

**PHYTOCHEMICAL AND BIOLOGICAL STUDIES ON
SWEET POTATO (*IPOMOEA BATATAS*)**

LAI CHAN WEI

PERPUSTAKAAN
UNIVERSITI MALAYSIA SABAH

**DISSERTATION SUBMITTED IN PARTIAL
FULFILMENT OF THE REQUIREMENTS FOR THE
DEGREE OF BACHELOR OF SCIENCE WITH HONOURS**

**INDUSTRIAL CHEMISTRY PROGRAMME
SCHOOL OF SCIENCE AND TECHNOLOGY
UNIVERSITI MALAYSIA SABAH**

APRIL, 2007



UMS
UNIVERSITI MALAYSIA SABAH

BORANG PENGESAHAN STATUS TESIS@

JUDUL: Phytochemical & Biological Studies on Sweet Potato (Ipomoea Batatas)

Ijazah: Degree of Bachelor of Science with Honours (Industrial Chemistry)

SESI PENGAJIAN: 6

Saya LAI CHAN WEI
(HURUF BESAR)

mengaku membenarkan tesis (LPS/Sarjana/Doktor Falsafah)* ini disimpan di Perpustakaan Universiti Malaysia Sabah dengan syarat-syarat kegunaan seperti berikut:

1. Tesis adalah hakmilik Universiti Malaysia Sabah.
2. Perpustakaan Universiti Malaysia Sabah dibenarkan membuat salinan untuk tujuan pengajian sahaja.
3. Perpustakaan dibenarkan membuat salinan tesis ini sebagai bahan pertukaran antara institusi pengajian tinggi.
4. **Sila tandakan (/)

SULIT

(Mengandungi maklumat yang berdarjah keselamatan atau kepentingan Malaysia seperti yang termaktub di dalam AKTA RAHSIA RASMI 1972)

TERHAD

(Mengandungi maklumat TERHAD yang telah ditentukan oleh organisasi/badan di mana penyelidikan dijalankan)

TIDAK TERHAD

[Signature]

(TANDATANGAN PENULIS)

Disahkan oleh

[Signature]

(TANDATANGAN PUSTAKAWAN)

Alamat Tetap: 35B, Jln Abdullah

84000, Muar, Johor

Nama Penyelia

Tarikh: 14/4/2007

Tarikh: _____

CATATAN: * Potong yang tidak berkenaan.
 ** Jika tesis ini SULIT atau TERHAD, sila lampirkan surat daripada pihak berkuasa/organisasi berkenaan dengan menyatakan sekali sebab dan tempoh tesis ini perlu dikelaskan sebagai SULIT dan TERHAD.
 @ Tesis dimaksudkan sebagai tesis bagi Ijazah Doktor Falsafah dan Sarjana secara penyelidikan, atau disertasi bagi pengajian secara kerja kursus dan penyelidikan, atau Laporan Projek Sarjana Muda (LPSM).



DECLARATION

I hereby declare that this dissertation is based on my original work, except for quotations and summaries each of which have been fully acknowledged.



APRIL, 2007

LAI CHAN WEI

HS2004-1051



VERIFICATION

NAME: LAI CHAN WEI

TITLE: PHYTOCHEMICAL AND BIOLOGICAL STUDIES ON SWEET POTATO
(*IPOMOEA BATATAS*)

DR. HOW SIEW ENG

MR. MOH PAK YAN

DR. MD. LUTFOR RAHMAN

PROF MADYA DR. SHARIFF

A.K. OMANG

School of Science and Technology

APRIL, 2007



ACKNOWLEDGEMENTS

I would like to take this opportunity to thank all people who have assisted me in the completion of this project. I would like to express my sincere gratitude and appreciation to Dr. How Siew Eng, my advisor, for her constant support and guidance throughout the long course of the entire study.

I also offer thanks to En. Samudi and En. Sani for their technical help, all final year students for their support and encouragement. The support of all my best friends Khoo Yau Liang, Yong Wu Hock, Teo Pick Yien, Kua Shwu Fun, Wong How Cai, Ng Seong Wooi, Bong Wan Yuen, Tang Iyu Ying, Chee Lye Yung, Chong Chia Chin, Lim Kim Lee, Wong Siew Ching and fellow friends are appreciated.

Special thanks are given to my parents and sister for their love, support, patience and encouragement with out whose help my degree's study would not have been possible. Last but not least, I would like to show my gratitude to my lovely girlfriend, Leo Bey Fen, who gave me invaluable support and concern, with understanding helped bring this effort to fruition.



ABSTRACT

Sweet potato has received a broad attention because it is an important resource of food and there is an abundance of pharmacologically active ingredients in it. Three genotypes (storage roots and leaves) of sweet potato commercially available in Sabah (orange and purple fleshed) were studied for its phytochemical contents, antioxidation, antimicrobial and anti-kinase (MAPK) properties. Methanol crude extracts were obtained which were further partitioned into petroleum ether, chloroform and butanol extracts using solvent-solvent extractions. Phytochemical screenings demonstrated that the petroleum ether and chloroform extracts of all the genotypes contained saponins, tannins and anthraquinones whereas the butanol extract contained more classes of compounds (saponins, tannins, flavonoids and anthraquinones). The petroleum ether extract of storage roots (purple fleshed, Tambunan) showed a very potent antioxidation activity with relative antioxidation value of 1.025 (compared to fullness BHT, a synthetic antioxidant) as evaluated using the ferric thiocyanate (FTC) method. Petroleum ether extracts showed moderate to strong antioxidation activities. All the extracts demonstrated moderate antimicrobial activities against *S. aureus* (S 277) and *B. cereus* (B 43/04B) as evaluated a disc diffusion method. Relatively, the butanol extract was the most potent antimicrobial agent among all the extracts. However, no inhibition against *E. coli* (E 91/026) was observed. It was interesting to found out that all the butanol extracts of the storage roots showed very potent MAPK kinase inhibition as evaluated using a yeast screening system (yeast growing zone was 17 mm). However, inhibition of GSK-3 β was only detected in the petroleum ether extract of the leaves with 15 mm inhibition zone. Hence, sweet potato can be used as an easy accessible source of natural antioxidants, as a food supplement, or in the pharmaceutical and medical industries.



**KAJIAN FITOKIMIA DAN AKTIVITI BIOLOGI KE ATAS
KELEDEK (IPOMOEA BATATAS)**

ABSTRAK

Keledek semakin mendapat perhatian yang meluas kerana ia merupakan sumber makanan yang penting serta kaya dengan kandungan farmakologikal yang aktif. Sifat tiga jenis (umbi dan daun) keledek di Sabah (berisi jingga dan ungu) telah dikaji bagi kandungan fitokimia, anti-pengoksidaan, antimikrob dan anti-kinase (MAPK). Ekstrak petroleum ether, ekstrak butanol dan ekstrak kloroform telah dihasilkan melalui pengestrakan pelarut. Penyaringan fitokimia mendapati bahawa ekstrak petroleum-ether dan ekstrak kloroform mengandungi saponin, tanin dan antrakuinon. Manakala, kandungan bahan kimia yang didapati dalam ekstrak butanol pula adalah saponin, flavonoid, tanin dan antrakuinon. Bagi aktiviti anti-pengoksidaan (FTC), ekstrak petroleum-ether dari umbi keledek yang berisi ungu (Tambunan) telah memberi kesan anti-oksida yang terbaik di mana kerelatifan ekstrak petroleum-ether terhadap BHT adalah paling rendah dan hampir dengan nilai BHT, iaitu 1.025. Ekstrak petroleum ether menunjukkan ciri perencat serdehana terhadap aktiviti antimikrob. Ekstrak-ekstrak dari keledek menunjukkan bahawa ciri perencat serdehana terhadap S. aureus (S 277) dan B. cereus (B 43/04B). Secara bandingan, keputusan menunjukkan bahawa ekstrak butanol mempunyai ciri perencat antimikrob yang paling kuat daripada ekstrak lain. Tetapi, semua ekstrak tidak berpotensi merencat terhadap E. coli (E 91/026). Bagi sistem penyaringan perencat MAPK kinase, ketiga-tiga ekstrak butanol dari ubi keledek didapati perencat sebesar 17 mm. Manakala, sistem penyaringan GSK-3 β hanya didapati satu perencat pada ekstrak petroleum-ether dari daun keledek dimana diameternya adalah sebesar 15 mm. kesimpulannya, keledek boleh diguna sebagai bahan anti-oksida semulajadi, makanan dan di industri perubatan.



CONTENTS

	Page
DECLARATION	ii
VERIFICATION	iii
ACKNOWLEDGEMENTS	iv
ABSTRACT	v
ABSTRAK	vi
LIST OF CONTENTS	vii
LIST OF TABLES	xi
LIST OF FIGURES	xiii
LIST OF SYMBOLS	xv
CHAPTER 1 INTRODUCTION	
1.1 Background	1
1.2 Objectives of the study	5
1.3 Scope of study	5
CHAPTER 2 LITERATURE REVIEW	
2.1 Phytochemistry	6
2.2 Phytochemicals	7
2.3 The chemical compounds from plants	7
2.3.1 Phenols	7
2.3.2 Simple phenols	8
2.3.3 Phenols ethers	9
2.3.4 Phenylpropanoids	9
2.3.5 Flavonoids	10
2.3.6 Anthocyanins	12
2.3.7 Tannins	13
2.3.8 Quinones	14



2.4	Taxonomy classification of sweet potato [<i>Ipomoea batatas</i> (L.) Lam.]	15
2.4.1	Origin and ecology	15
2.4.2	Scientific classification	16
2.4.3	Morphology	16
2.4.4	Usage	18
2.5	Chemical constituents of sweet potato	19
2.5.1	Phenol	21
2.5.2	Flavonoid	22
2.5.3	Anthocyanin	24
	a. Anthocyanins contained in purple-fleshed sweet potato	24
	b. Physiological functionality	25
	c. Utilization	27
2.5.4	Pectin	28
2.6.	Anti-oxidation activities	28
2.6.1	Anti-oxidation properties	31
	a. Reducing power activity	31
	b. Scavenging activity	32
2.7	Anti-microbial properties	34
2.7.1	Bacteria as infectious agents	34
	a. <i>Bacillus subtilis</i>	37
	b. <i>Escherichia coli</i>	38
	c. <i>Staphylococcus aureus</i>	39
2.7.2	Fungi as infectious agent	40
	a. <i>Candida albicans</i>	41
2.8	Inhibit HIV activity	42
2.9	Anti-cancer activity	42
2.10	Vascular relaxing properties	43

CHAPTER 3 MATERIALS AND METHODS

3.1	Introduction	45
3.2	Chemicals and apparatus	45
3.3	Sample preparation	48



3.4	Extraction	48
3.5	Phytochemical screening	49
3.5.1	Screening of alkaloids	49
	a. Preparation of reagents	49
	b. Dragendorff test	50
	c. Wagner test	50
3.5.2	Screening of saponins	51
	a. Foam test	51
	b. Liebermann-Burchadd test	52
3.5.3	Screening of flavonoids	53
	a. Wilstatter-Sianidin test	53
	b. Bate-Smith test	54
	c. Metacalf test	54
3.5.4	Screening of tannins	54
	a. Gelatin test	55
	b. Ferric chloride test	55
3.5.5	Screening of anthraquinones	55
	a. Borntrager test	55
	b. Derivative anthraquinones test.	56
3.6	Biological activities	56
3.6.1	Antioxidation test	56
3.6.2	Antimicrobial test	57
	a. Preparation of agar media	58
	b. Nutrient agar (NA)	58
	c. Preparation of sample concentration stock	58
	d. Bacteria culture media	59
	e. Preparation of stock culture	59
	f. Preparation of lactophenol cotton blue	59
	g. Preparation of discs	60
	h. Interaction between bacteria and control solution	60
	j. Procedures	61
3.6.3	Screening of MAPK kinase inhibitor	63
	a. Preparation of sample solution	63



b.	Preparation of yeast cultivation media	63
c.	Preparation of MAPK kinase media	65
d.	Expected results for MAPK kinase inhibitor screening system	67
e.	GSK-3 β screening system	68
3.7	Summary of methodology	70
CHAPTER 4	RESULTS AND DISCUSSION	
4.1	Preparation of crude extracts	71
4.2	Phytochemical of screenings	72
4.2.1	Screening of alkaloids	72
4.2.2	Screening of saponins and sapogenins	73
4.2.3	Screening of flavonoids	75
4.2.4	Screening of tannins	76
4.2.5	Screening of anthraquinones	78
4.2.6	Summary of the phytochemical screening tests	79
4.3	Antioxidation test	81
4.4	Antimicrobial test	85
4.4.1	Screening of plant fraction extracts against bacteria	86
4.4.2	Discussion of antimicrobial test	89
4.5	Screening of MAPK kinase inhibitor	91
4.6	GSK-3 β screening system	93
CHAPTER 5	CONCLUSION	96
REFERENCE		98
APPENDIX		109



LIST OF TABLES

Table No.		Page
2.1	The taxonomy of <i>Ipomoea batatas</i> .	16
2.2	Phytochemicals and antioxidants components of tuberous root and leaf of sweet potato.	20
2.3	Free radical chain reaction mechanism.	29
2.4	Some comparison characteristics of Gram-positive and Gram-negative bacteria.	36
2.5	The scientific classification of <i>Bacillus subtilis</i> .	38
2.6	The scientific classification of <i>Escherichia coli</i> .	39
2.7	The scientific classification of <i>Staphylococcus aureus</i> .	40
3.1	The chemicals for extraction, phytochemical screening, anti-oxidation, anti-kinase and anti-microbial test.	46
3.2	The apparatus used.	47
3.3	Expected observation of appearance of alkaloids.	51
3.4	Duration time of bubble and quantitative determination.	52
3.5	Flavonoid type and its colour showed in Wilstatter-Sianidin Test.	53
3.6	Yeast cultivation media.	64
3.7	MAPK Kinase inhibitor assay medium yeast strain MKK1 ^{P386}	65
3.8	Preparation of GSK-3 β inhibitor assay medium.	69
4.1	Fraction of samples and yields.	72
4.2	Results of alkaloid screening on fraction extracts of leaves and storage roots of sweet potato.	73
4.3	Results of saponins and sapogenins screening on fraction extracts of leaves and storage roots of sweet potato.	74
4.4	Results of flavonoids screening on fraction extracts of leaves and storage roots of sweet potato.	76
4.5	Results of tannins screening on fraction extracts of leaves and storage roots of sweet potato.	77
4.6	Results of anthraquinones screening on various extracts of leaves and storage roots of sweet potato.	78



4.7	A summary of phytochemicals analysis of sweet potato extracts (PE, CH and BUT respectively).	80
4.8	Relative absorbance values of fraction extracts and negative control toward BHT value.	85
4.9	Screening of MAPK kinase inhibitor for fraction extracts.	92
4.10	Screening of GSK-3 β for fraction extracts.	94



LIST OF FIGURES

Figure No.		Page
2.1	Some phenolic plant natural products.	9
2.2	Phenol ethers.	9
2.3	Examples of plant phenylpropanoids.	10
2.4	Common classes of flavonoids.	11
2.5	Quinones.	14
2.6a	Leaves of sweet potato.	17
2.6b	Flower of sweet potato	17
2.6c	Orange tuberous roots of sweet potato.	17
2.6d	Purple tuberous roots of sweet potato.	17
2.7	Structure of acylated flavonoid glucosides.	23
2.8	Major anthocyanins contained in purple-fleshed sweet potato.	25
2.9	Inhibit HIV compounds.	42
3.1	The effect of extracts on mutant yeasts, MKK1 ^{P386} .	68
3.2	Steps carried out in this study.	70
4.1	Absorbance of fraction extracts of storage root (Orange flesh, Ranau) as measured by the FTC method.	83
4.2	Absorbance of fraction extracts of storage root (Purple flesh, Ranau) as measured by the FTC method.	83
4.3	Absorbance of fraction extracts of storage root (Purple flesh, Tambunan) as measured by the FTC method.	84
4.4	Absorbance of fraction extracts of leaves as measured by the FTC method.	84
4.5	Average diameter inhibition zones of 3 fraction extracts of storage roots with orange fleshed (Ranau).	87
4.6	Average diameter inhibition zones of 3 fraction extracts of storage roots with purple fleshed (Ranau).	88
4.7	Average diameter inhibition zones of 3 fraction extracts of storage roots with purple fleshed (Tambunan).	88
4.8	Average diameter inhibition zones of 3 fraction extracts of leaves.	89



LIST OF SYMBOLS AND ABBREVIATIONS

NA	Nutrient Agar
PDA	Potato Dextrose Agar
MAPK	Mitogen-activated protein kinase
GSK	Glycogen synthase kinase
DMSO	Dimethyl sulfoxide
<i>E. coli</i>	<i>Escherichia coli</i>
<i>S. aureus</i>	<i>Staphylococcus aureus</i>
<i>B. cereus</i>	<i>Bacillus subtilis</i>
CH	Chloroform
PE	Petroleum Ether
BUT	n-Butanol



CHAPTER 1

INTRODUCTION

1.1 Background

Since the beginning of civilization, human have succeeded in using plants in various ways, such as for building shelters, food and also as a source for medicine (Ahmad & Raji, 1992). Exploration of the plant kingdom has been made in search of chemical compounds of medicinal values and has been going on ever since, resulting in many empirical discoveries uncovering the main sources of botanical drugs. Remedies for various ailments normally involve the use of roots, leaves and bark of the plants (Ahmad & Raji, 1992).

In recent years, there has been a global trend toward the use of natural phytochemical, as antioxidants and functional ingredients, which are present in natural resources such as vegetable, fruits, oilseeds and herbs (Huang *et al.*, 2006). The use of traditional medicine is widespread, and plants still represent a large source of natural



antioxidants that might serve as leads for the development of novel drugs (Barton *et al.*, 1999). Several anti-inflammatory, digestive, anti-necrotic, neuroprotective, and hepatoprotective drugs have recently been shown to have an anti-oxidation and radical-scavenging mechanism as part of their activity (Huang *et al.*, 2004). Phytochemicals and antioxidant constituents in plant material have raised interest among scientists, food manufacturers, producers, and consumers for their roles in the maintenance of human health (Milner, 1999). Numerous epidemiological studies suggest that diets rich in phytochemicals and antioxidants execute a protective role in health and disease (Lako *et al.*, 2006).

Moreover, fruits and vegetables in the diet have been found in epidemiology studies to be protective against several chronic diseases. Frequent consumption of fruits and vegetables is associated with a lowered risk of cancer, cardiovascular disease, cataracts, heart disease, hypertension and stroke (Lako *et al.*, 2006). The risk of macular degeneration and stroke is diminished in people consuming large amounts of fruits and vegetables. Over 170 epidemiological cancer studies have been showed that there is a lower risk with increasing intake of fruit and vegetables. It is generally assumed that the vitamin and pro-vitamin antioxidants in these foods (ascorbic acid, tocopherols, and carotenoids) account for the beneficial effects (Vinson *et al.*, 1998).

However, belief in the medicinal power of foods is not a recent event but has been a widely accepted philosophy for generations (Milner, 1999). Moreover, researchers have found that some foods have functional properties because they provide physiological benefits which may enhance health and reduce the risk of developing chronic diseases. Functional foods are a very important part of wellness



and are defined as those which are whole, fortified, enriched or enhanced, providing health benefits beyond basic nutrition when consumed as part of regular, varied diet (Simmons, 2005). The term "functional food" is surfacing as a generic descriptor of the benefits that accompany ingesting foods that go beyond those accounted for merely by the nutritive provided. The Institute of Medicine of the National Academy of Sciences, United State has expanded this definition to include "any food or food ingredient that may provide a health benefit beyond the traditional nutrients it contains" (Milner, 1999).

Sweet potato has received a broad attention because it is an important resource of food and there is an abundance of pharmacologically active ingredients in it. So far, sweet potato has been widely used as a food staple, vegetable, and animal feed for industrial starch extraction and various processed products (Huang *et al.*, 2003; Srisuwan *et al.*, 2006). At the same time, more modern research show that sweet potato had higher levels of both carbohydrate and dietary fiber than potato and also had a stronger anti-oxidation activity than most other vegetables in a typical Western diet (Guan *et al.*, 2006).

A lot of works have been performed on the health-related function of sweet potato, and several important biological activities are attributed to sweet potato (Zhao *et al.*, 2005). Sweet potato (*Ipomoea batatas* L.), in which vitamin C, chlorogenic acid, caffeic acid, quercetin, and rutin are abundant, is one of the functional food products aimed at introducing human dietary ingredients that aid specific body functions in addition to being nutritious (Guan *et al.*, 2006). Extracts from sweet potato show strong radical scavenging and anti-mutagenic activities, significantly



reduced high blood pressure and carbon tetrachloride-induced liver injury, anti-inflammatory, antimicrobial, and antihypertensive activities, and ultraviolet protection effects (Aruoma, 1998; Hou *et al.*, 2001). Furthermore, sweet potato was recently identified to possess a postprandial anti-hyperglycemic (anti-diabetic) effect in rats through retardation of maltase activity (Guan *et al.*, 2006; Konczak-Islam *et al.*, 2003).

Sweet potato is nutritionally valuable, with higher levels of both carbohydrate and dietary fiber than potato (*Solanum tuberosum*), and a strong anti-oxidation activity that has been claimed to surpass most other vegetables in a typical Western diet. Hydroxycinnamic acids (HCA) are the main phenolic antioxidants in most commercially available sweet potato varieties, which can vary in storage root size, shape, flavor, texture, and colour, with the most common being white-, cream-, yellow-, or orange-fleshed. Several varieties of sweet potato have been developed with intense purple coloration, and conferred by high anthocyanin content in Japan and New Zealand (Philpott *et al.*, 2004).



1.2 Objectives of the study

1. To determine phytochemicals in sweet potato (*Ipomoea batatas*).
2. To screen the extracts for anti-kinase, anti-oxidation and anti-microbial activities.

1.3 Scope of study

Three varieties of sweet potato tuberous roots and leaves were selected in these studies which were purple fleshed sweet potato and orange fleshed sweet potato. These sweet potatoes were originated from Ranau and Tambunan, Sabah. Tuberous roots and leaves of those variety sweet potatoes were selected in this study because no report on the anti-oxidation, anti-microbial and anti-kinase activities of those varieties are presently available. Solid-solvent extraction and solvent-solvent extraction were carried out for bioactive compounds extraction from sweet potato. Phytochemical test and biological activity test for sweet potato (*Ipomoea batatas*) were carried out in this study. Screening of alkaloid, flavonoid, saponin, tannin and anthraquinon were included in the phytochemical study. Biological activities included anti-oxidation test, anti-microbial test and MAPK kinase and GSK-3 β test which showed medicinal use bioactive compounds in sweet potato.



CHAPTER 2

LITERATURE REVIEW

2.1 Phytochemistry

The subject of phytochemistry, or plant chemistry, has developed in recent year as a distinct discipline, somewhere in between natural product organic chemistry and plant biochemistry and is closely related to both. It is concerned with the enormous variety of organic substances that are elaborated and accumulated by plants and deals with the chemical structures of these substances, their biosynthesis, turnover and metabolism, their natural distribution and their biological function (Harbone, 1998).

In all these operations, methods are needed for separation, purification and identification of the many different constituents present in plants. Phytochemistry is directly related to the successful exploitation of known techniques, and the continuing development of new techniques to solve outstanding problems as they appear. One of the challenges of phytochemistry are to carry out all the above operations on vanishingly small amounts of material (Harbone, 1998).



2.2 Phytochemicals

Phytochemicals from fruits and vegetables have been shown to exert varied beneficial biological actions (Simmons, 2005). Phytochemicals are biologically-active, non-nutritive secondary metabolites which provide plants with colour, flavour and natural toxicity to pests (Johnson & Williamson, 2003). They are usually used to refer to compounds found in plants which are not required for normal functioning of the body but which nonetheless have a beneficial effect on health or an active role in the amelioration of disease (Harborne, 1973).

The classification of this huge range of compounds is fall into three main groups which are phenolic compound, glucosinolates, and carotenoids. Many thousands of phenolic compounds have been identified. They include monophenols, the hydroxycinnamic, acid group which contain caffeic and ferulic acid, flavonoids and glycosides, phytoestrogens and tannins (Johnson & Williamson, 2003).

2.3 Chemical compounds from plants

2.3.1 Phenols

The vast majority of the plant-based aromatic natural products are phenols. Numerous categories of these compounds are derived from phenol which includes simple phenols, phenylpropanoids, flavonoids, tannins and quinines (Kaufman *et al.*, 1999).



Phenolic compounds are usually susceptible to different factors (e.g., acidic solution and high temperature) during the extraction process. Drying at room temperature may enhance the enzymatic degradation and thus lower the amount of phenolics in the samples. Increasing the temperature above 60 °C lowering the phenolic amount considerably. At high temperatures, certain phenolics may decompose or combine with the other plant components (Miean & Mohamed, 2001).

In addition, other phenolic and polyphenolic compounds are present in plants such as cinnamic acid derivatives, for example, chlorogenic acid, and isomers of flavones known as isoflavones. Many of these phenols have been found to be more powerful anti-oxidation activity than vitamins C, E, and carotene using an *in vitro* model for heart disease, namely the oxidation of lower density lipoproteins (Vinson *et al.*, 1998).

2.3.2 Simple phenols

Most of the simple phenols are monomeric components of the polymeric polyphenols and acids which make up plant tissues, including lignin, melanin, flavolan and tannins. These individual components are obtained by acid hydrolysis of plant tissues. The components include *p*-hydroxybenzoic acid, protocatechuic acid, vanillic, syringic and gallic acid. Free phenols which do not require degradation of cell-wall polymers are relatively rare in plants. Hydroquinone, catechol, orcinol, and other simple phenols are found in relatively low concentrations. Some examples are shown in Figure 2.1.



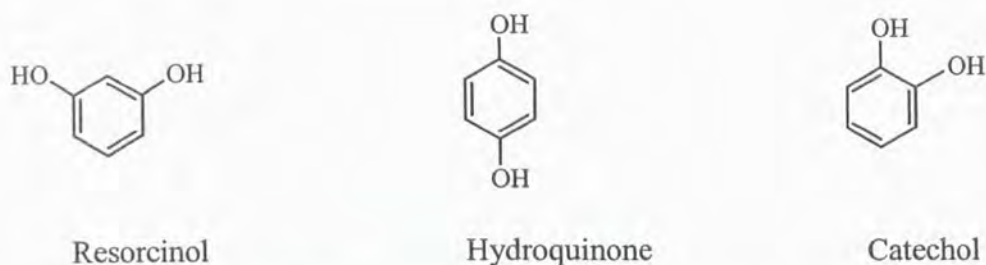


Figure 2.1 Some phenolic plant natural products (Kaufman *et al.*, 1999).

2.3.3 Phenols ethers

Many of the phenols also exist as their methyl ether; a few are shown in Figure 2.2. Khellin and visnagin are the active coumarin derivatives of the ammi visnaga fruit (*Ammi visnaga*). Trans Anethole is chiefly responsible for the taste and smell of anise seeds (*Pimpinella anisum*).

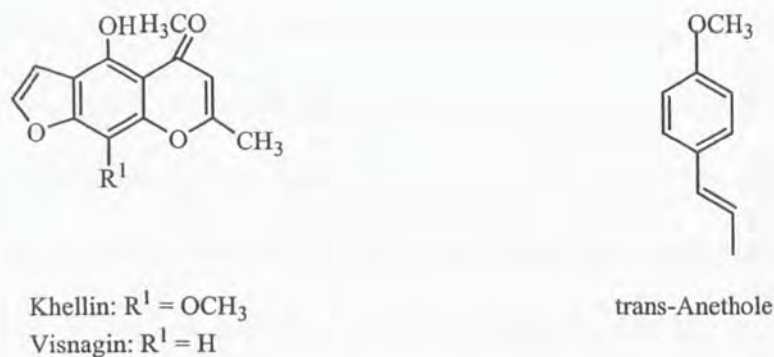


Figure 2.2 Phenol ethers (Kaufman *et al.*, 1999).

2.3.4 Phenylpropanoids

As the name implies, the phenylpropanoids contain a three-carbon side chain attached to a phenol. Common examples include the hydroxycoumarins, phenylpropenes and the lignans. Anethole and myristicin, the principles of nutmeg, are also representative

REFERENCES

- Afshari, A. T., Shirpoor, A., Farshid, A., Saadatian, R., Rasmi, Y., Saboory, E., Ilkhanizadeh, B. & Allameh, A. 2007. The effect of ginger on diabetic nephropathy, plasma antioxidant capacity and lipid peroxidation in rats. *Food Chemistry* **101**, pp. 148–153.
- Ahmad, F. B. & Raji, H. 1992. *Forest Biology and Conservation in Borneo*. Yayasan Sabah, Sabah, pp. 460-461.
- Ahmed, J. & Ramaswamy, H. S. 2006. Viscoelastic properties of sweet potato puree infant food. *Journal of Food Engineering* **74**, pp. 376–382.
- Antia, B. S., Akpan, E. J., Okon, P. A. & Umoren, I. U. 2006. Nutritive and Anti Nutritive Evaluation of Sweet Potatoes (*Ipomoea batatas*) Leaves. *Pakistan Journal of Nutrition* **5** (2), pp. 166-168.
- Aruoma, O. I. 1998. Free radicals, oxidative stress, and antioxidants in human health and disease. *Journal of the American Oil Chemists Society* **75**, pp. 199–212.
- Bauer, A. W., Kirby, W. M., Sherris, J. C. & Turck, M. 1966. *American Journal of Clinical Pathology* **45**, pp. 943–950.
- Barton, D. & Nakanishi, K. 1999. *Comprehensive natural products chemistry*. Pergamon, United State. pp. 3.
- Becker, B. F., Heindl, B., Kupatt, C. and Zahler, S. 2000. Endothelial function and hemostasis. *Zeitschrift für Kardiologie* **89**, pp. 160–167.



- Bonnefont-Rousselot, D. 2000. Consequences of diabetic status on the oxidant or antioxidant balance. *Diabetes Metabolism* **26**, pp. 163–173.
- Brown, A. A. & Hu, F. B. 2001. Dietary modulation of endothelial function: implications for cardiovascular disease. *American Journal of Clinical Nutrition* **73**, pp. 673–686.
- Burns, J., Gardner, P. T., Matthews, D., Duthie, G. G., Lean, M. E. & Crozier, A. 2001. Extraction of phenolics and changes in antioxidant activity of red wines during vinification. *Journal of Agricultural and Food Chemistry* **49**, pp. 5797–5808.
- Cambie, R. C. & Ferguson, L. R. 2003. Potential functional foods in the traditional Maori diet. *Mutation Research* **523**, pp. 109–117.
- Collins, I. & Workman, P. 2006. Design and Development of Signal Transduction Inhibitors for Cancer Treatment: Experience and Challenges with Kinase Targets. *Current Signal Transduction Therapy* **1**, pp. 13-23.
- Cowan, M. M., 1999. Plant products as antimicrobial agents. *Clinical Microbiology Reviews*. **12** (4), pp. 564-582.
- Dewanto, V., Wu, X., Adom, K. K. & Liu, R. H. 2002a. Thermal processing enhances the nutritional value of tomatoes by increasing total antioxidant activity. *Journal of Agricultural and Food Chemistry* **50**, pp. 3010–3014.
- Dewanto, V., Wu, X. & Liu, R. H. 2002b. Processed sweet corn has higher antioxidant activity. *Journal of Agricultural and Food Chemistry* **50**, pp. 4959–4964.
- Duke, J. A. 1985. *Handbook of medicinal herbs*. CRC Press, Inc., Boca Raton.



- Fasihuddin, A. & Hasmah, R. 1993. *Kimia hasilan semula jadi dan tumbuhan ubatan*. Dewan Bahasa dan Pustaka, Kuala Lumpur.
- Furuta, S. 1998. High tert-butylperoxyl radical scavenging activities of sweet potato cultivars with purple flesh. *Food Science Technology International* **4**, pp. 33–35.
- Guan, Y. Q., Wu, T., Lin, M. & Ye, J. N. 2006. Determination of Pharmacologically Active Ingredients in Sweet Potato (*Ipomoea batatas* L.) by Capillary Electrophoresis with Electrochemical Detection. *Journal of Agricultural and Food Chemistry* **54**, pp. 24-28.
- Halliwell, B. 1999. Food-derived antioxidants. Evaluation of their importance in food and in vivo. *Food Science and Agricultural Chemistry* **1**, pp. 67–109.
- Harborne, J. B. 1973. *Phytochemical Methods: A Guide To Modern Techniques of Plant Analysis*. Chapman and Hall, New York, pp. 2.
- Harbone, J. B. & Harbone, A. J. 1998. *Phytochemical methods*. Ed. ke-3. Thomson Science, London.
- Hardman, W. E. & Cameron, I. L. 1995. Site specific reduction of colon cancer incidence, without a concomitant reduction in cryptal cell proliferation, in 1,2-dimethylhydrazine treated rats by diets containing 10% pectin with 5% or 20% corn oil. *Carcinogenesis* **16**, pp. 1425–1431.
- Haug, J. C. & Sun, M. 2000. Genetic diversity and relationship of sweet potato and its wild relatives in *Ipomoea* series Batatas (Convolvulaceae) as revealed by inter-simple sequence repeat (ISSR) and restriction analysis of chloroplast DNA. *Theoretical and Applied Genetics* **100**, pp. 1050–1060.

- Hernandez, L., Munoz, R. A., Miro, G., Martinez, M., Silvia-Parra, J. & Chavez, P. 1984. Use of medicinal plants by ambulatory patients in Puerto Rico. *American Journal of Health-System Pharmacy* **41**, pp. 2060–2064.
- Hou, W. C., Chen, Y. C., Chen, H. J., Lin, Y. H., Yang, L. L. & Lee, M. H. 2001. Antioxidant Activities of Trypsin Inhibitor, a 33 KDa Root Storage Protein of Sweet Potato (*Ipomoea batatas* (L.) Lam cv. Tainong 57). *Journal of Agricultural and Food Chemistry* **49**, pp. 2978-2981.
- Hou, W. C., Han, C. H., Chen, H. J., Wen, C. L. & Lin, Y. H. 2005. Storage proteins of two cultivars of sweet potato (*Ipomoea batatas* L.) and their protease hydrolysates exhibited antioxidant activity in vitro. *Plant Science* **168**, pp. 449–456.
- Huang, D. J., Lin, C. D., Chen, H. J. & Lin, Y. H. 2004a. Antioxidant and antiproliferative activities of sweet potato (*Ipomoea batatas* [L.] Lam ‘Tainong 57’) constituents. *Botanical Bulletin of Academic Sinica* **45**, pp. 179-186.
- Huang, D. J., Chen, H. J., Hou, W. C., Lin, C. D. & Lin, Y. H. 2004b. Active Recombinant Thioredoxin h Protein with Antioxidant Activities from Sweet Potato (*Ipomoea batatas* [L.] Lam Tainong 57) Storage Roots. *Journal of Agricultural and Food Chemistry* **52**, pp. 4720-4724.
- Huang, D. J., Chen, H. J., Hou, W. C., Chen, T. E., Hsu, W. Y. & Lin, Y. H., 2005. Expression and function of a cysteine proteinase cDNA from sweet potato (*Ipomoea batatas* [L.] Lam ‘Tainong 57’) storage roots. *Plant Science* **169**, pp. 423–431.
- Huang, D. J., Chen, H. J., Hou, W. C., Lin, C. D. & Lin, Y. H. 2006. Sweet potato (*Ipomoea batatas* [L.] Lam ‘Tainong 57’) storage root mucilage with antioxidant activities in vitro. *Food Chemistry* **98**, pp. 774–781.



- Huang, W. C., Wang, A. W., Wang, L. T. & Sung, H. Y. 2003. Expression and Characterization of Sweet Potato Invertase in *Pichia pastoris*. *Journal of Agricultural and Food Chemistry* **51**, pp. 1494-1499.
- Huang, Y. C., Chang, Y. H. & Shao, Y. Y. 2006. Effects of genotype and treatment on the antioxidant activity of sweet potato in Taiwan. *Food Chemistry* **98**, pp. 529-538.
- Ishida, H., Suzuno, H., Sugiyama, N., Innami, S. & Maekawa, T. A., 2000. Nutritive evaluation on chemical components of leaves, stalks and stems of sweet potatoes (*Ipomoea batatas* poir). *Food Chemistry* **68**, pp. 359-367.
- Johnson, I. T. & Williamson, G. 2003. *Phytochemical Functional Foods*. Woodhead Publishing. England
- Kaufman, P. B., Cseke, L. J., Warber, S., Duke, J. A. & Brielmann, H. L. 1999. *Natural Products from Plants*. CRC Press. United State, pp. 20.
- Kazmi, M. H., Malik, A., Hameed, S., Akhtar, N. & Ali, S. N., 1994. An anthraquinone derivative from *Cassia italica*. *Phytochemistry*. **36**, pp.761-763.
- Keaney, J. F. & Loscalzo, J. 1999. Diabetes, oxidative stress and platelet activation. *Circulation* **99**, pp.189-191.
- Konczak-Islam, I., Yoshimoto, M., Hou, D. X., Terahara, N. & Yamakawa, O. 2003. Potential Chemopreventive Properties of Anthocyanin-Rich Aqueous Extracts from In Vitro Produced Tissue of Sweetpotato (*Ipomoea batatas* L.). *Journal of Agricultural and Food Chemistry* **51**, pp.5916-5922.



- Konczak-Islam, I., Terahara, N., Yoshimoto, M., Nakatanid, M., Yoshinagac, M. & Yamakawac, O. 2005. Regulating the composition of anthocyanins and phenolic acids in a sweet potato cell culture towards production of polyphenolic complex with enhanced physiological activity. *Trends in Food Science and Technology* **16**, pp.377–388.
- Kubo, L., Muroi, H. & Himejima, M. 1992. Antimicrobial activity of green teas flavor components and their combination effects. *Journal of Agricultural and Food Chemistry* **40**, pp.245-248.
- Lako, J., Trenerry, V. C., Wahlqvist, M., Wattanapenpaiboon, N., Sotheeswaran, S. & Premeir, R. 2006. Phytochemical flavonols, carotenoids and the antioxidant properties of a wide selection of Fijian fruit, vegetables and other readily available foods. *Food Chemistry*
- Lin, K. H., Tsou, C. C., Hwang, S. Y., Chen, L. F. & Lo, H. F. 2006. Paclobutrazol pre-treatment enhanced flooding tolerance of sweet potato. *Journal of Plant Physiology* **163**, pp.750-760.
- Liu, D. Z., Lu, Y. L., Cheng, H. C. & Hou, W. C., 2005. Immobilized Zinc Affinity Chromatography of Pectin Hydroxamic Acids for Purification of Trypsin Inhibitors from Soybean and Sweet Potato. *Journal of Agricultural and Food Chemistry* **53**, pp.10219-10223.
- Liu, Y., Ahmad, H., Luo, Y., Gardiner, D. T., Gunasekera, R. S. & McKeehan, W. L. 2001. Citrus pectin: characterization and inhibitory effects on fibroblast growth factor–receptor interaction. *Journal of Agricultural and Food Chemistry* **49**, pp.3051–3057.
- Madigan M. & Martinko J. 2005. *Brock Biology of Microorganisms*, Ed. Ke-11. Prentice Hall.



- Maeshima, M., Sasaki, T. & Asahi, T. 1985. Characterization of major proteins in sweet potato tuberous roots. *Phytochemistry* **24** (9), pp.1899-1902.
- Matsufuji, H. & Shibamoto, T. 2004. Inhibition of Malonaldehyde Formation in Oxidized Calf Thymus DNA with Synthetic and Natural Antioxidants. *Journal of Agricultural Food Chemistry* **52**, pp.5759-5763.
- Matsui, T., Ueda, T., Oki, T., Sugita, K., Terahara, N. & Kiyoshi Matsumoto, K. 2001. α -Glucosidase Inhibitory Action of Natural Acylated Anthocyanins. 1. Survey of Natural Pigments with Potent Inhibitory Activity. *Journal of Agricultural Food Chemistry* **49**, pp.1948-1951.
- Meijer, L. & Collin, O. 1999. KID, A Kinase Inhibitor Database Project. *Pharmacol therapeutic* **82** (2-3), pp. 165-168.
- Miean, K. H. & Mohamed, S. 2001. Flavonoid (Myricetin, Quercetin, Kaempferol, Luteolin, and Apigenin) Content of Edible Tropical Plants. *Journal of Agricultural Food Chemistry* **49**, pp.3106-3112.
- Milner, J. A. 1999. Functional foods and health promotion. *Journal of Nutrition* **129**, pp. 1395–1397.
- Mukherjee P. K. & Giri, S. N. 1995. Antifungal screening of *Nelumbo nucifera* (*Nymphaeaceae*) rhizome extract, *Indian Journal of Microbiology* **35**, pp. 320-327.
- Murray, R. J., Limb, T. T., Pearson, J. C., Grubb, W. B. & Lum, G. D. 2004. Community-onset methicillin-resistant *Staphylococcus aureus* bacteremia in Northern Australia. *International Journal of Infectious Diseases* **8**, pp. 275—283.



- Nakajima, N., Ishihara, K., Hamada, H., Kawabe, S. I. & Furuya, T. 2000. Regioselective Acylation of Flavonoid Glucoside with Aromatic Acid by an Enzymatic Reaction System from Cultured Cells of *Ipomoea batatas*. *Journal of Bioscience and Bioengineering* **90** (3), pp. 347-349.
- Nangia-Makker, P., Hogan, V., Honjo, Y., Baccarini, S., Tait, L. & Bresalier, R. 2002. Inhibition of human cancer cell growth and metastasis in nude mice by oral intake of modified citrus pectin. *Journal of the National Cancer Institute* **94**, pp. 1854–1862.
- Oki, T., Masuda, M., Furuta, S., Nishiba, Y., Terahara, N. & Suda, I. 2002. Involvement of anthocyanins and other phenolic compounds in radical-scavenging activity of purple-fleshed sweet potato cultivars. *Journal of Food Science* **67**, pp. 1752–1756.
- Osawa, T. & Namiki, M. 1981. A novel type of antioxidant isolated from leaf wax of Eucalyptus leaves. *Agricultural and Biological Chemistry* **45**, pp. 735-739.
- Philpott, M., Gould, K. S., Lim, C. & Ferguson, L. R. 2004. In Situ and In Vitro Antioxidant Activity of Sweet potato Anthocyanins. *Journal of Agricultural Food Chemistry* **52**, pp. 1511-1513.
- Platt, D. & Raz, A. 1992. Modulation of the lung colonization of B16-F1 melanoma cells by citrus pectin. *Journal of the National Cancer Institute* **84**, pp. 438–442.
- Rabah, I. O., Hou, D. X., Komine, S. I. & Fujii, M. 2004. Potential Chemopreventive Properties of Extract from Baked Sweet Potato (*Ipomoea batatas* Lam. Cv. Koganesengan) *Journal of Agricultural Food Chemistry* **52**, pp. 7152-7157.
- Ravi, V., Aked, J. & Balagopalan, C. 1996. Review on tropical root and tuber crops. I. Storage methods and quality changes. *Critical Review in Food Science and Nutrition* **36**, pp. 661–709.



- Robbers, J., Speedie, M. and Tyler, V. 1996. *Pharmacognosy and pharmacobiotechnology*. Trends in Wilkins, Baltimore, pp. 1-14
- Runnie, I., Salleh, M. N., Mohamed, S., Head, R. J. & Abeywardena, M. Y. 2004. Vasorelaxation induced by common edible tropical plant extracts in isolated rat aorta and mesenteric vascular bed. *Journal of Ethnopharmacology* **92**, pp. 311-316.
- Ryan, K. J. & Ray, C. G. 2004. *Sherris Medical Microbiology*, Ed. Ke-4. McGraw Hill.
- Sakakibara, H., Honda, Y., Nakagawa, S., Ashida, H. & Kanazawa, K. 2003. Simultaneous Determination of All Polyphenols in Vegetables, Fruits, and Teas. *Japan Agricultural Research Quarterly* **51**, pp. 571-581.
- Scalbert, A. 1991. Antimicrobial properties of tannins. *Phytochemistry* **30**, pp. 3875-3883.
- Simmons, P. C. 2005. *Harvest of the Month-Sweet potato. Family and Consumer Sciences*. University of Kentucky, United State, pp. 2-3.
- Srisuwan, S., Sihachakr, D. & Siljak-Yakovlev, S. 2006. The origin and evolution of sweet potato (*Ipomoea batatas* Lam.) and its wild relatives through the cytogenetic approaches. *Plant Science* **171**, pp. 424-433.
- Suda, I., Furata, S., Nishiba, Y., Yamakawa, O., Matsugano, K. & Sugita, K. 1997. Reduction of liver injury induced by carbon tetrachloride in rats administered purple-colored sweet potato juice. *Nippon Shokuhin Kagaku Kogaku Kaishi* **44**, pp. 315-318.



- Suda, I., Oki, T., Masuda, M., Kobayashi, M., Nishiba, Y. & Furuta, S. 2003. Physiological functionality of purple-fleshed sweet potatoes containing Anthocyanins and their utilization in food. *Japan Agricultural Research Quarterly* **37**(3), pp. 167-173.
- Tawfeek, H. I., Najim, N. H. & Al-Mashikhi, S. 2003. Efficacy of an infant formula containing anti-*Escherichia coli* colostral antibodies from hyperimmunized cows in preventing diarrhea in infants and children: a field trial. *International Journal of Infectious Diseases* **7** (2), pp. 120-128.
- Tian, Q. G., Konczak, I. & Schwartz, S. J. 2005. Probing Anthocyanin Profiles in Purple Sweet Potato Cell Line (*Ipomoea batatas* L. Cv. Ayamurasaki) by High-Performance Liquid Chromatography and Electrospray Ionization Tandem Mass Spectrometry. *Journal of Agricultural Food Chemistry* **53**, pp. 6503-6509.
- Tortora, G. J., Funke, B. R. & Case, C. L. 2004. *Microbiology*. Pearson Education Inc.
- Vinson, J. A., Hao, Y., Su, X. H. & Zubik, L. 1998. Phenol Antioxidant Quantity and Quality in Foods: Vegetables. *Journal of Agricultural Food Chemistry* **46**, pp. 3630-3634.
- Wang, B. H., Ternai, B. & Polya, G. 1997. Specific Inhibition of Cyclic Amp-Dependent Protein Kinases by Warangalone and Robustic. *Phytochemistry* **44** (5), pp. 787-796.
- Yoshimoto, M., Okuno, S., Yoshinaga, M., Yamakawa, O., Yamaguchi, M. & Yamada, J., 1999. Antimutagenicity of sweet potato (*Ipomoea batatas*) roots. *Bioscience, Biotechnology, Biochemistry* **63**, pp. 537-541.



Zhao, G. H., Kan, J. Q., Li Z. X. & Chen, Z. D. 2005. Characterization and immunostimulatory activity of an (1 \rightarrow 6)- α -D-glucan from the root of *Ipomoea batatas*. *International Immunopharmacology* 5, pp. 1436– 1445.

