

**DETECTION OF HOMOZYGOUS DELETION
OF SURVIVAL MOTOR NEURON 1 (SMN1)
GENE USING ALLELE-SPECIFIC PRIMER
IN SMA PATIENTS**

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**BIOTECHNOLOGY PROGRAMME
SCHOOL OF SCIENCE AND TECHNOLOGY
UNIVERSITI MALAYSIA SABAH**

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NEURON 1 (SMN1) GENE USING ALLELE-SPECIFIC PRIMER
IN SMA PATIENTS

HOO KIT MEI

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REQUIRMENT TO OBTAIN A BACHELOR DEGREE OF SCIENCE WITH
HONOURS

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APRIL 2007



DECLARATION

I hereby declare that the work in this project is of my own except quotations and summaries from the references which have been acknowledged.

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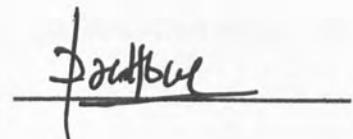
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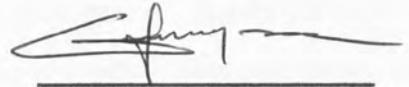
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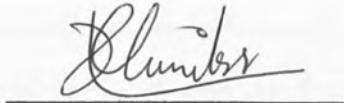
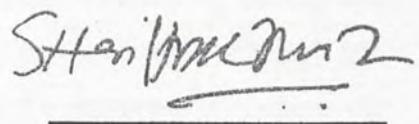
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ABSTRACT

This research was carried out to amplify the exon 7 of SMN1 gene by using allele-specific primer and optimize the PCR reagents and parameters for the detection of homozygous deletion of SMN1 gene in SMA patients. It had been noticed that 58.4 °C of annealing temperature gave the optimum condition in PCR amplification for both tel-SMN and β-globin primers. Based on the resulted PCR amplification profiles, both SMN1 and β-globin genes were performed in the normal healthy sample. Meanwhile, the identification of SMN1 gene deletion in SMA patients was verified by the absence of targeted band (SMN1 gene) together with the presence of β-globin gene band. This phenomenon could only be detected in the experiment with higher DNA concentration loaded, that was 68 ng. With the β-globin gene served as an internal control, the result was considered valid for any sample that showed β-globin gene band. In order to confirm the actual size of the PCR products, DNA sequencing was performed for both SMN1 and β-globin genes. However, the resulted sequencing electropherograms were failed to provide the complete sequence for all samples. Hence, PCR-allele specific method is specifically useful in the analysis of homozygous deletion of SMN1 gene in SMA disease but more optimizations are needed for DNA sequencing in order to make this study more reliable and success.



ABSTRAK

Penyelidikan ini telah dilakukan untuk mengamplifikasi ekson 7 dalam gen SMN1 dengan menggunakan primer alel-khusus dan mengoptimumkan reagen-reagen dan parameter-parameter bagi pengesanan pemotongan homozigus gen SMN1 dalam pesakit-pesakit SMA. Didapati bahawa suhu perlekatan primer pada 58.4 °C memberikan keadaan yang optimum dalam amplifikasi PCR bagi kedua-dua primer tel-SMN dan β-globin. Berdasarkan profil-profil amplifikasi PCR yang terhasil, kedua-dua gen SMN1 dan β-globin hadir dalam sampel yang normal dan sihat. Sementara itu, pengecaman pemotongan gen SMN1 di pesakit-pesakit SMA pula dapat disahkan dengan ketiadaan jalur sasaran (gen SMN1) bersama dengan kehadiran jalur gen β-globin. Fenomena ini hanya dapat dikesan dalam eksperimen yang dimuatkan dengan kepekatan DNA yang lebih tinggi, iaitu 68 ng. Dengan adanya gen β-globin sebagai kawalan dalaman, keputusan boleh dikatakan sahih bagi sebarang sampel yang menunjukkan jalur gen β-globin. Untuk memastikan saiz sebenar produk PCR, penujuukan DNA telah dijalankan bagi kedua-dua gen SMN1 dan β-globin. Walau bagaimanapun, elektroferogram-elektroferogram penujuukan yang terhasil adalah gagal untuk membekal jujukan yang lengkap bagi semua sampel. Maka, kaedah PCR alel-khusus ini adalah sangat berguna khususnya dalam analisis mengenai pemotongan homozigus gen SMN1 dalam penyakit SMA tetapi lebih optimasi adalah diperlukan untuk penujuukan DNA supaya kajian ini menjadi lebih menyakinkan dan berjaya.



LIST OF CONTENTS

	Page
DECLARATION	ii
VERIFICATION	iii
ACKNOWLEDGEMENT	iv
ABSTRACT	v
ABSTRAK	vi
LIST OF CONTENTS	vii
LIST OF TABLES	xii
LIST OF FIGURES	xiii
LIST OF SYMBOLS	xvi
LIST OF UNITS	xvii
LIST OF ABBREVIATIONS	xviii
CHAPTER 1 INTRODUCTION	1
CHAPTER 2 LITERATURE REVIEW	4
2.1 Spinal Muscular Atrophy (SMA) Disease	4
2.1.1 Types of SMA	4
a. Type I (Werdnig-Hoffmann Disease)	4
b. Type II (Dubowitz Disease)	5
c. Type III (Kugelberg-Welander Disease)	6
d. Type IV	7
2.1.2 Population Genetics	8



2.1.3	Gene Structure and Function	9
a.	Survival Motor Neuron 1 Gene (SMN1)	10
b.	Survival Motor Neuron 2 Gene (SMN2)	11
2.1.4	Molecular Diagnosis	11
2.1.5	Pattern of Inheritance	13
2.2	Polymerase Chain Reaction (PCR)	16
2.2.1	PCR-Allele Specific Method	18
2.2.2	Primers	19
a.	tel-SMN Primer	20
b.	β -globin Primer	22
2.3	Agarose Gel Electrophoresis	24
2.4	DNA Purification	26
2.5	DNA Sequencing	27
2.5.1	Maxam-Gilbert Sequencing	28
2.5.2	Sanger-Coulson Sequencing	28
2.6	DNA Cycle Sequencing	31
CHAPTER 3 MATERIALS AND METHODS		33
3.1	Materials	33
3.1.1	Blood Sample	33
3.1.2	DNA Molecular Weight Markers	33
3.1.3	DNA Extraction Kit	33
3.1.4	Primers	34
3.1.5	DNA Purification Kit	34
3.1.6	DNA Sequencing Kit	34

3.2	Methods	35
3.2.1	Sample Collection	35
3.2.2	Human Genomic DNA Extraction (QIAamp Mini Kit)	35
a.	Proteinase K	35
b.	Buffer AL	36
c.	Ethanol (96-100%)	36
d.	Buffer AW1	36
e.	Buffer AW2	36
f.	Buffer AE	37
3.2.3	PCR-Allele Specific Amplification	39
a.	Deionized Distilled Water (ddH ₂ O)	41
b.	PCR Buffer	41
c.	Magnesium Chloride (MgCl ₂)	41
d.	Deoxynucleotides (dNTPs)	42
e.	Primers (forward and reverse)	42
f.	<i>Taq</i> DNA Polymerase	42
g.	DNA Template	43
3.2.4	Agarose Gel Electrophoresis	45
a.	DNA Ladder	45
b.	Loading Dye	45
c.	SYBR GreenI	46
d.	Tris-borate EDTA (TBE) Buffer	46
3.2.5	DNA Purification	48
a.	Buffer PB	48
b.	Buffer NW	48



c.	Buffer EB	48
3.2.6	DNA Cycle Sequencing	50
a.	Deionized Distilled Water (ddH ₂ O)	50
b.	Sequencing Buffer	51
c.	Big Dye	51
d.	Primers (forward and reverse)	51
e.	DNA Template	51
CHAPTER 4 RESULTS		55
4.1	Human Genomic DNA Extraction	55
4.2	PCR Amplification of β-globin Gene (Internal Control)	57
4.3	Optimization of Annealing Temperature (T _m)	58
4.3.1	PCR Amplification of SMN1 and β-globin Genes Using T _m = 58.4°C	59
4.4	Optimization of DNA Concentration	60
4.4.1	PCR Amplification of SMN1 and β-globin Genes Using 34 ng/μl DNA Template	61
4.5	DNA Purification	64
4.6	DNA Sequencing	66
CHAPTER 5 DISCUSSION		72
5.1	PCR-Allele Specific Amplification	72
5.1.1	Primers	72
a.	tel-SMN Primer	73
b.	β-globin Primer	73

5.1.2 Optimizations of PCR-Allele Specific Amplification	74
a. Annealing Temperature	74
b. DNA Template Quantity	75
5.1.3 PCR Master Mix and Parameters	75
a. PCR Master Mix	76
b. PCR Parameters	78
5.2 DNA Sequencing Analysis	79
CHAPTER 6 CONCLUSION	81
REFERENCES	83
APPENDIX A	92
APPENDIX B	93
APPENDIX C	94
APPENDIX D	95
APPENDIX E	96
APPENDIX F	97
APPENDIX G	98



LIST OF TABLES

Table No.		Page
2.1	tel-SMN primer used for amplification.	22
2.2	β -globin primer used for amplification.	24
3.1	The primers sets that are used for PCR amplification.	34
3.2	Reagents used for PCR amplification.	40
3.3	PCR parameters.	40
4.1	DNA quantification for normal healthy samples.	56
4.2	DNA quantification for samples from SMA patients.	60



LIST OF FIGURES

Figure No.		Page
2.1	SMA Type I — Joseph Michael Alcott, born July 24, 1995, Mullica Hill, NJ.	5
2.2	SMA Type II — Brooke Alexis-Lynn Akers, born July 25, 2000, West Chester, Ohio.	6
2.3	SMA Type III — Brady Nelson, born October 29, 2002, Jenks, OK.	7
2.4	Location of SMN1 gene on the long (q) arm of chromosome 5 at position 13.	10
2.5	In SMN1, a cytosine (C) located six nucleotides inside exon 7 directs for exon 7 inclusion, whereas a thymine (T) in SMN2 causes exon 7 exclusion.	12
2.6	Explanation of Mendel's First Law with two pure lines of breeding peas.	15
2.7	Probability of occurrences as a SMA patient from SMA carrier parents.	16



2.8	Polymerase Chain Reaction.	18
2.9	The complete sequence of exon 7 and 8 in <i>Homo sapiens</i> SMN1 gene.	21
2.10	Location of HBB gene on the short (p) arm of chromosome 11 at position 15.5.	23
2.11	The Sanger-Coulson method for sequencing DNA	30
4.1	Genomic DNA isolated from different healthy individuals.	56
4.2	PCR amplification of β -globin gene at 61 °C annealing temperature.	57
4.3	Gradient profile of tel-SMN primer at ± 5 °C from 58 °C.	58
4.4	PCR amplification of SMN1 and β -globin genes at 58.4 °C in the same normal healthy sample.	59
4.5	1.0 μ l DNA loaded for PCR amplification of SMN1 and β -globin genes at 58.4 °C.	62



4.6	2.0 μ l DNA loaded for PCR amplification of SMN1 and β -globin genes at 58.4 °C.	63
4.7	PCR amplification of SMN1 and β -globin genes at 58.4 °C.	64
4.8	Purified DNA of β -globin and SMN1 genes from a healthy sample.	65
4.9	ABI Electropherogram of β -globin gene (forward).	67
4.10	ABI Electropherogram of β -globin gene (reverse).	68
4.11	ABI Electropherogram of SMN1 gene (forward).	70
4.12	ABI Electropherogram of SMN1 gene (reverse).	71



LIST OF SYMBOLS

α	Alpha
β	Beta
D	The nonsuperimposable stereoisomer of (L)
L	The nonsuperimposable stereoisomer of (D)
F_1	The first filial generation
F_2	The second filial generation
S	Smooth seed
s	Wrinkled seed
p	Short arm of chromosome
q	Long arm of chromosome
3'	Three prime end of a DNA strand
5'	Five prime end of a DNA strand
+	Position of a nucleotide located in a positive direction within the sequence of a gene
-	Negative/Minus
\pm	Plus and minus
∞	Infinity
/	Per
\sim	Approximate
&	And
x	Multiply
\therefore	As a result



LIST OF UNITS

%	Percent
°C	Degree Celsius
ft	Feet
g	Gram
in	Inch
kb	Kilobase
min	Minute
ml	Milliliter
mM	Millimolar
ng	Nanogram
ng/ μ l	Nanogram Per Microliter
pH	A standard measure of the acidity of a solution
pmol/ μ l	Picomole Per Microliter
rpm	Rotation Per Minute
U	Unit
U/ μ l	Unit Per Microliter
μ l	Microliter
V	Volt
X	Times in concentration



LIST OF ABBREVIATIONS

A	Adenine
ABI	Applied Biosystem
bp	Base pairs
C	Cytosine
ddH ₂ O	Deionized Distilled Water
DNA	Deoxyribonucleic Acid
dNTPs	Deoxynucleotide Triphosphates (dATP, dCTP, dGTP, dTTP)
ddNTPs	Dideoxyribonucleoside Triphosphates
dsDNA	Double-Stranded Deoxyribonucleic Acid
EDTA	Ethylenediaminetetraacetic Acid
EtOH	Ethanol
ssDNA	Single-Stranded Deoxyribonucleic Acid
fwd	Forward
G	Guanine
HBB	Beta-hemoglobin
KCl	Potassium Chloride
Mg ²⁺	Magnesium Ion
MgCl ₂	Magnesium Chloride
mRNA	Mesenger Ribonucleic Acid
³² P	Radioactive Phosphorus Group
PCR	Polymerase Chain Reaction
RE	Restriction Enzyme
rvs	Reverse



SMA	Spinal Muscular Atrophy
SMN	Survival of Motor Neuron
SMN <i>cen</i>	Centromeric Survival of Motor Neuron 2 Gene
SMN <i>tel</i>	Telomeric Survival of Motor Neuron 1 Gene
ssDNA	Single Stranded Deoxyribonucleic Acid
T	Thymine
TBE	Tris-borate EDTA
tel-SMN	Telomeric Survival of Motor Neuron Primer
tel-SMNex7	Telomeric Survival of Motor Neuron Forward Primer (exon 7)
tel-SMNint7	Telomeric Survival of Motor Neuron Reverse Primer (intron 7)
T _m	Annealing Temperature
Tris-Cl	Tris-Chloride
Tris-HCl	Tris-Hydrogen Chloride
UV	Ultraviolet

CHAPTER 1

INTRODUCTION

Spinal Muscular Atrophy (SMA) is a common autosomal recessive neuromuscular disease of early life. Childhood-onset proximal SMA is characterized by degeneration of anterior horn cells of the spinal cord and, in some cases, of motor nuclei in brain stem. This degeneration causes the deficiency in the production of SMN protein which is essential for the health of motor neurons. Motor neurons are nerve cells that control muscles used for normal activities such as walking, sitting and swallowing. Consequently, the loss of motor neurons will lead to symmetrical muscles weakness and atrophy, limb and trunk paralysis, respiratory failure, and even infant death among the SMA patients.

SMA affects the voluntary muscles used for crawling, walking, head and neck control, and swallowing. The proximal muscles such as shoulders, hips, and back are often the most severely affected part. However, weakness in the legs is usually more severe than in the arms. As the child with SMA grows, the decrease in motor neurons and the increase demands on the nerve cells make his body doubly stressed. The resulting muscle weakness may lead to further loss of function especially the respiratory system which can lead to an increased tendency for pneumonia and other lung problems.



Clinically, SMA is divided into four groups based on certain key motor function milestones such as the age of onset, the maximum muscular activity achieved, and survivorship. Type I is the most severe form of SMA with onset at birth or before six months and death of respiratory distress usually within two years. This type of patients will never be able to sit or walk and have difficulties in breathing and swallowing. Type II is the intermediate form SMA which strikes infants between seven and 18 months old. These patients may be able to sit unaided or even stand with support. Type III SMA affects children after the age of 18 months and patients are able to stand and walk but may be unable to run. Type IV or the adult-onset SMA usually shows symptoms after age 35. The affected patients have muscle weakness, tremor, and twitching from mild to moderate (Families of SMA, 2005).

The survival of motor neuron (SMN) gene is the causing gene for SMA disease. It is present on chromosome 5 at position 5q13 in two inverted copies, telomeric SMN1 and centromeric SMN2. Both genes are nearly similar and encode for the same SMN protein (Lefebvre *et al.*, 1995). However, only SMN1 gene produces the full-length proteins while most SMN2 proteins are shortened due to the lacking of exon 7 in its mRNA. Therefore, SMA disease is caused by the deletions or mutations in SMN1 but not SMN2. The number of copies of SMN2 is only related to the severity of SMA disease. SMA patients are people who have both SMN1 copies deleted while the carriers are missing only one copy (Families of SMA, 2005). Approximately 95% of SMA patients are having homozygous deletion of SMN1 gene whereas the subsequent 5% are having other mutations (Lefebvre *et al.*, 1995).

Polymerase Chain Reaction (PCR)-based assays are always used for molecular diagnosis. A method called PCR-Allele Specific is one of the tests that developed on the basis of conventional PCR. The speciality of this method is the allele-specific primer that used for amplification because it is designed to only amplify particular desired allele. Since the primer has a high level of specificity in amplifying desired DNA fragment, thus, the resulted PCR products are always highly reliable (Cantafora *et al.*, 1998).

In this study, the PCR-Allele Specific method was applied to detect the homozygous deletion of SMN1 gene in three SMA patients from three to 14 years old. All these patients are referred to SMA molecular genetic test conducted at Human Genome Centre, Universiti Sains Malaysia Health Campus, Kubang Kerian, Kelantan. The first objective of this study is to amplify the specific exon 7 of SMN1 gene by using allele-specific tel-SMN primer and the second objective is to optimize the PCR reagents and parameters for the detection of homozygous deletion of SMN1 gene in SMA patients. The third objective is to confirm the actual size of both SMN1 and β -globin genes by using DNA sequencing.

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