

ANTIMICROBIAL POTENTIAL OF *Lepiota* sp. EXTRACT

TAN PEY JUAN

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(TANDATANGAN PENULIS)

(TANDATANGAN PUSTAKAWAN)

Alamat Tetap: 633, Kampung Aston,
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Pulau Pinang.

Prof. madya Dr. Markus Atong

Nama Penyelia

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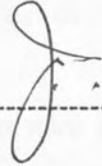


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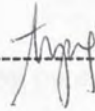
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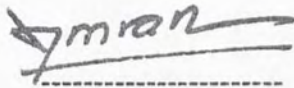
3. PEMERIKSA 2

(CIK. CHEE FONG TYNG)



4. DEKAN SEKOLAH SAINS DAN TEKNOLOGI

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ABSTRAK

Kajian telah dijalankan ke atas *Lepiota* sp. untuk menentukan potensi antimikrob cendawan tersebut dalam mengawal pertumbuhan mikrob patogen yang menyebabkan penyakit manusia khususnya penyakit kulit. Antara mikrob yang digunakan dalam ujian ini ialah *Staphylococcus aureus*, *Escherichia coli* dan *Candida albicans*. Ujian ini bertujuan untuk menentukan sama ada ekstrak *Lepiota* sp. mempunyai kesan antimikrob terhadap *Staphylococcus aureus*, *Escherichia coli* dan *Candida albicans*. Ujian ini juga bertujuan untuk menentukan faktor-faktor yang mempengaruhi keberkesanan ekstrak cendawan ini dalam aktiviti antimikrob. Antara faktor-faktor yang dikaji ialah kepekatan ekstrak, suhu penyimpanan ekstrak, tempoh masa penyimpanan ekstrak dan masa tindakan ekstrak dengan mikroorganisma. Ujian antimikrob dijalankan dengan menentukan Unit Pembentukan Koloni (UPK) mikroorganisma yang diuji. Hasil kajian yang didapati menunjukkan bahawa *Lepiota* sp. mempunyai potensi antimikrob dalam mengawal pertumbuhan *S. aureus*, *E. coli* dan *C. albicans*. Keberkesanan ekstrak dapat dilihat melalui pengurangan dalam nilai UPK mikroorganisma. Kepekatan ekstrak 350 mg/ml didapati paling berkesan dalam aktiviti antimikrob. Kesan suhu penyimpanan dan tempoh masa penyimpanan memberi kesan yang berbeza terhadap mikroorganisma yang berlainan. Dalam ujian antifungi pula, ekstrak *Lepiota* sp. dibuktikan dapat menghadkan percambahan *C. albicans*. Peratusan spora yang bercambah akan dikurangkan apabila kepekatan ekstrak bertambah. Kepekatan ekstrak tidak memberi kesan yang signifikan dalam mengawal pemanjangan tiub germa *C. albicans*.



ABSTRACT

The antimicrobial potential of *Lepiota* sp. extract in inhibiting the growth of pathogenic microbes isolated from human disease has been studied. *Staphylococcus aureus*, *Escherichia coli* and *Candida albicans* are the three microorganisms that were selected for this study. The objective of this study is to test for the antimicrobial activities of *Lepiota* sp. against *Staphylococcus aureus*, *Escherichia coli* and *Candida albicans*. This study also aimed to compare the factors that affect the effectiveness of the extract in antimicrobial activities. The factors which were considered in study are the concentrations of the extract, temperature of storage, period of storage and the mix time between the extract and the bacteria suspension. The value of Colony Forming Unit (CFU) was determined in the antimicrobial test. The results showed that *Lepiota* sp possessed antimicrobial potential in inhibiting the growth of *S. aureus*, *E. coli* and *C. albicans*. The extract was effective in decreasing the value of CFU of the microorganisms. The extract concentration of 350 mg/ml was proved to be the most effective in antimicrobial activities. The storage temperature and period of storage of the extract gave different effect to different microorganisms. In the antifungal test, *Lepiota* sp. extract gave favorable results in inhibiting the spore germination of *C. albicans*. The percentage of spore germination was reduced when the extract concentration was increased. Extract concentration has no significant effect on germ tube elongation.



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CHAPTER 1

INTRODUCTION

1.1 INTRODUCTION

Fungi are a diverse group of organisms that are classified within their own kingdom as they are neither plant nor animal (Lessee and Conte, 1996). There are estimated to be over 100,000 different fungi, most of which form only tiny threads (hypha) that can only be seen through a microscope. Fungi are thallophytes that have no chlorophyll (Svrcek, 1996). This means that they cannot manufacture their own organic food such as sugars, starches, celluloses, proteins and fats, as the green plants do (Christensen, 1972).

As presently delimited, the kingdom Fungi is believed to constitute a monophyletic group that shares some characters with animals such as chitinous structures, storage of glycogen, and mitochondrial UGA coding for tryptophan (Baier, 1991).



Being unable to produce energy through photosynthesis, fungi acquire their organic matter and energy from the decomposition of dead or live matter of other organisms. This method of getting nourishment is called heterotrophic (Baier, 1991). Fungi play a key role in the natural cycle as decomposers, returning nutrients to the soil (Lessee and Conte, 1996).

The structures of fungi are microscopic and not visible to the naked eye. Some are unicellular like yeast, but most string their cells together in long, thread-like strands called hypha. The hypha is essentially a tube with a rigid wall, containing a moving slug of protoplasm (Deacon, 1997). The shape and size of these hyphae varies in different groups of fungi, 1.5-12 μ m in diameter and from a few up to 100 μ m in length. The walls of hyphae are often strengthened with chitin, a polymer of N-acetylglucosamine. Most fungi produce an extensive system of hyphae, which may be visible when growing thickly in a mass called mycelium (commonly referred to as mould).

Hyphae can be either continuous tubes or divided by septa (walls) into individual cells, arranged in an end-to-end order. These septa are however not entirely closed; single or several openings forming a multi-perforate structure allow protoplasmic streaming. The protoplasm which fills the cells at the growing tips of the mycelium manufactures dozens of different kinds of enzymes and a number of simple and not so simple organic acids (Christensen, 1972). Mycelium can be of any size from tiny clusters to massive acre wide systems, which effectively form the feeding and growing body of the fungus.



Fungi also possess a true nucleus (Scvrek, 1996) and the reproductive unit in fungi is primarily represented by the spore. Spores may be uni or pluricellular, of many different colours and shapes, depending on the species and growth conditions. In size, spores vary from 5-20 μ m in diameter (Florian, 1993). Spores are the results of either sexual or asexual reproduction.

Asexual spore formation is often quick and profuse, making possible rapid dissemination of the fungus during favourable conditions. Sexual spore formation is a considerably more complex process, and is believed to be far more resistant since they mature more slowly and are both sexually and asexually produced spores can, if environmental conditions do not allow immediate germination, go into a state of dormancy, stasis, with a hold-over function, in which the water contents of the spore is lowered, and the metabolism inactivated but reversible. When dormancy of a spore is broken through activation, germination starts with the formation of a germ tube. Long, branching, threadlike structures, hyphae, develop from these germ tubes, eventually forming a complex mycelium.

When sporulation begins, the mature spores are very easily separated from their fruiting bodies, and due to their low weight, spreading and distribution can take place by a minute current of air. Spores can remain dispersed in air for very long time, are easily spread and therefore ubiquitous on our planet. The fungi may impose a potential health-hazard to humans causing mycoses, mycoallergies and mycotoxicoses. These diseases include invasions of the skin, lungs, eyes and brain.



All species of fungi have specific requirements of moisture and temperature before they will produce a fruit. Some mushrooms have a very restrictive set of requirements while others have very broad sets of requirements. Even under the proper climatic conditions, mushrooms may not develop if the mycelium lacks the necessary food energy. Many fungi will not fruit for several years; then one year when everything is just right, the woods will seem full of them. Other fungi with a broader range of temperature and moisture requirements appear annually.

The organisms of the fungal lineage include mushrooms, rusts, smuts, puffballs, truffles, morels, molds, and yeasts, as well as many less well-known organisms (Alexopoulos *et al.*, 1996). About 70,000 species of fungi have been described; however, some estimates of total numbers suggest that 1.5 million species may exist (Hawksworth, 1991; Hawksworth *et al.*, 1995). The mycota of Borneo has much in common with the mycotas found throughout Southeast Asia and Australia, and much less with that of Europe. The true fungi form a separate kingdom, which includes three phyla, which are Ascomycota (Cup fungi and allies), Basidiomycota (Mushrooms and allies), Zygomycota (Pin-moulds and allies), and Chytrids are often included but are not true fungi (Pegler, 1992). Placement into a division is based on the way in which the fungus reproduces sexually. The shape and internal structure of the sporangia, which produce the spores, are the most useful character for identifying these various major groups.



Fungi that will be studied in this paper have the mycological data as follows:

Kingdom: Fungi

Phylum: Basidiomycota

Class: Holobasidiomycetes

Order: Agaricales

Family: Lepiotaceae

Genus: *Lepiota*



Photograph 1.1 *Lepiota* sp.

2 cm

The Lepiotaceae was formerly considered to consist of a single genus, *Lepiota*, but some mycologists have felt that a number of changes were needed. First, there was the matter of spore colour, because one species had green spores, while the rest have white, cream, or pale buff spores. It was generally conceded that the green-spore member belonged in a separate genus, and it is now called *Chlorophyllum molybdites*..

Mushrooms in this group have white spore prints (greenish in the case of *Chlorophyllum molybdites*), free gills that are white to yellow and are a central stipe easily separable from the pileus, partial veils that typically leave a ring on the stem, and dry caps that are often scaly but do not have patches or warts. There is no volva, though one species has a rimmed basal bulb (Orr and Orr, 1979). Although there was no report on the antimicrobial activities of *Lepiota* sp, this study is still carried on to determined the antimicrobial potential of the studied fungi.

The Basidiomycota or club fungi are those fungi in which the sexual spores (basidiospores) are borne externally on special club-shaped cells, called basidia, which may develop singly or in layers known as hymenial layers (Gray, 1959). They range from common mushroom forms to some of the most important plant pathogens. In accordance with their way of life, mushrooms are divided into three basic categories:

- i) Symbiotic fungi (mycorrhizal fungi)
- ii) Fungi subsisting on dead or decaying matter (saprophytic fungi)
- iii) Fungi feeding on living organisms (parasitic fungi)



The objectives of this research are to determine the antimicrobial potential of mushroom against *Staphylococcus aureus*, *Escherichia coli* and *Candida albicans* in human disease especially skin disease.

1.2 OBJECTIVES

The objectives that are aimed to achieved are as follows:

1. To compare the antimicrobial activities in different concentrations of *Lepiota* sp. extract against *Staphylococcus aureus*, *Escherichia coli* and *Candida albicans*.
2. To compare the effect of storage temperature and period of storage of *Lepiota* sp. in antimicrobial activities.
3. To test for the effect of extract concentrations to spore germination and germ tube elongation of *C. albicans*.



1.3 HYPOTHESES EXPECTED

The hypotheses that are expected are as follows:

1. The higher concentrations of mushroom extract will have a greater antimicrobial potential.
2. For antimicrobial test, the shorter period and lower temperature of storage of the extract is more effective and the colony formed is lesser.
3. For antifungal test, higher concentrations of the extract will inhibit the spore germination and germ tube elongation of *C. albicans* more effectively.



CHAPTER 2

LITERATURE REVIEW

2.1 MEDICINAL USE OF MUSHROOMS

For hundreds of years mushrooms have excited both the minds and stomachs of millions of people. They have been used for food, medicine, religious ceremonies, and for poisoning enemies. Mushrooms have been an important part of many cultures around the world. In California some Native Americans have made soups out of shredded mushrooms. The number of mushrooms on Earth is estimated at 140,000, yet maybe only 10% (approximately 14,000 named species) are known (Christensen, 1972).

Mushrooms comprise a vast and yet largely untapped source of powerful new pharmaceutical products. In particular, and most importantly for modern medicine, they represent an unlimited source of polysaccharides with antitumor and immunostimulating properties. Several classes of compounds, such as proteins, peptides, lipopolysaccharides, glycoproteins, and lipid derivatives, have all been classified as molecules that have potent effects on the immune system (Tzianabos, 2000). Some of these compounds exert immunomodulatory and antitumor actions and are marketed



drugs. Extensive studies have revealed that a number of mushroom species are of value in the prevention a treatment of cancer, viral diseases, hypercholesterolemia, and hypertension.

Since the introduction of penicillin into therapeutics more than 50 years ago, numerous drugs with antibacterial activity have been obtained from the Actinomycetes and Bacillus (Smania *et al.*, 1995). Many of the Basidiomycetes mushrooms contain biologically active polysaccharides in fruit bodies, cultured mycelium, culture broth (Wasser, 2002). However, the resistance effect from prolonged usage of antibiotics were caused scientist to search for new source of antibiotics from various organisms including fungi. Commonly known mushrooms that are both edible and have functional properties include *Auricularia* (Mu-er), *Flammulina* (Enokitake), *Grifola* (Maitake), *Hericium*, *Lentinus* (Shiitake), *Pleurotus* (Oyster), and *Tremella* (Yiner).

Recently, many species of fungi have been found to be highly potent immune system enhancers, potentiating animals and human immunity against cancer (Feng *et al.*, 2001). Fungi such as Shittake and Agaricus have been used widely as medicinal mushroom to cure several of diseases and to enhance body immune system. Researches also show that many mushroom extractions contain antibacterial and antiviral effect against microorganisms. For example, recent study on *Pycnoporus sanguineus* has demonstrated that its extract was shown to contain a compound with biological activity against strains of *Escherichia coli*, *Klebsiella pneumoniae*, *Salmonella typhi*, *Staphylococcus aureus*, and others bacteria (Smania *et al.*, 1995).



REFERENCES

- Alexopoulos, C. J., Mims, C. W. and Blackwell, M., 1996. *Introductory Mycology 4th ed.* John Wiley and Sons, New York, USA.
- Atong, M., 1979. Kesan *in vitro* turasan *Tinospora crispa* Miers ke atas mikroorganisma dan fungsi leukosit polimorfonukleus. Unit Mikrobiologi, Universiti Kebangsaan Malaysia. 24-25.
- Baier, J., 1991. *Mushrooms and Toadstools- An Illustrated Guide.* Sunburst Books, London, 16-17.
- Boyce, T.G., Swerdlow, D.L. and Griffin, P.M., 1995. *Escherichia coli O157:H7* and the hemolytic uremic syndrome. *New England Journal of Medicine* **333**, 364-368.
- Breene, W.M., 1990. Nutritional and medicinal value of specialty mushrooms. *Journal of Food Protection* **53**, 883 -894.
- Chandrasekaran, M. and Venkatesalu, V., 2004. Antibacterial and antifungal activity of *Syzygium jambolanum*. *Journal of Ethnopharmacology* **91**, 105-108.
- Christensen, C.M., 1972. *The Molds and Man.* University of Minnesota Press, Minneapolis, 6-8.
- Cruickshank, R., Duguid, J.P., Marmion, B.P. and Swain, R.H.A., 1973. *Medical Microbiology- A Guide To The Laboratory Diagnosis And Control of Infection* 12th ed. Volume 1. Churchill Livingstone, London.
- Deacon, J.W., 1997. *Modern Mycology 3rd ed.* Blackwell Science Ltd, London WC1N2BL.



- El-Mekkawy, S., Meselhy, M.R., Nakamura, N., Tezuka, Y., Hattori, M., Kakiuchi, N., Shimotohno, K., Kawahata, T. and Otake, T., 1998. Anti-HIV-I and anti-HIV I-Protease substances from *Ganoderma lucidum*. *Phytochemistry* **49**, 1651– 1657.
- Eo, S. K., Kim, Y. S., Lee, C. K. and Han, S.S., 1999. Antiviral activities of various water and methanol soluble substances isolated from *Ganoderma lucidum*. *Journal of Ethnopharmacology* **68**, 129–136.
- Feng, W., Nagai, J. and Ikekawa, T. 2001. A clinical pilot study of EEM for advanced cancer treatment with EEM for improvement of cachexia and immune function compared with MPA. *Biotherapy* **15**, 691-696.
- Florian, M.L., 1993. Conidial Fungi (mould) Activity on artifact materials – A new look at prevention, control and eradication ICOM. *Committee for Conservation 10th Triennial Meeting*, Washington , 868–874.
- Ganiere, J.P., Medaille, C. and Etoire, F., 2004. In vitro antimicrobial activity of orbifloxacin against *Staphylococcus intermedius* isolates from canine skin and ear infections. *Research in Veterinary Science* **77**, 67-71.
- Gray, W.D., 1959. *The Relation of Fungi to Human Affairs*. Henry Holt and Company, Inc., New York, 16-21.
- Hawksworth, D. L., 1991. The fungal dimension of biodiversity: Magnitude, significance, and conservation. *Mycological Research* **95**, 641-655.
- Hawksworth, D. L., Kirk, P. M., Sutton, B. C. and Pegler, D. N., 1995. *Ainsworth and Bisby's Dictionary of the Fungi 8th ed*. CAB International, Wallingford, United Kingdom.



- Higgins, V.J. and Millar, R.L., 1968. Phytoalexin production by alfalfa in response to infection by *Colletotrichum phomoides*, *Helminthosporium turcicum*, *Stemphyllium loti* and *Stemphyllium botryosum*. *Phytopathology* **58**, 1377-1383.
- Hirasawa, M., Shoujii, N., Neta, T., Fukushima, K. and Takada, K., 1999. Three kinds of antibacterial substances from *Lentinus edobes* (Berk) Sing, (Shitake, an edible mushroom). *Int. J. of Ant. Agents* **11**, 151 -157.
- Hwang, J.S., Song, K.S., Kim, W.G., Lee, T.H., Koshino, H. and Yoo, I.D., 1997. Polyozellin, A New inhibitor of prolyl endo-peptidase from polyozellus multiplex. *Journal of Antibiotics* **50**, 773-777.
- Ito, H., Shimura, K. and Kawade, M., 1997. Antitumor effects of a new polysaccharide-protein complex (ATOM) prepared from *Agaricus blazei* (Iwade strain 101) "himematsutake" and its mechanisms in tumor-bearing mice. *Anticancer Research* **17**, 277-284.
- Jong, S. C. and Birmingham, J. M., 1992. Medicinal benefits of the mushroom *Ganoderma*. *Advances in Applied Microbiology*, **37**, 101-134.
- Jonathan, S.G. and Fasidi, I.O., 2003. Antimicrobial activities of Nigerian edible macrofungi- *Lycoperdon pusillum* (Bat. Ex) and *Lycoperdon giganteum* (Pers.). *African Journal of Biomedical Research* **6**, 85-90.
- Kasuga, A., Aoyagi, Y. and Sugahara, T., 1995. Antioxidant activity of fungus *Suillus ovinus* (L:Fr.) O.Kuntze. *Journal of Food Science* **60**, 1113 -1115.
- Kawagishi, H.A., Nomura, T., Yumen, T., Mizuno T., Hagwara, A. and Nakamura, T., 1988. Isolation and properties of lectin from the fruiting bodies of *Agaricus blazei*. *Carbohydrate Research*, **183** (1), 150-154.



- Kelmanson, J.E., Jager, A.K., and Van Staden, J., 2000. Zulu medicinal plants with antibacterial activity. *Journal of Ethnopharmacology* **69**, 241-246.
- Lerner, P.I., 1974. Susceptibility of Pathogenic *Actinomycetes* to antimicrobial compounds. *Antimicrob Agents Chemother* **5**, 302-309.
- Lessee, T. and Conte, A.D., 1996. *The Mushroom Book- Practical Know-how on Identifying, Gathering, and Cooking Wild Mushrooms and Other Fungi*. Dorling Kindresley Book, London WC2E 8PS, 6-7.
- Lin, J.M., Lin, C.C., Chen, M.F., Ujiie, T. and Takada, A., 1995. Radical scavenger and antihepatotoxic activity of *Ganoderma formosanum*, *Ganoderma lucidum* and *Ganoderma neojaponicum*. *Journal of Ethnopharmacology* **47**, 33-41.
- Mizuno, T., 1995. Bioactive biomolecules of mushrooms: Food function and medicinal effect of mushroom fungi. *Food Reviews International* **11**,7-21.
- Mizuno,T., Hagiwara,T., Nakamura,T., Ito,H., Shimura,K., Sumiya,T. and Asakura, A.,1990. Antitumor activity and some properties of water-soluble polysaccharides from "himematsutake", the fruiting body of *Agaricus blazei* Murill. *Agricultural and Biological Chemistry* **54**, 2889-2896.
- Moffet, H.L., 1980. *Clinical Microbiology 2nd ed.* J.B. Lippincott Company, Toronto.
- Nanba, H. and Kuroda, H., 1987. Antitumor mechanisms of orally administered Shiitake fruit bodies. *Chemical and Pharmacological Bulletin* **35**, 2459- 2464.



- Ng, T. B., 1998. A review of research on the protein-bound polysaccharide (Polysaccharopeptide, PSP) from the mushroom *Coriolus versicolor*. *General Pharmacology* **30**, 1-4.
- Ngai, H.K. and NG, T.B., 2004. A mushroom (*Ganoderma capense*) lectin with spectacular thermostability, potent mitogenic activity on splenocytes, and antiproliferative activity toward tumor cells. *Biochemical and Biophysical Research Communication* **314**, 988-993.
- Odds, F. C., 1979. *Candida and Candidosis*. London, Leicester University Press.
- Oloke, J.O. and Kolawole, D.O., 1998. The antibacterial and antifungal activities of certain components of *Aframomum melegueta* fruits. *Fitoterapia* **59** (5), 384-388.
- Orr, R.T. and Orr, D.B., 1979. *Mushrooms of Western North America*. University of California Press, Ltd, London.
- Pegler, D. and Spooner, B., 1992. *The Mushroom Identifier*. Apple Press, London N79BH, 12-13.
- Piraino, F., Brandt, C.R., 1999. Isolation and partial characterization of an antiviral, RC-183, from the edible mushroom *Rozites caperata*. *Antiviral Research* **43**, 67-68.
- Seong, K.E., Young, S.K., Chong, K.L. and Seong, S.H., 1999. Antiviral activities of various water and methanol soluble substances isolated from *Ganoderma lucidum*. *Journal of Ethnopharmacology* **68**, 129-136.



- Shahidi, F., and Amarowicz, R., 1996. Antioxidant activity of protein hydrolyzates from aquatic species. *Journal of the American Oil Chemists' Society* **73**, 1197 -1199.
- Smania, A., Monache, F.D., Smania, E.F.A., Gil, M.L., Benchetrit, L.C. and Cruz, F.S.,1995. Antimicrobial activity of substance produced by the fungus *Pycnoporus sanguineus* (Fr.) Murr. *Journal of Ethnopharmacology* **45**, 177-181.
- Smania, A., Monache, F.D., Loguericio-Leite, C., Smania, E.F.A. and Gerber, A.L., 2001. Antimicrobial activity of Basidiomycetes. *International Journal of Medicinal Mushrooms* **3**, 87.
- Suay, I., Arenal, F., Asenio, F., Basilio, A., Cabello, M. and Diez, M.T., 2000. Screening of Basidiomycetes for antimicrobial activities. *Antonie van Leeuwenhoek* **78**, 129-39.
- Svrcek, M., 1996. *A Field Guide in Colour to Mushrooms*. Aventinum Publishing House, Leicester.
- Tarr, P.I., 1995. *Escherichia coli O157:H7*: Clinical diagnostic and epidemiological aspects of human infection. *Clinical Infectious Diseases* **20**, 1-10.
- Toda, M., Okubu, S., Ikigai, H., Suzuki, T., Suzuki, Y. and Shimamura, T., 1991. The protective activity of tea against infection by *vibro cholerae*. *J. Appl. Bacteriol* **70**, 109 -112.
- Tong, L.M., Sasaki, S., McClements, D.J. and Decker, E.A., 2000. Mechanisms of the antioxidant activity of a high molecular weight fraction of whey. *Journal of Agricultural and Food Chemistry* **48**, 1473 -1478.



- Tzianabos, A., 2000. Polysaccharide immunomodulators as therapeutic agents: Structural aspects and biologic function. *Clinical Microbiology Reviews* **13**, 523-533.
- Victor, L., 1980. *Antibiotics in Laboratory Medicine*. The Williams and Wilkins Company, USA.
- Wang, H.X., Ng, T.B., Ng, V.E. and Ooi, C., 1998. Lectins from mushrooms - A review. *Mycology Research* **102**, 897-906.
- Wasser, A.L. and Weis, A.L., 1999. Medicinal properties of substances occurring in higher Basidiomycetes mushrooms: Current perspectives (review). *International Journal of Medicinal Mushrooms* **1**, 31-62.
- Wasser, S., 2002. Medicinal mushrooms as a source of antitumor and immunomodulating polysaccharides. *Applied Microbiology and Biotechnology* **60** (3), 258-274.
- Yun, S.S., Sun, H.K., Jae, H.S., Chang, B.J. and Eun, H.P., 2004. Anti-angiogenic and inhibitory activity on inducible nitric oxide production of the mushroom *Ganoderma lucidum*. *Journal of Ethnopharmacology* **90**, 17-20.
- Zhu, J. S., Halpern, G. and Jones, K., 1998. The scientific rediscovery of an ancient chinese herbal medicine: *Cordyceps sinensis* *The Journal Of Alternative And Complementary Medicine* **4**(3), 289.

