiwara, A., Gallardo, W.G., Assavaaree, M., Kotani, T. & de Araujo, A.B. 2001. Live Food in Japan: Recent Progress and Future Aspects. *Aquaculture*. **200**: 111-127.
- Hansson, L.J., & Kiørboe, T. 2006. Prey-specific Encounter Rates and Handling Ambushfeeding Hyromedusae. *Limnology and Oceanography* . **51**: 1849-1858.
- Hossain, Q., Hossain, M.A. & Parween S. 2006. Artificial Breeding and Nursery Practices of *Claias batrachus* (Linnaeus, 1758). *Scientific World*. **4**: 32-37.
- Ignacio, A.C.A. 2003. Condition Indices and Their Relationship with Environmental Factors in Fish Larvae. Department of D'Ecologia: Universitat De Barcelona.
- Ingram, B., Sungan, S., Gooley, G., Sim, S.Y., Tinggi, D. & De Silva, S.S. 2005. Induced Spawning, Larval Development and Rearing of Two Indigenous Malaysian Mahseer, *Tor tambroiders* and *T. douronesis. Aquaculture Research.* 36: 983-995.
- Iguchi, K. & Maizuno, N. 1998. Early Starvation Limits Survival in Amphidromous Fishes. Journal of Fish Biology. 54:705-712.
- Jennings, B.R. & Parslow, K. 1988. Particle Size Measurement: The Equivalent Spherical Diameter. Mathematical and Physical Sciences. 419: 137-149.
- Juanes, F., J.A. Buckel, and F.S. Scharf. 2001. Predatory Behaviour and Selectivity of a Primary Piscivore: Comparing Fish and Non-fish Prey. *Marine Ecology Progress* Series. 217:157-165.
- Johnson, J.H. & Dropkin, D.S. 1995. Effects of Prey Density and Short Term Food Deprivation on the Growth and Survival of American Shad Larvae. *Journal of Biology*. 46: 872-879.
- Juario, J.V., Duray, M.H., Duray, V.M., Nacario, J.F. & Almendras, M.E. 1985. Breeding and Larval Rearing of the Rabbitfish, *Siganus guttatus* (Bloch). *Aquaculture*. 44: 91-101.
- Kaji, T., Kosama, M., Arai, H., Tanaka, M. & Tagawa, M. 2003. Prevention of Surface Death of Marine Fish Larvae by the Addition of Egg White into Rearing Water. *Aquaculture*. 224: 313-322.
- Kailasam, M., Thirunavukkarasu, A.R., Seluaraj, S. & Stalin, P. 2007. Effects of Delayed Initial Feeding on Growth and Survival of Asian Sea Bass, *Lates calcarifer* (Blotch) Larvae. *Aquaculture*. 271: 298-306.



- Kanazawa, A. 2003. Nutrition of Marine Fish Larvae. In Jana, B. B & Webster, C. D. (eds.). Sustainable Aquaculture : Global Perspectives. 103-144. New York: The Haworth Press, Inc.
- Karjalainen, J.J. & Viljanen, M. 1991. The Gastric Evacuation Rate of Vendae Coregonus albula Larvae Predating on Zooplankter in the Laboratory. Aquaculture. 96: 343-351.
- Khan, M.H.K. & Mollah, M.F.A. 2004. Further Trials on Induced Breeding od *Pangasius* pangasius (Hamilton) in Bangladesh. Asian Fisheries Science. **17**: 135-146.
- King, A.J. 2004. *Density and Distribution of Potential Prey for Larval Fish in the Main Channel of a Floodplain River: Pelagic versus Epibenthic Meiofauna*. 883-897. San Francisco : John Wiley & Sons Ltd.
- Kohno, H. 1998. Early History Features Influencing Larval Survival of Cultivated Tropical Finfish. In De Silva, S.S. (ed.). Tropical Mariculture. 71-110. California: Academic Press.
- KolKovski, S. 2001. Digestive Enzymes in Fish Larvae and Juvenile- Implications and Application to Formulated Feed. *Aquaculture*. **200**: 181-201.
- Koven, M., Kolkovski, S., Hadas, E., Gamsiz, K. & Tandler, A. 2000. Advances In The Development Of Microdiets For Gilthead Seabream, *Sparus Aurata*: A Review. *Aquaculture*. **194**: 107-121.
- Kraul, S. 1989. Production of Live Prey for Marine Fish Larvae. Advances in Tropical Aquaculture. 9:595-607.
- Kraul, S. 2006. Live Food for Marine Larvae. In Elizabeth-Cruz, L., Suarez-Dennis, R.M., Salazar, M.T., Martha, G., David, A., Ana, C. & Puello-Cruz, Y.A.G. (eds.). Proceeding of Avances en Nutricion Acuicola VIII. Sinaloa: Mexico.
- Kuiter, R. & Tonozuka, T. 2001. Pictorial Guide to Indonesia Reef Fishes. Part 1. Eels-Snappers, Muranidae-Lutjanidae. Zoonetics. 302:294.
- Kungyankij, P. (ed.). 1989. *Hatchery Management Techniques in Marine Fish Culture Development*. Food and Agriculture Organization (FAO). 1989.
- Langdon, C. 2003. Microparticle Types for Delivering Nutrients to Marine Fish Larvae. Aquaculture. 227: 259-275.
- Lavens, P. & Sorgeloos, P. 1996. *Manual on the Production and Use of Live Food for Aquaculture*. Food and Agriculture Organization (FAO). 1996.
- Lavens, P. Sorgeloos, P., Dhert, P. & Devresse, B. 1995. Larval Foods. In Bromage, N. & Robert, R.J. (eds.). Broodstock Management and Egg and Larval Quality. 373-394. Oxford: Blackwell Publishing.



- Lee, C.S. 1998. Culture of Marine Finfish Species of the Pacific. In De Silva, S.S. (ed.). Tropical Mariculture. 361-380. California: Academic Pres.
- Lee, C. S & Sadovy, Y. 1998. A Taste for Live Fish: Hong Kong's Live Reef Fish Market. Naga ICLARM. 21: 38-42.
- Leong, T.S. 1998.Grouper Culture. In De Silva, S.S. (ed.). Tropical Mariculture. 423-448. California: Academic Pres.
- Liao, I.C., Su, H.M. & Chang, E.Y. 2001. Techniques in Finfish Larviculture in Taiwan. Aquaculture. 200: 1-31.
- Lim, L. C., Dher, P. & Sorgeloos, P. 2004. Recent Developments in the Application of Live Feed in the Ornamental Fish Culture. *Aquaculture*. **227**: 319-331.
- Marcus, N.H. 2005.Calanois Copepodes, Resting Eggs and Aquaculture. In Lee, C.S., O'Bryen, P.J. & Marcus, N.H. (eds.). *Copepodes in Aquaculture*. 3-10. Oxford: Blackwell Publishing.
- Mayer, C.M. & Wahl, D.H. 1997. The Relationship Between Prey Selectivity and Growth and Survival in a Larval Fish. *Canadian Journal Fish Aquatic Science*. **54**: 1504-1512.
- McKinnon, A.D., Duggan, S., Nicholas, P.D., Rimmer, M.A., Semmens, G., & Robino, B. 2003. The Potential of Tropical Parsacalanis Copepode as Live Feeds in Aquaculture. Aquaculture. 223: 89-106.
- Meeren, T.V.D. 1991.Selective Feeding and Prediction of Food Consumption in Turbot Larvae, (*Scophthalmus maximus* L) Reared on the Rotifer, *Brachionus plicatilis* and Natural Zooplankton. *Aquaculture*. **93**: 35-55.
- Michael, J.D. & Walsey, W.E. 1987. Ingestion, Gut Passage and Egestion by the Copepod Neocalanus plumchrus in the Laboratory and in the Subartic Pacific Ocean. Limnology and Oceanography. **32**: 178-188.
- Mohsin, M., Abd Khair & Ambak, M., Azmi. 1996. *Marine Fishes and Fisheries of Malaysia* and Neighboring Countries. Kuala Lumpur: Universiti Pertanian Malaysia.
- Molejon, O.G.H. & Lajonchere, L.A. 2003. Culture Experiments With Oithona Oculata Farran, 1913 (Copepoda: Cyclopoida), and its Advantages as Food for Marine Fish Larvae. Aquaculture. 219: 471-483.
- Mookerji, N., & Rao, T.R. 1999. Rates of Yolk Utilization and Effects of Delayed Initial Feeding in the Larvae of Freshwater Fishes Rohu and Singhi. Aquaculture International. 7: 1-12.
- Morehead, D. T., Battaglene, S.C., Metillo, E.B., Bransden, M.P. & Dunstan, G. A. 2005. Copepodes as a Live Feed for Striped Trumpeter *Latris lineate* Larvae. *In* Lee,



C.S., O'Bryen, P.J. & Marcus, N.H. (eds.). *Copepodes in Aquaculture*. 195-206: Blackwell Publishing.

- Moteki., M., Yoseda, K., Sahin, T., Ustundag, C. & Kohno, H. 2001.Transition from Endogenous to Exogenous Nutritional Sources in Larvae of Black Sea Turbot, *Psetta maxima. Fisheries Science.* **67**: 571-578.
- Nakagawa, Y., Senoo, S. & Miyashita, S. 2007. Protozoa and Diatoms as Primary Food Sources for Larvae of the Blue Spotted Snapper, *Lutjanus rivulatus*. *Aquaculture Science*. **55**: 125-130.
- Ng, W.K., Lu, K.S., Hashim, R. & Alis, A. 2000. Effects of Feeding Rate on Growth, feed Utilization and Body Composition of A Tropical Bagrid Catfish. *Aquaculture International.* 8: 19-29.
- O' Bryen, P.J. & Lee, C.S. 2005 Culture of Copepods and Applications to Marine Finfish Larvae Rearing, Workshop Discussion Summary. *In* Lee, C.S., O'Bryen, P.J. & Marcus, N.H. (eds.). *Copepodes in Aquaculture*. 245-255. Oxford: Blackwell Publishing.
- Ogle, J.T., Lemus, J.T., Nicholson, L.C., Barnes, D.N. & Lotz, J.M. 2005. Characterization of an Extensive Zooplankton Culture System Coupled with Intensive Larval Rearing of Red Snapper, *Lutjanus campechanus*. *In* Lee, C.S., O'Bryen, P.J. & Marcus, N.H. (eds.). *Copepodes in Aquaculture*. 225-244. Oxford: Blackwell Publishing.
- Opuszynski., K. & Shireman, J.V. 1991.Food Passage Time and Daily Ration of Bighead Carp, Aristichthys nobilis, Kept in Cages. Environmental Biology of Fishes. 30: 387-393.
- Pankhurst, P.M., & Eager, R. 1998. Changes in Visual Morphology Through Life History stages of the New Zealand Snapper, *Pagrus auratus. Journal of Marine and Freshwater Research.* 30: 79-90.
- Pankhurst, P.M., Montgomery, J.C. & Pankhurst, N.W. 1991. Growth, Development and Behavior of Artificial Reared Larval Pagrus auratus (Bloch and Schnidae, 1801) (Sparidae). Australian Journal of Marine and Freshwater Research. 42: 391-398.
- Payne, M.F. & Rippunagle, R.J. 2000. Effects of Salinity, Cold Storage and Enrichment on the Calanoid Copepod Gladioferens imparupes. Aquaculture. 201: 251-262.
- Payne, M.F., Rippingale, R.J. & Cleary, J.J. 2001. Cultured Copepods As Food For West Australian Huffish (*Glaucosoma Hebraicum*) and Pink Snapper (*Pagrus Auratus*) Larvae. *Aquaculture*. **194**: 137-150.
- Pena, R. & Dumas, D. 2005. Effects of Delayed First Feeding on Development and Feeding Ability of *Paralabrax maculatofasciatus* Larvae. *Journal of Fish Biology*. 67:640-651.



- Petkem, R. & Moodie, G.E.E. 2001. Food Particles Size, Feeding Fequency, and the Use of Prepared Food to Culture Larval Walking Catfish, *Clarias macrocephalus*. *Aquaculture*. **194:** 349-362.
- Phelps, R. P., Sumiaarsa, G. S., Lipman, E.E., Hsiang, P.L., Moss, K.K. & Davis, A.D. 2005. Intensive and Extensive Production Techniques to Provide Copepod Nauplii for Feeding Larval Red Snapper, *Lutjanus campechanus. In* Lee, C.S., O'Bryen, P.J. & Marcus, N.H. (eds.). *Copepodes in Aquaculture*. 151-168. Oxford: Blackwell Publishing.
- Planas, M. & Cunha, I. 1998. Laviculture of Marine Fish: Problems and Perspectives. Aquaculture. 177: 171-190.
- Polovina, J.J. & Ralston, S. 1987. Tropical Snappers and Groupers: Biology and Fisheries Management. 37-67. Colorado: Westview Press.
- Puvanendran, V. & Brown, J.A. 1999. Foraging, Growth and Survival of Atlantic Cod Larvae Reared in Different Prey Concentration. Aquaculture. 175: 77-92.
- Qin, J.G. & Hillier. 2000. Live Food and Feeding ecology of Larval Snapper, Pagrus auratus. Proceeding of Hatchery Feeds. March 9-10, 2000. Cairns, Australia.
- Rajkumar, M & Kumaraguru, K.P. 2006.Sustainability of the Copepods, Acartia clause as a Live Feed for Seabass Larvae, Lates calcarifer\_: Compared to Traditional Live feed Organisms with Special Emphasize on the Nutritional Value. Aquaculture. 261: 649-658.
- Rao, T.R. 2003.Ecological and Ethological Perspective in Larval Fish Feeding. In Jana, B.B. & Webster, C.D. (eds.). Sustainable Aquaculture Global Perspective. 145-178. New York: The Haworth Press, Inc.
- Rettig, J.E. 2003. Zooplankton Responses to Predation by Larval Bluegill: an Enclosure Experiment. *Freshwater Biology*. **48**: 636-648.
- Rhodes, A., & Boyd, L. 2005. Formulated Feeds for Harpacticoid Copepodes: Implications for Populations and Growth and Fatty Acid Composition. *In* Lee, C.S., O'Bryen, P.J. & Marcus, N.H. (eds.) *Copepodes in Aquaculture*. 75-98. Oxford: Blackwell Publishing.
- Rimmer, M & Russell, 1998. Aspects of the Biology and Culture of *Lates calcarifer*. *Tropical Mariculture*. 449-476. Califorina: Academic Press.
- Rimmer, M. 2000. Issues Raised In General Discussion at The Hatchery Feeds Workshop. Proceeding of Hatchery Feeds. March 9-10, 2000. Cairns, Australia.
- Rippingale, R.J. & Payne, M.F. 2005. Suitability of the Copepode *Gladioferens imparipes* for Intensive Cultivation of Aquaculture. *In* Lee, C.S., O'Bryen, P.J. & Marcus, N.H. (eds.). *Copepodes in Aquaculture* . 102-135. Oxford: Blackwell Publishing.



- Roo, F.J., Socorro, J., Izquierdo, M.S., Caballero, M.J., Hernandez-Cruz, M., Fernandez, A. and Hernandez-Palacious, H. 1999. Development of Red Porgy, *Pagrus pagrus* Visual System in Relation with Changes in the Digestive Tract and Larval Feeding Habits. *Aquaculture*. **179**: 499-512.
- Sabatez, A. & Saiz, E. 2000. Intra and Interspecific Variability In Prey Size and Niche Breath of Myctophiform Fish Larvae. *Marine Ecology Progress Series.* **201**: 261-271.
- Santiago, .B., Ricci, M. & Lampa, A.R. 2004.Effects of Nematode Panagreallus redivivus Density on growth, Survival, Feed Consumption and Carcass Composition of Bighead Carp, Avistichthys nobilis Larvae. Journal of Applied Ichthyology. 6: 22-27.
- Schabetsberger, R., Sztatecsny, M., Drozdowski, G., Broder, R.D., Swartzman, G.L., Wilson, M.T., Winter, A.G. & Napp, J.M. 2003. Size Dependent, Spatial and Temporal viability of Juvenile Walleye Pallock, *Theragra chalcograma* Feeding at a Structure Front in the Southeast Bering Sea. *Marine Ecology*. 24: 141-164.
- Scharf, F.S., James, F. & Rountree, R.A. 2000. Predator Size-Prey Size Relationships of Marine Fish Predators : Interspecific Variation and Effects on Ontogeny and Body Size on Tropic Niche Breadth. *Marine Ecology Progress Series*. 208: 229-248.
- Schipp, G.R., Bosmans, J.M.P. & Marshall, A. 1999. A Method for Hatchery Culture of Tropical Calanois Copepods, *Arcatia* spp. *Aquaculture*. **174**. 81-88.
- Schmitt, P.D. 1986. Prey Size Selectivity and Feeding Rate of Larvae on the Northern Anchovy, *Engraulis mordazgirard. California Cooperative Oceanic Fisheries Investigation* (CalCOFI ). 1986.
- Senoo, S., Nguang, S.I. & Miyashita, S. 2006. Observation on the Development of Early Larval Stage of Hoi Tai Kai, *Lutjanus rivlatus* for the Artificial Seed Production. *Proceeding of The Japanese Fisheries Society Conference*. March 20-23, 2006. Kochi University, Japan.
- Shaw, G.H., Pankhurst, P.M. & Battaglen, S.C. 2006. Effect of turbidity, Prey Density and Culture History on Prey Consumption by Greenback Flounder, *Rhombosolea tapirina*. Aquaculture. 253: 447-460.
- Shoji, J., Aoyama, M., Fujimoto, H., Iwanoto, A.& Tanaka, M. 2002. Susceptibility to Starvation by Piscivorous Japanese Spanish Mackerel Larvae at First Feeding. *Fisheries Science*. 68:59-64.
- Sommer, C., Schneider, M & Poutiers, J.M. 1996. FAO Species Identification Field Guide for Fisheries Purposes: The Living Marine Resources of Somalia. Food and Agriculture Organization (FAO). 1996.
- Southgate, P. & Kolkovski, S. 2000. Development of Artificial Diets for Fish Larvae. Proceeding of Hatchery Feeds. March 9-10, 2000. Cairns, Australia.



- Southgate, P. & Partridge, G. 1998. Development of Artificial Diets for Marine Finfish Larvae: Problems and Prospects. *Tropical Mariculture*. 151-170. California: Academic Press.
- Støttrup, J.G. & Norsker, N.H. 1997. Production and Uses of Copepod in Marine Fish Laviculture. Aquaculture. 155: 231-247.
- Su, H.M., Chneg, S.H., Chen, T.I. & Su, M.S. 2005. Culture of Copepods and Application to Marine Finfish Larval Rearing in Taiwan. *In* Lee, C.S., O'Bryen, P.J. & Marcus, N.H. (eds.). *Copepodes in Aquaculture*. 183-194. Oxford: Blackwell Publishing.
- Suchar, V.A. & Chigbu, P. 2006. The Effects of Algae Species and Densities on the Population Growth of the Marine Rotifer, *Colurella dicentra. Journal of Experimental Marine Biology and Ecology*. 337: 97-102.
- Szlaminska, M., Zarubov, A., Hamackova, J., Kouril, J., Vachta, R., Adamkova, I. & Munozasenio, C. 1999. Food Passage and Food Selectivity of Tench, *Tinca tinca* Larvae Fed with Zooplankton. *Fish biology*. **41**: 41-47.
- Tacon, A.G.J. & Barg, U.C. 1998. Major Challenge to Feed Development for Marine and Diadromous Finfish and Crustacean Species. In De Silva, S.S. (eds.). Tropical Mariculture . 171-208. California: Academic Press.
- Toledo, J.D., Salvacio-Golez, M., & Ohno. 2005. A. Studies of the Use of Copepods in the Semi-Intensive Seed Production of Grouper *Epinephelus coioides*. In Lee, C.S., O'Bryen, P.J. & Marcus, N.H. (eds.). *Copepodes in Aquaculture*. 169-182. Oxford: Blackwell Publishing.
- Tucker, B. T., Booth, M.A., Allen, G.L., Booth, D. & Fielder, D.S. 2006. Effect of Photoperiod and Feeding Freuency on Performance of Newly Weaned Australian Snapper, *Pagrus auratus. Aquaculture.* 258: 514-520.
- Turingan, R.G., Beck, J.L., Krebs, J.M. & Licamele, J.D. 2005. Development of Feeding Mechanisms in Marine Fish Larvae and the Swimming Behavior of Zooplankton Prey : Implications for Rearing Marine Fishes. *In* Lee, C.S., O'Bryen, P.J. & Marcus, N.H. (eds.). *Copepodes in Aquaculture*. 119-132. Oxford: Blackwell Publishing.
- Treece, G.D. & Davis, D. A. 2000. *Culture of Small Zooplanktons for the Feeding of Larval Fish*. Southern Regional Aquaculture (SRAC). 2000.
- Tzeng, W.M. & Yu, S.Y.1992. Effects of Starvation on the Formation of Daily Growth Increment in the Otoliths of Milkfish, *Chanos chanos* (Forsskal). *Journal of Fish Biology.* 40: 39.
- Walford, J., Lim, T.M. & Lam, T.J. 1991. Replacing Live Food with Microcapsulated Diets in the Rearing of Seabass, *Lates calcarifer* Larvae: Do They Ingest and Digest Protein Membrane Micro capsulate? *Aquaculture*. 92: 225-235.



- Wang, Q., Takeuchi, T., Hirota, T., Ishida, S., Miyakwa, H. & Hayasawa, H. 2004. Application of Microparticles Diets for Japanese Flounder, *Paralichthys olivaceus* Larvae. *Fisheries Science*. **70**: 611-619.
- Watanabe, W.O. 2001. Species Profile Mutton Snapper. Southern Regional Aquaculture Centre (SRAC). 2001.
- Watanabe, T. & Kiron, V. 1994. Prospects in Larval Fish Dietics. Aquaculture. 124: 223-251.
- Waynarovich, E. 1980, *Technical Assistance for Inland Fish Culture and Fishery Improvement*. Food and Agriculture Organization (FAO). 1980.
- Wilcox, J.A., Tracy, P.L. & Marcus, N.H. 2006. Improving Live Feeds: Effects of a Mixed Diet of Copepod Nauplii, *Acartia tonsa* and Rotifer on the Survival and Growth of Frist Feeding Larvae of the Southern Flounder, *Paralichthys lethostigma. Journal* of World Aquaculture Science. **37**: 113-120.
- Williams, K., Papanikos, N., Phelps, R.P. & Shardo, J.D. 2004. Development, Growth, and Yolk Utilization of Hatchery- reared Red Snapper, *Lutjanus campechanus* Larvae. *Marine Ecology Progress Series*. 275: 231-239.
- Wuenschel, M.J. & Werner, R.G. 2004. Consumption and Gut Evacuation of Laboratoryreared Spotted Sea trout Larvae and Juvenile. *Journal of Fish Biology*. 65: 723-743.
- Wyatt, T. 1972. Some Affects of Food Density on the Growth and Behavior of Plaice Larvae. *Marine Biology*. 14: 210–216.
- Yamamoto, M., Makino, H., Kobasyashi, J. & Tominaga, O. 2004. Food Organisms and Feeding Habits of Larval and Juvenile Japanese Flounder, *Paralichthya oliveceus* at Ohama Beach in Hiuchi-Nasa, the Central Seto Inland Sea, Japan. *Fisheries Science*. **70**: 1098-1105.
- Yamaoka, K., Nanbu, T., Miyagawa, M., Isshiki, T. & Kusaka, A. 2000. Water Surface Tension Related Death in Pre-Larval Red Spotted Grouper, *Epinephelus akaara*. *Aquaculture*. **189**: 165-176.
- Yilmaz, E., Bozkurt, A. & Gokcek, K. 2005. Prey Selection by African Catfish, Clarias gariepinus Larvae Fed Different Feeding Regimes. Turkey Journal of Zoo. 30: 59-66.
- Yufera, M., Fernhdez-Diaz, C. & Pascual, E. 1995.Feeding Rates of Gilthead Sea Bream, Sparus auratus Larvae on Microcapsules. Aquaculture. **134**: 257-268.

