ANTIBACTERIAL AND ANTIOXIDANT ACTIVITY OF ALOE VERA

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ABSTRAK

AKTIVITI-AKTIVITI ANTIBAKTERIA DAN ANTIOKSIDAN ALOE VERA

Objektif am bagi kajian ini adalah mengaji potensi aktiviti-aktiviti antibakteria dan antioksidan tentang ekstrak aloe vera. Fenolik, flavonoid dan antrakuinon diekstrak dari kedua-dua daun Aloe yuenjiangensis dalam bentuk segar dan kering, masingmasing. Ekstrak fenolik dari daun dalam bentuk segar and bentuk kering tidak menunjukkan sebarang aktiviti antibakteria, manakala ekstraks flavonoid dari daun segar dan antrakuinon dari kedua-dua daun segar and kering menunjukkan aktiviti antibakteria terhadap patogen makanan serta bakteria perosak makanan melalui kaedah penyebaran disk, Kepekatan penyekatan minimum ekstrak ditentukan dengan kaedah dalam kalang dari 3.90 ke 41.1 mg/ml manakala kepekatan pembasmian minimum ekstrak dikalang dari 3.90 ke 50.1mg/ml. Kandungan jumlah fenolik setiap ekstrak dikira dengan reagen Folin-Ciocalteau, Aktiviti antioksidan ekstraks Aloe yuenjiangensis dinilai dengan ujian pembangkaian radikal 2,2-diphenyl-1picrylhydrazyl (DPPH), kesan chelating ion ferum dan sistem asid linoleik beta karotin. Kandungan jumlah fenolik berbeza di antara daun mentah dan daun kering, dan sama juga dengan kaedah pengekstrakan yang berlainan. Ekstraks anthraquinone daun kering menunjukkan signifikan yang lebih terhadap kedua-dua jenis radikal larut air dan lemak dalam ujian pembangkaian radikal DPPH dan sistem asid linoleik beta karotin, di mana dirujuk kepada kandungan jumlah fenolik yang terdapat dalam ekstraks. Kesan chelating ion ferum menunjukkan perbezaan signifikan dalam ekstraks fenolik kering (p<0.05). Dalam kesimpulan, keputusan menunjukkan ektraks Aloe yuenjiangensis mempunyai kapasiti untuk membangkai radikal bebas dan merencat pertumbuhan kedua-dua jenis bakteria patogen dan perosak makanan. Namun, ektraks ini adalah sesuai untuk digunakan sebagai agen antimikrobia dan antioksidan semulajadi yang berasal dari tumbuhan dalam industri makanan.



ABSTRACT

ANTIBACTERIAL AND ANTIOXIDANT ACTIVITY OF ALOE VERA

The main objective of the study was to examine the potential of antibacterial and antioxidant activities of aloe vera extracts. Phenolic, flavonoid and anthraquinone extracted from fresh and dried leaves of Aloe yuenjiangensis, respectively. Phenolic extracts from fresh and dry sample show no antibacterial activity, whereas flavonoid extract from fresh form of sample and anthraquinone extracts from fresh and dry form of sample against Staphylococcus aureus S277, Escherichia coli IMR E91/02C, Pseudomonas aeruginosa ATCC 10145 and Bacillus cereus exhibited antimicrobial activity through disk diffusion assay. The minimum inhibitory concentrations (MICs) of extracts determined by the broth dilution method ranged from 3.90 to 41.1 mg/ml while the minimum bacteriacidal concentrations (MBCs) of extracts ranged from 3.90 to 50.1mg/ml. Total phenolic content of each extract was quantified with the Folin-Ciocalteau reagent, Antioxidant activities of Aloe yuenjiangensis extracts were evaluated using 2,2-diphenyl-1-picrylhydrazyl (DPPH) radical scavenging assay, ferrious ion chelating effect and B-carotene linoleic acid system. Total phenolic content varied between fresh and dried leaves, as well as different extraction methods. Dried anthraquinone extracts exhibited significantly (p<0.05) greater DPPH radical scavenging assay and beta carotene linoleic acid system towards both lipidand water-soluble radicals, which was attributed to the total phenolic content. Ferrous ion-chelating effect was significantly (p < 0.05) greater in the dried phenolic extracts. In conclusion, the results indicate that the extracts of Aloe yuenjiangensis have the capacity to scavenge free radicals and to inhibit the growth of both spoilage and pathogenic bacteria. Therefore they could be suitable for using as natural plantderived antimicrobial and antioxidant agents in the food industry.



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LIST OF ABBREVIATION

AAPH	2,2'-azobis- (2-amidinopropane) dihydrochloride
ABTS	2,2'-azino-bis-(3-ethylbenzthiazoline-6-sulfonic
aw	water activity
BHA	butylated hydroxyanisole
BHT	butylhydroxytoluene
CFU	Colony form unit
DPPH	1,1-Diphenyl-2-picrylhydrazyl or 2,2-Diphenyl-1-picrylhydrazyl
EDTA	Ethylenediaminetetraacetic acid
EPR	electron paramagnetic resonance
ET	electron transfer
Fe ²⁺	ferric ion (II)
FeCl ₂ · 4H ₂ O	ferric chloride hydrated
FRAP	ferric ion reducing antioxidant parameter
GC	Gas chromatography
GRAS	Generally recognized as safe
НАТ	hydrogen atom transfer
HPLC	high performance liquid chromatography
IASC	Institute Aloe Science Council
IMR	Institute of Medical Research
LDL	low density lipoprotein
MIC	Minimal inhibitory concentrations
ORAC	Oxygen radical absorbance capacity
SPSS	Statistical Packages for Social Science
TBHQ	tert-butylhydroquinone
TEAC	Trolox equivalent antioxidant capacity



- TSA tryptic soy agar
- TSB tryptic soy broth
- WHO World Health Organization

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LIST OF SYMBOL

%	percent
α	alpha
β	beta
°C	degree Celsius
kg	kilogram
g	gram
mg	microgram
μg	microgram
ml	milliliter
μL	microliter
mM	milimolarity
Acontrol	Absorbance of control
Atest	absorbance of test
t	time
±	positive and negative
>	more than
<	less than



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CHAPTER 1

INTRODUCTION

The spoilage and poisoning of foods by microorganisms is a problem that has not yet been brought under adequate control despite the range of roust preservation techniques available (Rauha et al., 2000). Foodborne diseases are still a major problem in the world, even in well-developed countries. A variety of microorganisms also lead food spoilage that is encountered as one of the most important matter concerning food industry. So far, many pathogenic microorganisms, such as Escherichia coli, Staphylococcus aureus, Listeria monocytogenes, or non pathogenic microorganisms such as Pseudomonas spp. and Lactobacillus spp. have been reported as the causal agents of foodborne diseases and/or food spoilage (Sokmen et al., 2004). Besides microbial spoilage and contamination, chemical spoilage is also another problem occur in food industry. For an example, rancidity rapidly appears in high fat foods such as bakery products and edible oils. Therefore, synthetic antioxidants, such as butylated hydroxyanisole (BHA), butylated hydroxytoluene (BHT), and tertiary butyhydroquinone (TBHQ) have been added as inhibitors of lipid oxidation and stabilizing fat containing food stuffs. However due to possible toxic and carcinogenic effects on health, their usage are being questioned (Amarowiczet et al., 2004; Hu et al., 2003; Sokmen et al., 2004).

Consumers increasingly avoid foods prepared with preservatives of chemical origin. A desire for high quality foods that are more natural, minimally processed and



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yet preservative free is highly demanding (Fitzgerald *et al.*, 2003). Natural alternatives are required to prolong shelf life of foods as well as provide a high degree of safety with respect to foodborne pathogenic microorganisms. The application of naturally derived plant extracts to fulfill this aim of growing interest has challenged food industry to increase research on bioactive components from plants (Baydar *et al.*, 2004).

Phytochemicals occurring in herb that play a significant role in disease prevention and health promotion was first recognized as a result of epidemiological studies using both animal and human subjects (Shahidi & Naczk, 2004). This led to an ever-growing interest in herbs or botanicals and nutraceutical products. Bioactives in herbal and nutraceutical products constitute a myriad of chemical compounds, among which phenolic substances often play a primary or a synergistic function. Phytochemicals exhibited various degrees of antimicrobial activity occur in plant after molecular separation techniques to the isolation of various compounds (Vinha *et al.*, 2005; Karadeniz *et al.*, 2000; Yao *et al.*, 2003; Chavan *et al.*, 2003; Tsao & Deng, 2004). The increased demand for minimally processed foods or food with extended shelf-life has further revived interest in exploitation of these natural antimicrobial agents.

Phenolic compounds, at low concentration, may act as an antioxidant in our diet (Blasa *et al.*, 2006) and protect foods from oxidative deterioration. However, at high concentrations, they or their oxidation products may interact with proteins, carbohydrates, and minerals. Phenols are important compounds because of their contribution to human health and their multiple biological effects, such as



antioxidant activity, antimutagenic or anticarcinogenic activities, and antiinflammatory action (Karakaya, 2004).

Antioxidant compounds include vitamins, phenols, carotenoids, and flavonoids. These antioxidants may prevent the incidence of cardiovascular disease (Pastrana-Bonilla et al., 2003). Over 4000 different flavonoids have been described and they are categorized into several subgroups. The four major classes of flavonoid are the 4-oxoflavonoids (flavones, flavonols and others.), anthocyanins, isoflavones, and the flavan-3-ol derivatives (catechin and tannins) and exhibit substantial antioxidant activity (Rhodes & Price, 1996). Flavonoids are a group of C15 aromatic plant pigments, which are biosynthesized via a confluence of the acetate/malonate and shikimate pathways. They are group of natural benzo-y-pyran derivatives and are ubiquitous in photosynthesizing cells and they occur as aglycones, glycosides and methylated derivatives (Skerget et al., 2005). Flavonols and flavones are usually found in plants bound to sugars as O-glycosides. Flavones may also occur as Cglycosides. The glycosidic form is a general feature of flavonoids, with the exception of flavanols where glycosides are rare. Free flavonoids, flavonoids without their attached sugars, are called aglycones. Aglycones of flavonols and flavones are not present in fresh plants but may be present as a result of food processing (Hollman & Arts, 2000). A single plant may contain different flavonoids, and their distribution within a plant family could be useful in the taxonomy. In plants, the flavonoids occur as white and yellow pigments in flowers, fruits, barks and roots and because of their favorable UV-absorbing properties, they also protect the plant from harmful UV radiation from the sun (Naidu et al., 2000). These polyphenolic compounds are widely found in various types of edible plants, especially in vegetables, fruits, and tea. Flavonols (quercetin, myricetin and kaempferol) and flavones (apigenin and



luteolin) are the most common phenolics in plant-based foods. Flavanones are typically present in citrus fruit, and flavanols in green tea (Puupponen-Pimiä *et al.*, 2001).

The antimutagenic (Park *et al.*, 2004), antibacterial (Turkoglu *et al.*, 2006), antiviral (Du *et al.*, 2003; Evers *et al.*, 2005), anticarcinogenic (Merken & Beecher, 2000), anti-inflammatory and antithrombotic actions (Di Carlo *et al.*, 1999) of flavonoids are well characterised. Flavonoids can act as vasodilators and platelet disaggregators and also possess efficient antioxidant and free radical scavenging abilities (Bahorun *et al.*, 2004). Acylation of anthocyanins with *p*-coumaric and caffeic acids is common in fruits, and it is responsible for a better color stability in food products (Mazza & Miniati, 1993). Among numerous substances identified in medicinal plants, flavonoids represent one of the most interesting classes of bioactive compounds.

Anthraquinone is one of the members in quinine group under the big group of phenolics (Cowan, 1999). Quinones are aromatic rings with two ketone substitutions. They present everywhere in nature and are characteristically highly reactive. These compounds, being colored, are responsible for the browning reaction in cut or injured fruits and vegetables and are an intermediate in the melanin synthesis pathway in human skin (Schmidt, 1988). In addition to provide a source of stable free radicals, quinones are known to irreverse nucleophilic amino acids in proteins (Stern et al., 1996), often leading to inactivation of the protein and loss of function.



The potential range of quinone antimicrobial effects is great. Probable targets in the microbial cell are surface-exposed adhesins, cell wall polypeptides, and membrane-bound enzymes (Cowan, 1999). For all the plant-derived antimicrobials included quinone, the possibility of toxic effects must be thoroughly examined before it is applied in food system. Anthraquinone included emodin, rhein and physcion that isolated from *Cassia tora* seed inhibited phytopathogenic fungi such as *Pyricularia grisea, Botrytis cinerea, Phytophythora infestans* and *Erysiphe graminis* (Kim *et al.*, 2004). Since anthraquinone was proofed to exhibit anti-inflammatory and immunomodulatory effects of anthraquinones, the mechanism was believed involves antioxidant. Therefore, anthraquinones may act as antioxidants and radical scavengers (Choi & Chung, 2003).

Aloe vera is a member of liliaceae family and has been widely applied for centuries as traditional garden herbs. There are more than 360 different species of aloes grown in the dry regions of North American, Europe, and Asia (Hu *et al.*, 2003). Aloe is a succulent plant which is xerophytes and is adapted to live in low water availability area and is characterized by possessing a large water storage tissue. Aloe succulents is one of the xerophytates use crassulacean acid metabolism, an additional photosynthetic pathway involving malic acid (Ni *et al.*, 2004) that commonly found in cacti to store simple sugar and organic compounds in stomata. Of over 300 Aloe species, Aloe vera Linn. is most widely accepted and used for various medical, cosmetic and nutraceutical purposes. In Japan, *Aloe arborescens* Miller *var. natalensis* Berger is used as a folk remedy, and *Aloe barbadensis* Miller attracts much attention as a health food (Okamura *et al.*, 1996). Aloe Vera products have long been used in health foods for medical and preservative purposes. In the food industry, Aloe vera has been utilized as a source of functional food, especially



PERPUSTAKAAN UNIVERSITI MALAYSIA SABAH for the preparation of health drinks and other beverages, including tea yogurt and fruit juice. Bottled aloe vera juice or gel is widely available for internal consumption as a tonic, and it has been claimed to cure many illness, such as gout, constipation and arthritis (Eshun & He, 2004). It has been also applied in pharmaceutical industry due to its positive effect on the inhibition of tumor growth and fight against various skin disorders. An extensive application of aloe vera gel in the cosmetic and toiletry industries was found because of its valuable moisturizing emollient effect and widely applied in creams, cleansers and soups (Eshun & He, 2004). It has been reported to posses immunomodulatory, antiimflammatory, UV protective, antiprotozoal and wound and burn healing promoting properties (Choi & Chung, 2003).

Aloe vera is a rich source of polysaccharides and has various carbohydrate constituents. Mannose-6-phosphate is the major sugar in aloe vera gel. As it is linked to a protein and form mucopolysaccharide, it may produce even greater wound-healing effects (Grey *et al.*, 1991). Aloe vera contains anthraquinone as its major active principles hydroxyanthrone derivatives, mainly of the aloe-emodin-anthrone 10-*C*-glucoside type. The major constituent is known as barbaloin (aloin) (15–40%). It also contains hydroxyaloin (about 3%). Barbaloin (= aloin) is in fact a mixture of aloin A (10*S*) and B (10*R*). Another aloe species origin from Japan, *Aloe ferox* also contains aloinoside A and B. Aloin A and B interconvert through the anthranol form as do aloinoside A and B. The yellow exudates from the inner epidermal cell layers are known for its purgative activity. Purgative principles from aloe have been identified as an anthrone-C-glucosyl, barbaloin (aloin A) and homonataloin (Okamura *et al.*, 1996).



Hu *et al.* (2003) suggest that growth stage plays a vital role in the composition and antioxidant activity of Aloe vera. Free radical-scavenging activity of Aloe vera (*Aloe barbadensis* Miller) extracts in aloatic epidermis is stronger than pulp as more active free radical scavengers were in the skin (Hu *et al.*, 2005). Components in the rind of Aloe vera are responsible for the higher antioxidant activity of Aloe vera extracts. In fact, there are many cases reported that rind and pomace of plant are higher in phenolic content than pulp or flesh that commonly served as edible part during processing. They can be utilized as good source to recycle for extraction of polyphenol. Grape seed and skin (Baydar *et. al.* 2004), apple pomace (Lu & Foo, 2000), almond skin (Sang *et al.*, 2002), peanut skin (Nepote *et al.*, 2005) are examples of it.

Previous studies on aloe vera are focused on their health contribution to human body (Borrelli & Izzo, 2000; Choi *et al.*, 2001; Schiller, 2001; Eshun & He, 2004) but potential of aloe vera as preservative for food safety is lacking of focus (Hu *et al.*, 2003; Valverde *et al.*, 2005). Freeze dried aloe vera extracts are high in total flavonoid content and exhibit strong antioxidant activity (Hu *et al.*, 2003). Flavonoid of aloe vera plant has not been determined for its potential to exhibit antioxidant activities as well as antimicrobial activities. Anthraquinone give a promising effect against fungi (Jasso de Rodriguez *et al.*, 2005; Ali et al., 1999) and inhibit a possible causative factor of gastric cancer, *Helicobacter pylori* (Choi & Chung, 2003). Information of aloe vera against food spoilage bacteria and foodborne pathogen is still not available. Thus, in this study was carried out to determine antibacterial as well as antioxidant activities of targeted groups of compounds of aloe vera.



REFERENCE

- Aburjai, T. & Natsheh, F. M., 2003. Plants used in cosmetics. *Phytotherapy Research*. 17: 987-1000.
- Ahmad, R., Shaari, K., Lajis, N. H., Hamzah, A.S., Ismail, N.H. & Kitajima, M., 2005. Anthraquinones from *Hedyotis capitellata*. *Phytochemistry*. **66**: 1141-1147.
- Ali, M.I.A., Shalaby, N.M.M., Elgamal, M.H.A. & Mousa, A.S.M., 1999. Antifungal Effects of Different Plant Extracts and their Major Components of Selected Aloe Species. *Phytotherapy Research.* 13: 401–407.
- Alves, D.S., Perez-Fons, L., Estepa, A. & Micol, V., 2004. Membrane-related effects underlying the biological activity of the anthraquinones emodin and barbaloin. *Biochemical Pharmacology.* 68: 549–561.
- Ames, B.N., Shigenga, M.K. & Hagen, T.M., 1993. Oxidants, antioxidants and degenerative diseases of ageing. *Proceedings of National Academy of Sciences* USA. 90: 7915-7922.
- An, M., Haig, T. & Hatfield, P., 2001. On-site sampling and analysis of fragrance from living lavender (*Lavendula augustiflora* L.) flowers by solid phase microextraction coupled to gas chromatography and ion trap mass spectrometry. *Journal of Chromatography A.* 917: 245-250.
- Arias, M.E., Gomeza, J.D., Cudmani, N.M., Vattuone, M.A. & Isla, M.I., 2004. Antibacterial activity of ethanolic and aqueous extracts of Acacia aroma Gill. ex Hook et Arn. *Life Sciences.* **75**: 191–202.
- Arts, M. J. T. J., Dallinga, J. S., Voss, H.P., Haenen, G. R. M. M. & Bast, A., 2003. A critical appraisal of the use of the antioxidant capacity (TEAC) assay in defining optimal antioxidant structures. *Food Chemistry*, 80: 409–414.
- Arts, M. J. T. J., Haenen, G. R. M. M., Voss, H.P., & Bast, A., 2001. Masking of antioxidant capacity by the interaction of flavonoids with protein. *Food and Chemical Toxicology*. **39**: 787–791.
- Arts, M. J. T. J., Haenen, G. R. M. M., Voss, H. P. & Bast, A., 2004. Antioxidant capacity of reaction products limits the applicability of the Trolox equivalent antioxidant capacity (TEAC) assay. *Food Chem. Toxicol.* **42**: 45-49.

Atherton P., 1998. Aloe vera: magic or medicine? Nurs Stand. 12: 49–52, 54.



- Atta-ur-Rahman & Choudhary, M.I., 1995. Diterpenoid and steroidal alkaloids. Nat. Prod. Rep. 12: 361–379.
- Bahorun, T., Luximon-Ramma, A., Crozier, A. & Aruoma, O., 2004. Total phenol, flavonoid, proanthocyanidin and vitamin C levels and antioxidant activities of Mauritian vegetables. *Journal of the Science of Food and Agriculture*. 84: 1553– 1561.
- Bassole, I.H.N., Ouattara, A.S., Nebie, R., Ouattara, C.A.T., Kabore, Z.I. & Traore, S.A., 2003. Chemical composition and antibacterial activities of the essential oils of *Lippia chevalieri* and *Lippia multiflora* from Burkina Faso. *Phytochemistry.* 62: 209–212.
- Baydar, N.G., Ozkan, G. & Sagdic, O., 2003. Total Phenolic contents and antibacterial activities of frappe (*Vitis vinifera* L.) extracts. *Food Control.* 15: 335-339.
- Beppu, H., Kawai, K., Shimpo, K., Chihara, T., Tamai, I., Ida, C., Ueda, M. & Kuzuya, H., 2004. Studies on the components of *Aloe arborescens* from Japan—monthly variation and differences due to part and position of the leaf. *Biochemical Systematics and Ecology.* **32**: 783–795.
- Bouchey, G.D. & Gjerstad, G., 1969. Chemical studies of Aloe vera juice. Quarterly Journal of Crude Drug Research. 9:1445–1453.
- Blitz, J., Smith, J.W. & Gerard, J.R., 1963. Aloe vera gel in peptic ulcer therapy: preliminary report. Journal of the American Osteopathic Association. 62: 731– 735.
- Bondet V., Brand-Williams, W. & Berset, C., 1997. Kinetics and mechanisms of antioxidant activity using the DPPH free radical method. *Lebensm Wiss Technol.* 30: 609–615.
- Boussouel, N., Mathieu, F., Revol-Junelles, A.M. & Milliere, J.B., 2000. Effect of combinations of lactoperoxidase system and nisin on the behaviour of *Listeria monocytogenes* ATCC 15313 in skim milk. *International Journal of Food Microbiology*. 61: 169–175.
- Boziaris, I.S. & Adams, M.R., 2001. Temperature shock, injury and transient sensitivity to nisin in Gram negatives. *Journal of Applied Microbiology*. 91: 715–724.
- Brune, M., Hallberg, L. & Skanberg, A., 1991. Determination of iron-binding phenolic groups in foods. J. Food Sci. 56: 128–131.
- Budu-Amoako, E., Ablett, R.F., Harris, J. & Delves-Broughton, J., 1999. Combined effect of nisin and moderate heat on destruction of *Listeria monocytogenes* in coldpack lobster meat. *Journal of Food Protection.* 62: 46–50.



- Burt, S.A. & Reinders, R.D., 2003. Antibacterial activity of selected plant essential oils against *Escherichia coli* O157:H7. *Letters in Applied Microbiology*. **36**: 162–167.
- Burt, S., 2004. Essential oils: their antibacterial properties and potential applications in foods-a review. *International Journal of Food Microbiology*. **94**: 223-253.
- Cao, G., Verdon, C. P., Wu, A. H. B., Wang, H. & Prior, R. L., 1995. Automated assay of oxygen radical absorbance capacity with COBAS FARA II. *Clin. Chem.* 41: 1738-1744.
- Capellas, M., Mor-Mur, M., Gervilla, R., Yuste, J. & Guamis, B., 2000. Effect of high pressure combined with mild heat or nisin on inoculated bacteria and mesophiles of goat's milk fresh cheese. *Food Microbiology*. **17**: 633–641.
- Carson, C.F. & Riley, T.V., 1995. Antimicrobial activity of the major components of the essential oil of *Melaleuca alternifolia*. Journal of Applied Bacteriology. **78**: 264– 269.
- Carson, C.F., Hammer, K.A. & Riley, T.V., 1995. Broth microdilution method for determining the susceptibility of *Escherichia coli* and *Staphylococcus aureus* to the essential oil of *Melaleuca alternifolia* (tea tree oil). *Microbios.* 82:181–185.
- Chang, X.L., Wang, C., Feng, Y. & Liu, Z., 2005. Effects of heat treatment on the stabilities of polysaccharides substances and barbaloin in gel juice from *Aloe vera* Miller. *Journal of Food Engineering*. **75**: 245-251.
- Chavan, U.D., McKenzie, D.B., Amarowicz, R. & Shahidi, F., 2003. Phytochemical components of beach pea (*Lathyrus maritimus* L.). Food chemistry. 81: 61-71.
- Chen, Y., Wang, M. F., Rosen, R. T., & Ho, C. T., 1999. 2,2-Diphenyl-1-picrylhydrazyl radical-scavenging active components from *Polygonum multiflorum Thunb. Journal of Agricultural and Food Chemistry*. 47: 2226–2228.
- Chithra, P., Sajithlal, G.B. & Chandrakasan, G., 1998. Influence of Aloe vera on the glycoaminoglycans in the matrix of healing dermal wounds in rats. Journal of Ethnopharmacology. 59: 179–186.
- Choi, J. S., Lee, H. J., Park, K. Y., Ha, J. O. & Kang, S. S., 1997. In Vitro antimutagenic effects of anthraquinone aglycones and naphthopyrone glycosides from Cassia tora. Planta Medica. 63: 11-14.
- Choi, S.W., Son, B.W., Son, Y.S., Park, Y.I., Lee, S.K. & Chung, M.H., 2001. The woundhealing effect of a glycoprotein fraction isolated from *Aloe vera*. *The British Journal of Dermatology* .145: 535–545.
- Choi S. W. & Chung, M. Y., 2003. A Review on the Relationship between Aloe vera Components and Their Biologic Effects. Seminars in Integrative Medicine. 1: 53-62.



Clarke, M.P., 1999. Can food forestall ageing. Agriculture Research USDA. 2: 15-17.

- Connor, A. M., Luby, J. J., Hancock, J. F., Berkheimer, S. & Hanson, E. J., 2002. Changes in fruit antioxidant activity among bluberry cultivars during coldtemperature storage. J. Agric. Food Chem. 50: 893-898.
- Cosby, D.A., Harrison, M.A., Toledo, R.T. & Craven, S.E., 1999. Vacuum or modified atmosphere packaging and EDTA–nisin treatment to increase poultry product shelf life. *Journal of Applied Poultry Research.* 8: 185–190.
- Cosentino, S., Tuberoso, C.I.G., Pisano, B., Satta, M., Mascia, V., Arzedi, E. & Palmas, F., 1999. In vitro antimicrobial activity and chemical composition of Sardinian *Thymus* essential oils. *Letters in Applied Microbiology*. **29**: 130–135.
- Coskun, M., Satake, T., Hori, K., Saiki, Y. & Tankr, M., 1990. Anthraquinone glycosides from *Rhamnus libanoticus*. *Phytochemistry*. **29**: 2018-2020.
- Cox, S.D., Mann, C.M., Markham, J.L., Bell, H.C., Gustafson, J.E., Warmington, J.R., Wyllie, S.G., 2000. The mode of antimicrobial action of essential oil of *Melaleuca alternifola* (tea tree oil). *Journal of Applied Microbiology*. 88: 170–175.
- Cowan, M. M., 1999. Plant Products as Antimicrobial Agents. *Clinical Microbiology Reviews.* **12**: 564–582.
- Cressy, H.K., Jerrett, A.R., Osborne, C.M., Bremer, P.J., 2003. A novel method for the reduction of numbers of *Listeria monocytogenes* cells by freezing in combination with an essential oil in bacteriological media. *Journal of Food Protection*. 66: 390–395.
- Cutter, C.N. & Siragusa, G.R., 1995. Population reductions of Gram-negative pathogens following treatments with nisin and chelators under various conditions. *Journal* of Food Protection. 58: 977–983.
- Dagne, E., Yenesew, A., Asmellash, S., Demissew, D. & Maw, S., 1994. Anthraquinones, preanthraquinones and isoeleutherol in the roots of aloe species. *Phytochemistry*, 35: 401-466.
- Davis, R.H., Leitner, M.G., Russo, J.M. & Byrne, M.E., 1989. Anti inflammatory activity of Aloe vera against a spectrum of irritants. Journal of the American Podiatric Medical Association. 79: 263–276.
- Deans, S.G., Simpson, E., Noble, R.C., MacPherson, A. & Penzes, L., 1993. Natural antioxidants from *Thymus vulgaris* (thyme) volatile oil: the beneficial effects upon mammalian lipid metabolism. *Acta Horticulturae*. 332: 177–182.
- Delaquis, P.J., Stanich, K., Girard, B. & Mazza, G., 2002. Antimicrobial activity of individual and mixed fractions of dill, cilantro, coriander and eucalyptus essential oils. *International Journal of Food Microbiology*. **74**: 101–109.



- Deshpande, S.S. & Cheryan, M., 1987. Determination of phenolic compounds of dry beans using vanillin, redox and precipitation assays. *Journal of Food Science*. 52: 332-334.
- Dorantes, L., Colmenero, R., Hernandez, H., Mota, L., Jaramillo, M.E., Fernandez, E. & Solano, C., 2000. Inhibition of growth of some food-borne pathogenic bacteria by *Capsicum annum* extracts. *International Journal of Food Microbiology*. 57: 125–128.
- Dorman, H.J.D. & Deans, S.G., 2000. Antimicrobial agents from plants: antibacterial activity of plant volatile oils. *Journal of Applied Microbiology*. 88: 308–316.
- Dragsted, C.O., Strube, M. & Larsen, L.C., 1993. Cancer protective factors in fruits and vegetables. Biochemical and biological background. *Pharmacology and Toxicology*. 72: 116-135.
- Duh, P.D., Yen, D.B. & Yen, G.C., 1992. Extraction and identification of an antioxidative component from edible oils. *Food Chemistry*. 14: 45-51.
- Dziezak, J.D., 1986. Preservatives., Antioxidants. The ultimate answer to oxidation. Food Technology. 40 (9): 94-102.
- Elamthuruthy, A. T., Shah, C.R., Khan, T.A., Tatke, P.A. & Gabheb, S.Y., 2005. Standardization of marketed Kumariasava—an Ayurvedic *Aloe vera* product. *Journal of Pharmaceutical and Biomedical Analysis.* **37**: 937–941.
- Elgayyar, M., Draughon, F.A., Golden, D.A. & Mount, J.R., 2001. Antimicrobial activity of essential oils from plants against selected pathogenic and saprophytic microorganisms. *Journal of Food Protection.* 64: 1019–1024.
- Eichhorn, S. & Winterhalter, P., 2005. Anthocyanins from pigmented potato (Solanum tuberosum L.) varieties. Food Research International. 38: 943–948.
- Ekstrand, B. 1994. Lactoperoxidase and lactoferrin. V.M. Dillon and R.G. Board (ed). Natural antimicrobial systems and food preservation. Wallingford: CAB International. 15-63.
- Eloff, J.N.1998. Which extractant should be used for the screening and isolation of antimicrobial components from plants? *Journal of Ethnopharmacology*. 60:1-8.
- Ennahar, S., Assobhel, D. & Hasselmann, C., 1998. Inhibition of *Listeria monocytogenes* in a smear-surface soft cheese by *Lactobacillus plantarum* WHE92, a pediocin AcH producer. *Journal of Food Protection.* **61**: 186–191.
- Erdogrul, O.T., 2000. Antibacterial activities of some plant extracts used in folk medicine. *Pharmaceutical Biology*. **40**: 269–273.



- Eshun, K. & He, Q., 2004. Aloe Vera: A Valuable Ingredient for the Food, Pharmaceutical and Cosmetic Industries – A Review. *Critical Reviews in Food Science and Nutrition.* 44: 91–96.
- Farag, R.S., Daw, Z.Y., Hewedi, F.M. & El-Baroty, G.S.A., 1989. Antimicrobial activity of some Egyptian spice essential oils. *Journal of Food Protection*. 52: 665-667.
- Femenia, A., Sanchez, E.S., Simal, S. & Rossello, C., 1999. Compositional features of polysaccharides from Aloe vera (*Aloe barbadensis* Miller) plant tissues. *Carbohydrate Polymers.* **39**: 109-117.
- Fereira, M.A.S.S. & Lund, B.M., 1996. The effect of nisin on *Listeria monocytogenes* in culture medium and long-life cottage cheese. *Letter Applied Microbiology*. 22: 433–438.
- Ferreres, F., Tomas-Barberan, F.A., Soler, C., Garcia-Viguera, C., Ortiz, A. & Tomas-Lorente, F., 1994. A simple extractive technique for honey flavonoid HPLC analysis. *Apidologie*. 25: 21-30.
- Fessenden, R. J. & Fessenden, J.S., 1982. Organic chemistry. Boston: Willard Grant Press.
- Firouzi, R., Azadbakht, M. & Nabinedjad, A., 1998. Anti-listerial activity of essential oils of some plants. *Journal of Applied Animal Research.* 14: 75-80.
- Francis, F.J., 1993. Polyphenols as natural colorants. In *Polyphenolic Phenomena*. Scalbert, A. (ed). Paris: Institut National de la Rocherche Agronomique. 209–220.
- Frankel, E.N., Waterhouse, A.L. & Teissedre, P., 1995. Principal phenolic phytochemicals in selected California wines and their antioxidant activity in inhibiting oxidation of human low density lipoproteins. *Journal of Agricultural Food Chemistry*. 43: 890-894.
- Freiburghaus, F., Kaminsky, R., Nkunya, M.H.H. & Brun, R., 1996. Evaluation of African medicinal plants for their *in vitro* trypanocidal activity. *J. Ethnopharmacol.* 55: 1-11.
- Gage, D. 1996. Aloe Vera: Nature's Soothing Healer. United Kingdom: Healing Arts Press.
- Garcia-Delgado, R.A., Cotoruelo, L.M. & Rodriguez, J.J., 1992. Adsorption of anionic mixtures by polymeric resins. *Separation Science and Technology*. 27: 1065– 1076.
- Gazzani, G., Papetti, A., Massolini, G. & Daglia, M.,1998. Antioxidative and pro-oxidant activity of water soluble components of some common diet vegetables and the effect of thermal treatment. *Journal of Food Chemistry*. 6: 4118-4122.



- Geissman, T. A. 1963. Flavonoid compounds, tannins, lignins and related compounds. In Pyrrole pigments, isoprenoid compounds and phenolic plant constituents. Florkin, M. & Stotz, E.H. (ed). New York: Elsevier. 265.
- Gil, M.I., Ferreres, F. & Tomas-Barberan, F.A., 1998. Effect of modified atmosphere packaging on the flavonoids and vitamin C content of minimally processed swiss chard (*Beta vulgaris* subspecies *cycla*). *Journal of Agricultural & Food Chemistry*. 46: 2007-2012.
- Gill, A.O. & Holley, R.A., 2000. Inhibition of bacterial growth on ham and bologna by lysozyme, nisin and EDTA. Food Research International. 33: 83–90.
- Gill, A.O., Delaquis, P., Russo, P. & Holley, R.A., 2002. Evaluation of antilisterial action of cilantro oil on vacuum packed ham. *International Journal of Food Microbiology*. 73: 83-92.
- Govindarajan, V. S., & Sathyanarayana, M. N., 1991. Capsicum-production, technology, chemistry and quality. Part. V. Impact on physiology, pharmacology, nutrition and metabolism, structure, pungency, pain, and desensitization sequences. *Food Science and Nutrition.* 29: 435-471.
- Grimmer, H.R., Parbhoo, V. & Mc Grath, R.M., 1992. Antimutagenicity of polyphenol-rich fractions from Sorghum bicolor grain. J. Sci. Food Agric. 59: 251–256.
- Gustafson, J.E., Liew, Y.C., Chew, S., Markham, J.L., Bell, H.C., Wyllie, S.G. & Warmington, J.R., 1998. Effects of tea tree oil on *Escherichia coli*. Letters in Applied Microbiology. 26: 194–198.
- Gutterman, Y. & Chauser-Volfson, E., 2000. The distribution of the phenolic metabolites barbaloin, aloeresin and aloenin as a peripheral defense strategy in the succulent leaf parts. *Biochemical Systematics and Ecology.* 28: 825-838.
- Hammer, K.A., Carson, C.F. & Riley, T.V., 1999. Antimicrobial activity of essential oils and other plant extracts. *Journal of Applied Microbiology*. 86: 985–990.
- Haslam, E. 1996. Natural polyphenols (vegetable tannins) as drugs: possible modes of action. *Journal of Natural Product.* 59: 205–215.
- Hatano, T., Kusuda, M., Inada, K., Ogawa, T., Shiota, S., Tsuchiya, T. & Yoshida, T. 2005. Review: Effects of tannins and related polyphenols on methicillin-resistant *Staphylococcus aureus. Phyochemistry.* 66 (17): 2047-2055.
- He, Q. Liu C., Eshun K. & Zhang T. 2005. Quality and safety assurance in the processing of *Aloe vera* gel juice. *Food Control.* 16: 95–104.



Helander, I.M., Alakomi, H.-L., Latva-Kala, K., Mattila-Sandholm, T., Pol, I., Smid, E.J., Gorris, L.G.M. & Von Wright, A., 1998. Characterization of the action of selected essential oil components on Gram-negative bacteria. *Journal of Agricultural and Food Chemistry*. **46**: 3590–3595.

- Hemwimol, S., Pavasant, P. & Shotipruk, A., 2005. Ultrasound-assisted extraction of anthraquinones from roots of *Morinda citrifolia*. Ultrasonics Sonochemistry.
- Hennessee, O.M. & Cook, B.R., 1994. Aloe myth, magic and medicine. New York: Raven Press.
- Henry, R. 1979. An updated review of aloe vera. Cosmetics and Toiletries, 94:42-50.
- Hiramatsu, T., Imoto, M., Koyano, T. & Umezawa, K., 1993. Inductionof Normal Phenotypes in Rat-transformed Cells by Damnacanthal from *Morinda citrifolia*. *Cancer Lett.* **73**: 161-6.
- Hollman, P.C.H. & Arts, I.C.W., 2000. Review Flavonols, flavones and flavanols nature, occurrence and dietary burden. *Journal of the Science of Food and Agriculture*. 80: 1081-1093.
- Hollman, P.C.H. & Katan, M.B., 1997. Absorption, metabolism, and health effects of dietary flavonoids in man. *Biomed. Pharmacother.* 51:305–310.
- Hsu, C.-L., Chen, W., Weng, Y.-M., & Tseng, C.-Y. 2003. Chemical composition, physical properties, and antioxidant activities of yam flours as affected by different drying methods. *Food Chemistry*. 83: 85–92.
- Hu, Y., Xu, Q. & Hu, Q., 2003. Evaluation of Antioxidant Potential of Aloe vera (Aloe barbadensis Miller) Extracts. Journal of Agricultural and Food Chemistry. 51: 7788-7791.
- Hu, Q., Hu, Y. & Xu, J., 2005. Free radical-scavenging activity of Aloe vera (Aloe barbadensis Miller) extracts by supercritical carbon dioxide extraction. Food Chemistry. 91: 85–90.
- Huang, D, OU, B. & Prior, R.L. 2005. Review: the chemistry behind antioxidant capacity assays. *Journal of Agricultural and Food Chemistry*. 53: 1841-1856.
- Hyronimus, B., Le Marrec, C., Hadj Sassi, A. & Deschamps, A., 2000. Acid and bile tolerance of spore-forming lactic acid bacteria. *International Journal of Food Microbiology*. 61: 193–197.
- International Aloe Science Council (IASC). 2004. How large is the aloe market? Texas, USA. <u>http://www.iasc.org/aloemarket.html</u>. Updated on 19 October 2004.



International Aloe Science Council (IASC). 2006. Facts, procedures, an information for IASC's "Must Have" certification program. Texas, USA. http://www.iasc.org/certification_facts.html. Updated on 6 January 2006.

- International Aloe Science Council (IASC). 2006. Why buy products certified by the IASC? Texas, USA. <u>http://www.iasc.org/why_certified.html</u>. Updated on 25 April 2006.
- Joseph, J.A., Shukit-Hale, B. & Denisova, N.A., 1999. Reversal of age-related declines in neuronal signal transduction, cognitive, and motor behavioural deficits with blue berry, spinach, or strawberry dietary supplementation. *Journal of Neuroscience*, 19: 8114-8812.
- Juven, B.J., Kanner, J., Schved, F. & Weisslowicz, H., 1994. Factors that interact with the antibacterial action of thyme essential oil and its active constituents. *Journal* of Applied Bacteriology. **76**: 626–631.
- Karadeniz, F., Durst, R.W. & Wrolstad, R.E., 2000. Polyphenolic Composition of Raisins. Journal of Agricultural and Food Chemistry. 48: 5343-5350
- Karakaya, S., 2004. Bioavailability of phenolic compounds. Critical Reviews in Food Science and Nutrition. 44: 453–464.
- Kaur, C. & Kapoor, H. C., 2001. Antioxidants in fruits and vegetables-the millennium's health. Int. J. Food Sci. Technol. 36: 703-725.
- Kim, Y.M. Lee, C.H., Kim, H.G. & Lee, H.S., 2004. Anthraquinone isolated from Cassia tora (Leguminosae) seed show an antifungal property against phytopathogenic fungi. Journal of Agricultural and Food Chemistry. 52: 6096-6100.
- Kim, H.M. & Cho, S.H., 1999. Lavender inhibits immediate type allergic reaction in mice and rats. J. Pharmac. Pharmaco. 51: 221-226.
- Kim J.M., Marshall, M.R. & Wei, C.I. 1995a. Antibacterial activity of some essential oil components against five foodborne pathogens. *Journal of Agricultural & Food Chemistry*. 43: 2839-2845.
- Kim, J.M., Marshall, M.R., Cornell, J.A., Preston, J.F. & Wei, C.I., 1995b. Antibacterial activity of carvacrol, citral, and geraniol against *Salmonella typhimurium* on culture medium and on fish cubes. *Journal of Food Science*. 60: 1364–1374.
- Koutsoudaki, C., Krsek, M. & Rodger, A., 2005. Chemical composition and antibacterial activity of the essential oil and the gum of *Pistacia lentiscus var. chia. Journal of Agriculture and Food Chemistry*. 53: 7681-7685.
- Lambert, R.J.W. & Pearson, J., 2000. Susceptibility testing: accurate and reproducible minimum inhibitory concentration (MIC) and non-inhibitory concentration (NIC) values. *Journal of Applied Microbiology*. 88: 784–790.



- Lambert, R.J.W., Skandamis, P.N., Coote, P. & Nychas, G.-J.E., 2001. A study of the minimum inhibitory concentration and mode of action of oregano essential oil, thymol and carvacrol. *Journal of Applied Microbiology*. 91: 453–462.
- Lawless J. & Allan J., 2000. Aloe Vera-Natural Wonder Cure. London: Harper Collins Publishers.
- Lee, K.Y., Weintraub, S.T. & Yu, B. P., 2000. Isolation and Identification of a phenolic antioxidant from Aloe barbandensis. Free Radical Biology & Medicine. 28: 261-265.
- Lorian, V., 1995. Antibiotics in laboratory medicine. In *Disk susceptibility test (4th ed).* Acar, J.F. & Goldstein, F.W. (ed). London: Williams & Walkins Awaverly.
- Lu Yinrong & Foo L. Y., 2000. Antioxidant and radical scavenging activities of polyphenols from apple pomace. *Food Chemistry*. 68: 81-85.
- Madsen, H. L. & Bertelsen, G., 1995. Spices as antioxidants. Trends in Food Science and Technology. 6: 271–277.
- Mann, C.M. & Markham, J.L., 1998. A new method for determining the minimum inhibitory concentration of essential oils. *Journal of Applied Microbiology*, 84: 538–544.
- Marino, M., Bersani, C. & Comi, G., 2001. Impedance measurements to study the antimicrobial activity of essential oils from *Lamiacea* and *Compositae*. *International Journal of Food Microbiology*. 67: 187–195.
- Martianez-Romero, D., Alburquerque, N., Valverde, J.M., Guillean, F., Castillo, S., Valero, D. & Serrano, M. 2006. Postharvest sweet cherry quality and safety maintenance by *Aloe vera* treatment: A new edible coating. *Postharvest Biology and Technology*. **39**: 93-100.
- Mason, T. L. & Wasserman, B.P., 1987. Inactivation of red beet beta glucan synthase by native and oxidized phenolic compounds. *Phytochemistry*. 26: 2197–2202.
- Mazza, G. & Miniati, E. 1994. Anthocyanins in Fruits, Vegetables and Grains. Bota Raton, Florida: CRC Press.
- McDevitt, J. T., Schneider, D.M., Katiyar, S.K. & Edlind, T.D., 1996. Berberine: a candidate for the treatment of diarrhea in AIDS patients. *Program and Abstracts* of the 36th Interscience Conference on Antimicrobial Agents and Chemotherapy. Washington: American Society for Microbiology.
- McGimpsey, J.A. & Porter, M.G., 1999. A grower's guide for commercial production. New Zealand: New Zealand Institute for Crop & Food Research Ltd.



- McMahon, J. B., Currens, M.J., Gulakowski, R.J., Buckheit, R.W.J., Lackman-Smith, C., Hallock, Y.F. & Boyd, M.R., 1995. Michellamine B, a novel plant alkaloid, inhibits human immunodeficiency virus-induced cell killing by at least two distinct mechanisms. *Antimicrob. Agents Chemother*. 39:484–488.
- Miguel, G., Simoes, M., Figueiredo, A.C., Barroso, J.G., Pedro, L.G. & Carvalho, L., 2004. Composition and antioxidant activities of the essential oils of *Thymus caespititius*, *Thymus camphoratus* and *Thymus mastichina*. *Food Chemistry*. **86**: 183-188.
- Miller, H. E., 1971. A simplified method for the evaluation of antioxidants. Journal of the American Oil Chemists Society. 48: 91-97.
- Miller, N. J., Rice-Evans, C. A., Davies, M. J., Gopinathan, V., & Miller, A., 1993. A novel method of measuring antioxidant capacity and its application to monitoring the antioxidant status in premature neonates. *Clinical Science*. 84: 407–412.
- Moure, A., Franco, D., Sineiro, J., Dominguez, H., Nunez, M. J., & Lema, J. M., 2000. Evaluation of extracts from *Gevuina avellana* hulls as antioxidants. *Journal of Agricultural and Food Chemistry*. **48**: 3890–3897.
- Mourey, A. & Canillac, N., 2002. Anti-Listeria monocytogenes activity of essential oils components of conifers. Food Control. 13: 289-292.
- Naidu, A.S., 2000. Natural food antimicrobial systems. Bota Raton, Florida: CRC Press.
- National Committee for Clinical Laboratory Standards (NCCLS), 2000. Approved Standard M7-A5: Methods for Dilution Antimicrobial Susceptibility Test for Bacteria that Grow Aerobically, 5th edition. Wayne, PA: NCCLS.
- Nawar, W. W., 1996. Lipids. In Food chemistry. Fennema, O.R. (ed). New York: Marcel Dekker, Inc. 225-313.
- Nazrul Islam, S.K., Gray, A.I., Waterman, I. & Ahasan, M., 2002. Screening of eight alkaloids and ten flavonoids isolated from four species of the genus *Boronia* (*Rutaceae*) for antimicrobial activities against seventeen clinical microbial strains. *Phytotherapy Research*. **16**: 672–674.
- Negi, P.S., Jayaprakasha, G.K., Jagan Rao Mohan, L. & Sakariah, K.K., 1999. Antibacterial activity of turmeric oil: a byproduct from curcumin. *Journal of Agricultural and Food Chemistry*. 47: 4297-4300.
- Nepote, V., Grosso, N.R. & Guzman, C.A., 2005. Optimization of extraction of phenolic antioxidants from peanut skins. J Sci Food Agric. 85: 33–38.
- Ng, T.B., Liu, F., Lu, Y, Cheng, C.H.K. & Wang, Z., 2003. Antioxidant activity of compounds from the medicinal herb Aster tataricus. Comparative Biochemistry and Physiology Part C. 136: 109–115.



- Nikaido, H. & Vaara, M., 1985. Molecular basis of bacterial outer membrane permeability. Microbiological Reviews. 1, 1-2.
- Okamura, N., Asai, M., Hine, N., Akira Yagi, A., 1996. High-performance liquid chromatographic determination of phenolic compounds in *Aloe* species. *Journal* of Chromatography A. 746: 225-231.
- Okuda, T., Yoshida, T. & Hatano, T., 1992. Antioxidant effects of tannins and related phenols. In *Phenolic Compounds in foods and their effect on health*. Huang, M.T., Ho, C.T. & Lee, C.Y. (ed). Washington: American Chemical Society. 87-97.
- Omulokoli, E., Khan, B. & Chhabra, S.C., 1997. Antiplasmodial activity of four Kenyan medicinal plants. J. Ethnopharmacol. 56:133–137.
- Oomah, B.D. & Mazza, G. 2000. Functional Foods. In The Wiley Encyclopedia of Science & Technology, 2nd edition, Vol. 2. Francis, F.J. (ed). New York: Wiley. 1176-1182.
- Oosterhaven, K., Poolman, B. & Smid, E.J., 1995. S-carvone as a natural potato sprout inhibiting, fungistatic and bacteristatic compound. *Industrial Crops and Products*. 4: 23–31.
- Orafidiya, L.O., Agbania, E.O., Oyedelea, A.O., Babalolab, O.O., Onayemic, O. & Aiyedun, F.F. 2004. The effect of *aloe vera* gel on the anti-acne properties of the essential oil of *Ocimum gratissimum* Linn leaf – a preliminary clinical investigation. *The International Journal of Aromatherapy*. 14: 15–21.
- Ormancey, X., Sisalli, S., & Coutiere, P., 2001. Formulation of essential oils in functional perfumery. *Parfums, Cosmetiques, Actualites.* 157: 30–40.
- Packiyasothy, E.V. & Kyle, S., 2002. Antimicrobial properties of some herb essential oils. Food Australia. 54: 384-387.
- Pardue, K. 1997. Versatile Amberlite XAD-2 Resin Now Available. Supelco: The Reporter. 16: 1.
- Park, M. K., Park, J.H., Kim, N.Y., Shin, Y.G., Choi, Y.S., Lee, Y.G., Kim, K.H. & Lee, S.K., 1998. Analysis of 13 Phenolic Compounds in *Aloe* species by High Performance Liquid Chromatography. *Phytochemical Analysis.* 9: 186-191.
- Parr, A.J. & Bolwell, G.P., 2000. Phenols in the plant and in man. The potential for possible nutritional enhancement of the diet by modifying the phenols content or profile. *Journal of the Science of food and agriculture*. 80: 985-1012.



- Paster, N., Juven, B.J., Shaaya, E., Menasherov, M., Nitzan, R., Weisslowicz, H. & Ravid, U., 1990. Inhibitory effect of oregano and thyme essential oils on moulds and foodborne bacteria. *Letters in Applied Microbiology*. **11**: 33–37.
- Pastrana-Bonilla, E., Akoh, C.C., Sellappan, S. & Krewer, G., 2003. Phenolic content and antioxidant capacity of Muscadine grape. *Journal Agricultural and Food Chemistry*. 51: 5497-5503.
- Perucka, I., 1996. Effect of 2-chloroethylphpsphonic acid on pheny-lalanine ammonia lyase activity and formation of capsaicinoids in hot pepper fruits. Acta Physiologie Plantarum. 18: 7-12.
- Phillipson, J. D. & O'Neill, M.J., 1987. New leads to the treatment of protozoal infections based on natural product molecules. Acta Pharm. Nord. 1:131-144.
- Pintore, G., Usai, M., Bradesi, P., Juliano, C., Boatto, G., Tomi, F., Chessa, M., Cerri, R. & Casanova, J., 2002. Chemical composition and antimicrobial activity of *Rosmarinus officinalis* L. oils from Sardinia and Corsica. *Flavour and Fragrance Journal.* **17**: 15-19.
- Pol, I.E. & Smid, E.J., 1999. Combined action of nisin and carvacrol on Bacillus cereus and Listeria monocytogenes. Letters in Applied Microbiology. 29: 166-170.
- Prior, R.L. & Cao, G., 2000. Antioxidant phytochemicals in fruits and vegetables: diet and health implications. *Horticulture Science*. 35: 588-592.
- Prior, R. L., Hoang, H., Gu, L., Wu, X., Bacchioca, M., Howard, L., Hampsch-Woodill, M., Huang, D., Ou, B., Jacob, R., 2003. Assays for hydrophilic and lipophilic antioxidant capacity (oxygen radical absorbance capacity (ORAC) of plasma and other biological and food samples. *Journal of Agriculure & Food Chemistry*. 51: 3273-3279.
- Prudent, D., Perineau, F., Bessiere, J.M., Michel, G.M. & Baccou, J.C., 1995. Analysis of the essential oil of wild oregano from Martinique (*Coleus aromaticus Benth.*) evaluation of its bacterioatatic and fungistatic properties. *Journal of Essential Oil Research.* 7: 165-173.
- Pugh N., Ross S.A., ElSohly M.A., Pasco D.S., 2001. Characterization of Aloeride, a new high-molecular-weight polysaccharide from *Aloe vera* with potent immunostimulatory activity. *Journal of Agriculture and Food Chemistry.* 49: 1030–1034.
- Puupponen-Pimiä R., Nohynek, L., Meier, C., Kähkönen, M., Heinonen, M., Hopia A. & Oksman-Caldentey, K.M., 2001. Antimicrobial properties of phenolic compounds from berries. *Journal of Applied Microbiology*. 90: 494.



- Puupponen-Pimia, R., L. Nohynek, Hartmann-Schmidlin, S., Kahkonen, M., Heinonen, M., Maatta-Riihinen, K & Oksman-Caldentey, K.M., 2005. Berry phenolics selectively inhibit the growth of intestinal pathogens. *Journal of Applied Microbiology*. 98: 991–1000.
- Rauha, J-P., Remesa, S., Heinonen, M., Hopia, A., Kahkonen, M., Kujala, T., Pihlaja, K., Vuorelaa, H. & Vuorelaa, P., 2000. Antimicrobial effects of Finnish plant extracts containing flavonoids and other phenolic compounds. *International Journal of Food Microbiology*. 56: 3-12.
- Rice-Evans, C.A., Miller, N.J. & Paganga, G., 1996. Structure-antioxidant activity relationships of flavonoids and phenolic acids. *Free Radical Biology and. Medicine*. 20: 933–956.
- Rios, J.L., Recio, M.C. & Villar, A., 1988. Screening methods for natural products with antimicrobial activity: a review of the literature. *Journal of Ethnopharmacology*. 23: 127-149.
- Rival, S.G., Boeriu, C.G. & Wichers, H.J., 2001. Caseins and casein hydrolysates antioxidative properties and relevance to lipoxygenase inhibition. *Journal of Agricultural and Food Chemistry*. **49**: 295-302.
- Reynolds, T. & Dweck, A.C., 1999. Aloe vera leaf gel: a review update. Journal of Ethnopharmacology. 68: 3–37.
- Rhodes, M. J. C. & Price, K. R., 1996. Analytical problems in the study of flavonoid compounds in onions. *Food Chemistry*. 57: 113-117.
- Robards, K., Prenzler, P.D., Tucker, G., Swatsitang, P. & Glover, W. 1999. Phenolic compounds and their role in oxidative processes in fruits. *Food Chemistry*. 66: 401–436.
- Roginsky, V. & Lissi, E. A., 2005. Review of methods to determine chainbreaking antioxidant activity in food. *Food Chemistry*. 92: 235-254.
- Rowe, T.D. & Parks, L.M., 1941. Phytochemical study of Aloe vera leaf. Journal of the American Pharmaceutical Association, 30:262–266.
- Sacchetti, G., Maietti, S., Muzzoli, M., Scaglianti, M., Manfredini, S., Radice, M., Bruni, R., 2005. Comparative evaluation of 11 essential oils of different origin as functional antioxidants, antiradicals and antimicrobials in foods. *Food Chemistry*. **91**: 621– 632.
- Sang, S., Lapsley, K., Jeong, W.S., Lachance, P.A., Ho, C.T. & Rosen, R.T., 2002. Antioxidative phenolic compounds isolated from almond skins (*Prunus amygdalus* Batsch). J. Agric. Food Chem. 50: 2459-2463.



- Saria, A., Lembeck, F. & Skotsch, G., 1981. Determination of capsaicin in tissues and separation of capsaicin analogues by high-performance liquid chromatography. *Journal of Chromatography.* 201: 41-46.
- Sawamura, M., 2000. Aroma and functional properties of Japanese yuzu (*Citrus junos Tanaka*) essential oil. *Aroma Research*. **1**(1): 14–19.
- Scalbert, A. & Williamson, G. 2000. Dietary intake and bioavailability of polyphenols. Journal of Nutrition. 130:20735–2085S.
- Scherrer, R. & Gerhardt, P., 1971. Molecular sieving by the *Bacillum megaterium* cell wall and protoplast. *Journal of Bacteriology*. **107**: 718–735.
- Schiller, L.R., 2001. Review article: the therapy of constipation. Aliment Pharmacology Therapy. 15: 749-763.
- Schmidt, H., 1988. Phenol oxidase (E.I.14.18.1), a marker enzyme for defense cells. Progress in histochemistry and cytochemistry, 17. New York: Gustav Fischer.
- Senatore, F., Napolitano, F. & Ozcan, M., 2000. Composition and antibacterial activity of the essential oil from *Crithmum maritimum* L. (*Apiaceae*) growing wild in Turkey. *Flavour and Fragrance Journal.* **15**: 186–189.
- Serafini, M., Ghiselli, A. & Ferro-Luzzi, A., 1994. Red wine, tea and anti-oxidants. Lancet 344: 626.
- Serrano, M., Valverde, J.M., Guillean, F., Castillo, S., Martianez-Romero, D. & Valero, D. 2006. Use of Aloe vera gel coating preserves the functional properties of table grapes. *Journal of Agricultural & Food Chemistry*. 54: 3882-3886.
- Sethi, M. L., 1979. Inhibition of reverse transcriptase activity by benzophenanthridine alkaloids. J. Nat. Prod. 42:187–196.
- Shahidi, F. & Naczk, M. 1995. Food phenolics, sources, chemistry, effects, applications. Lancaster, PA: Technomic Publishing Co. Inc.
- Shahidi, F. & Naczk, M., 2004. Phenolics in Food and Nutraceutical. Boca Raton, Florida: CRC Press.
- Sherwin, E. R., 1990. Antioxidants. In *Food additives*. Branen, R. (ed). New York: Marcel Dekker. 139-193.
- Sikkema, J., De Bont, J.A.M. & Poolman, B., 1994. Interactions of cyclic hydrocarbons with biological membranes. *Journal of Biological Chemistry*. **269**: 8022–8028.
- Singh, B & Singh, S., 2003. Antimicrobial activity of terpenoids from *Trichodesma* amplexicaule Roth. *Phytotherapy Research.* 17: 814–816.



- Sivropoulou, A., Kokkini, S., Lanaras, T. & Arsenakis, M., 1995. Antimicrobial activity of mint essential oils. J. Agric. Food Chem. 43: 2384–2388.
- Skandamis, P., Koutsoumanis, K., Fasseas, K. & Nychas, G.-J.E., 2001b. Evaluation of the inhibitory effect of oregano essential oil on *Escherichia coli* O157:H7, in broth culture with or without EDTA, using viable counts, turbidity and impedance, *Italian Journal of Food Science & Technology*. **13**: 65–75.
- Skerget, M., Kotnik, P., Hadolin, M., Rizner, Hras, A. R., Simoni, M. & Knez, Z., 2005. Phenols, proanthocyanidins, flavones and flavonols in some plant materials and their antioxidant activities. *Food Chemistry*. 89: 191–198.
- Slinkard, K. & Singleton, V. L., 1977. Total Phenol Analysis: Automation and Comparison with Manual Methods. American Journal of Enology and Viticulture, 28: 49-55.
- Sokmen, A., Gulluce, M., Akpulat, H.A., Daferera, D., Tepe, B., Polissiou, M., Sokmen, M. & Sahin, F., 2004. The in vitro antimicrobial and antioxidant activities of the essential oils and methanol extracts of endemic *Thymus spathulifolius*. *Food Control.* **15**: 627-634.
- Stavric, B. 1994. Antimutagens and anticarcinogens in foods. *Food Chem. Toxicol.*, **32**: 79–90.
- Stern, J. L., A. E. Hagerman, P. D. Steinberg & P. K. Mason., 1996. Phlorotannin-protein interactions. J. Chem. Ecol. 22: 1887–1899.
- Stintzing, F. C.; Stintzing A. S.; Carle, R.; Frei, B.; Wrolstad, R. E., 2002. Color and antioxidant properties of cyanidin-based anthocyanin pigments. J. Agric. Food Chem. 50: 6172-6181.
- Sudhakar, T., Ravishankar, G. A. & Venkataraman, L. V., 1992. Separation of capsaicin from phenylpropanoid compounds by high-performance liquid chromatography to determine the biosynthetic status of cells and tissues of *Capsicum frutescens* Mill. in vivo and in vitro. *Journal of Agricultural and Food Chemistry*. **40**: 2461-2463.
- Tsao, R. & Deng, J. 2005. Review: Separation procedures for naturally occurring a ntioxidant phytochemicals. *Journal of Chromatography B.* 812: 85-99.
- Thomson, W. A. R., 1978. *Medicines from the Earth*. United Kingdom: McGraw-Hill Book Co., Maidenhead.
- Tsigarida, E., Skandamis, P. & Nychas, G.-J.E., 2000. Behaviour of *Listeria monocytogenes* and autochthonous flora on meat stored under aerobic, vacuum and modified atmosphere packaging conditions with or without the presence of oregano essential oil at 5°C. *Journal of Applied Microbiology*. 89: 901–909.



- Xu, Ning, Gruber, M., Westcott, N., Soroka, J., Parkin, I. & Hegedus, D., 2005. A Method for the solvent extraction of low-boiling-point plant volatiles. *Phytochemical Analysis.* 16: 239–245.
- Ultee, A., Kets, E.P.W. & Smid, E.J., 1999. Mechanisms of action of carvacrol on the food-borne pathogen *Bacillus cereus*. *Applied and Environmental Microbiology*. 65: 4606–4610.
- Ultee, A., Kets, E.P.W., Alberda, M., Hoekstra, F.A. & Smid, E.J., 2000a. Adaptation of the food-borne pathogen *Bacillus cereus* to carvacrol. *Archives of Microbiology*. 174: 233–238.
- Ultee, A., Bennink M.H.J. & Moezelaar, R., 2002. The phenolic hydroxyl group of carvacrol is essential for action against the food-borne pathogen *Bacillus cereus*. *Applied and Environmental Microbiology*. **68**: 1561–1568.
- Urch, D. 1999. Aloe Vera: Nature's Gift. England: Blackdown Publications.
- Yang, B., & Kallio, H. 2001. Fatty acid composition of lipids in sea buckthorn (*Hippophae rhamnoides* L.) berries of different origins. *Journal of Agricultural and Food Chemistry*. 49: 1939–1947.
- Valverde, J.M., Valero, D., Martianez-Romero, D., Guillean, F., Castillo, S. & Serrano, M. 2005. Novel edible coating based on Aloe vera gel to maintain table grape guality and safety. *Journal of Agricultural & Food Chemistry*. 53: 7807-7813.
- Van de Braak, S.A.A.J. & Leijten, G.C.J.J., 1999. Essential Oils and Oleoresins: A Survey in the Netherlands and other Major Market in the European Union. Rotterdam: CPI, Centre for the Promotion of Imports from Developing Countries. 116.
- van Overveld, F. W. P. C., Haenen, G. R. M. M., Rhemrev, J., Vermeiden, J. P. W. & Bast, A., 2000. Tyrosine as important contributor to the antioxidant capacity of seminal plasma. *Chemico Biological Interactions.* **127**: 151–161.
- Villasenor, I.M., Gajo, R.M.T. & Gonda, R.C., 1997. Bioactivity Studies on the alkaloid extracts from seeds of *Leucaena leucocephala*. *Phytotherapy Research*. 11: 615– 617.
- Vinha, A.F., Ferreres, F., Silva, B.M., Valentao, P., Goncalves, A., Pereira, J.A., Oliveira, M.B., Seabra, R.M. & Andrade, P.B., 2005. Phenolic profiles of Portuguese olive fruits (*Olea europaea* L.): Influences of cultivar and geographical origin. *Food chemistry*. 89: 561-568.
- Vishwakarma, R. A., 1990. Stereoselective synthesis of a-arteether from artemisinin. Journal of Natural Product. 53: 216–217.

Wang, Y-T., 1993. Bases of Aloe Certification. Aloe Today, Winter. 27–29.



- Wargovich, M.J., 2000. Anticancer properties of fruits and vegetables. *Horticulture Science*. 35: 573-575.
- Wamer W.G., Vath P. & Falvey, D.E., 2003. In vitro studies on the photobiological properties of aloe emodin and aloin A. *Free Radical Biological Medicine*. 34: 233-242.
- Wendakoon, C.N. & Sakaguchi, M., 1995. Inhibition of amino acid decarboxylase activity of *Enterobacter aerogenes* by active components in spices. *Journal of Food Protection.* 58: 280–283.
- Wilkins, K.M. & Board, R.G., 1989. Mechanisms of actions of food preservation procedures. In *Natural antimicrobial systems*. Gould, G.W. (ed). London: Elsevier. 285-362.
- Wilkinson, J.M., Hipwell, M., Ryan, T. & Cavanagh, H.M.A., 2003. Bioactivity of Backhousia citriodora: Antibacterial and antifungal activity. Journal of Agricultural and Food Chemistry. 51: 76– 81.
- Wu, X., Beecher, G. R., Holden, J. M., Haytowitz, D. B., Gebhardt, S. E., Prior, R. L., 2004. Lipophilic and hydrophilic antioxidant capacities of common foods in the United States. J. Agric. Food Chem. 52: 4026-4037.
- Xie, B., Shi, H., Chen, Q. & Ho, C.T., 1993. Antioxidant properties of fractions and polyphenol constituents from green, oolong, and black teas. *Proceedings of the National Science Council, ROC, Part B, Life Sciences.* 17: 77–84.
- Yamamoto, I., 1973. Aloe ulcin, a new principle of Cape Aloe and gastrointestinal function, especially experimental ulcer in rats. Journal of The Medical Society of Toho University. 20: 342–347.
- Yao, L., Datta, N., Tomas-Barberan, F.A., Ferreres, F., Martos, I. & Singausong, R., 2003. Flavonoids, phenolic acids and abscisic acid in Australian and New Zealand *Leptospermum* honeys. *Food Chemistry*. 81: 159-168.
- Yen, G. C. & Chung, D. Y., 1999. Antioxidant effects of extracts from *Cassia tora* L. prepared under different degrees of roasting on the oxidative damage to biomolecules. *J. Agric. Food Chem.* 47: 1326-1332.
- Yen, G.C. & Hsieh., C.L. 1998. Antioxidant Activity of Extracts from Du-zhong (*Eucommiaulmoides*) toward Various Lipid Peroxidation Models in Vitro. *Journal* of Agricultural and Food Chemistry. 46: 3952-3957.
- Yen, G.C., Duh, P.D. & Chuan, D.Y., 2000. Antioxidant activity of anthraquinones and anthrone. *Food Chemistry*. **70**: 437-441.



- Yen, G.C., Chen, H.W. & Duh, P.D., 1998. Extraction and identification of an antioxidative component from Jue Ming Zi (*Cassia tora* L.). *Journal of Agricultural Food Chemistry.* 46: 820-824.
- Yusuf, S., Agunu A. & Diana M., 2004. The effect of *Aloe vera A*. Berger (*Liliaceae*) on gastric acid secretion and acute gastric mucosal injury in rats. *Journal of Ethnopharmacology.* 93: 33–37.
- Zawahry, M.E., Hegazy, M.R. & Helal, M., 1973. Use of aloe in treating leg ulcers and dermatoses. *International Journal of Dermatology*. **12**: 68 –73.
- Zhu, X., Zhang, H. & Lo, R., 2004. Phenolic compounds from the leaf extract of artichoke (*Cynara scolymus* L.) and their antimicrobial activities. J. Agric. Food Chem. 52: 7272-7278.

