

ABILITY OF *trnH-psbA* AND *trnL-trnF* INTERGENIC SPACERS  
TO DISCRIMINATE BETWEEN CLOSELY RELATED  
*Paphiopedilum* SPECIES

YEOH KEAT AI

PERPUSTAKAAN  
UNIVERSITI MALAYSIA SABAH

THESIS SUBMITTED TO FULFILL PART OF THE  
REQUIREMENTS TO OBTAIN A BACHELOR DEGREE IN  
SCIENCE WITH HONORS IN PLANT TECHNOLOGY

PLANT TECHNOLOGY PROGRAMME  
SCHOOL OF SCIENCE AND TECHNOLOGY  
UNIVERSITI MALAYSIA SABAH

2007



**UMS**  
UNIVERSITI MALAYSIA SABAH

## BORANG PENGESAHAN STATUS TESIS@

JUDUL: ABILITY OF TRNH-PSBA AND TRNL-TRNF INTERGENIC SPACER  
TO DISCRIMINATE BETWEEN CLOSELY PAPHIOPEDILUM SPECIES

Ijazah: ISAZAH SARJANA MUDA SAINS (TEKNOLOGI TUMBUHAN) DENGAN KEPUNJUAN

SESI PENGAJIAN: 2004

Saya YEON KEAT AI

(HURUF BESAR)

mengaku membenarkan tesis (LPS/Sarjana/Doktor Falsafah)\* ini disimpan di Perpustakaan Universiti Malaysia Sabah dengan syarat-syarat kegunaan seperti berikut:

- PERPUSTAKAAN**  
**UNIVERSITI MALAYSIA SABAH**
1. Tesis adalah hakmilik Universiti Malaysia Sabah.
  2. Perpustakaan Universiti Malaysia Sabah dibenarkan membuat salinan untuk tujuan pengajian sahaja.
  3. Perpustakaan dibenarkan membuat salinan tesis ini sebagai bahan pertukaran antara institusi pengajian tinggi.
  4. \*\*Sila tandakan ( / )

☐

SULIT

(Mengandungi maklumat yang berdarjah keselamatan atau kepentingan Malaysia seperti yang termaktub di dalam AKTA RAHSIA RASMI 1972)

☐

TERHAD

(Mengandungi maklumat TERHAD yang telah ditentukan oleh organisasi/badan di mana penyelidikan dijalankan)

☐

TIDAK TERHAD

Disahkan oleh

Keat Ai  
 (TANDATANGAN PENULIS)

Jey  
 (TANDATANGAN PUSTAKAWAN)

Alamat Tetap: NO 40, JLN 28/28,  
TAMAN ALAM MEGA, 40400

SHAH ALAM, SELANGOR, D.E.

Nama Penyelia

Tarikh: 16/4/2007

Tarikh: \_\_\_\_\_

CATATAN: \* Potong yang tidak berkenaan.

\*\* Jika tesis ini SULIT atau TERHAD, sila lampirkan surat daripada pihak berkuasa/organisasi berkenaan dengan menyatakan sekali sebab dan tempoh tesis ini perlu dikelaskan sebagai SULIT dan TERHAD.

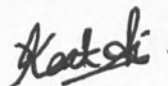
@ Tesis dimaksudkan sebagai tesis bagi Ijazah Doktor Falsafah dan Sarjana secara penyelidikan, atau disertasi bagi pengajian secara kerja kursus dan penyelidikan, atau Laporan Projek Sarjana Muda (LPSM).



## DECLARATION

I hereby declare that no part of this thesis has been previously submitted for a degree in this or any other university. All work were based on my own research except as cited in references and other part of the thesis where indicated.

12 March 2007



---

YEOH KEAT AI

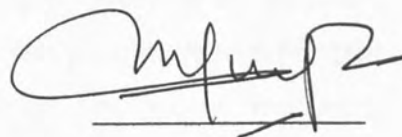
HS2004-2775

PERPUSTAKAAN  
UNIVERSITI MALAYSIA SABAH

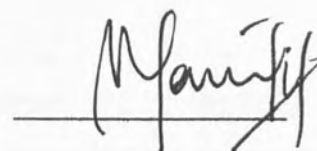


**VERIFIED BY****Verified by:****Signature**

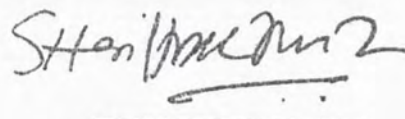
1. **SUPERVISOR**  
**(DR. VIJAY KUMAR)**



2. **EXAMINER 1**  
**(ASSOCIATE PROFESSOR**  
**DR. MARIAM ABD. LATIP)**



3. **DEAN**  
**(SUPT/ KS ASSOCIATE PROFESSOR**  
**DR. SHARIFF A. K OMANG)**



**PERPUSTAKAAN**  
**UNIVERSITI MALAYSIA SABAH**



**UMS**  
UNIVERSITI MALAYSIA SABAH



## ACKNOWLEDGEMENT

With the completion of this project, first and foremost, I would like to express my utmost appreciation and gratitude to my supervisor Dr. Vijay Kumar for his guidance, advice and supervision throughout the entire duration of this project. Special heartfelt thanks are also extended to Mr. Kenneth Rodrigues for his tireless assistance, guidance and advice during the whole duration of this project. I thank them very much for their patience and tolerance.

My sincere appreciation also goes to other postgraduate students of Biotechnology Research Institute especially Awang, Thien, Melvin and Adrian who had provided me guidance, assistance and support whenever I needed them. I would also like to thank Mr. Richard, Ms. Vidarita and Cik Azian for their assistance in the laboratory.

A token of appreciation goes also to all other Biotechnology Research Institute undergraduate students who have been through thick and thin with me. Doing research with all of them were enjoyable and it was a memorable experience for me. Last but not least, I would like to extend my gratitude to my family members, friends and coursemates who has provided me their endless love and also emotional and moral support.



## ABSTRACT

Two intergenic spacers, *trnH-psbA* and *trnL-trnF* of the chloroplast DNA (cpDNA) were used to determine whether they can be used to distinguish between three closely related *Paphiopedilum* species namely *P. rothschildianum*, *P. lowii* and *P. dayanum*. Genomic DNA was extracted from the leaves of the three *Paphiopedilum* species without the use of phenol chloroform. Next, the two loci were amplified through PCR using two primers pairs (*psbAF* – *trnHR* and *trnLF* – *trnFR*). The amplified PCR products were ligated into the pCR 2.1 – TOPO vector, transformed using TOP 10 chemically competent cells. Then, plasmid miniprep was performed and the plasmids obtained were sequenced. Data analysis was performed using the resulting sequences. A total of 29 indels mutations points with 95 indels were observed, accounting for *trnH-psbA* intergenic spacer length of *P. rothschildianum*, *P. lowii* and *P. dayanum* with a range of 924 to 1000 bp. In the *trnL-trnF* intergenic spacer, 11 indels mutation points with 48 indels have been discovered for the three *Paphiopedilum* species with a range of between 462 to 490 bp. The genetic similarity and differences between the *Paphiopedilum* species was examined by calculating the pairwise nucleotide differences that occurred in each intergenic region. The result showed that *P. rothschildianum* and *P. lowii* were more closely related compare to *P. dayanum*. Single nucleotide polymorphisms (SNP) were then determined and *trnH-psbA* intergenic spacer was found to provide 96 informative sites which indicate the variations that occur in this spacer region was higher compared to *trnL-trnF* spacer region which provide 48 informative sites. The phylogenies of the three species were also examined by constructing phylogenetic trees and confirmed by performing bootstrapping analysis. The results obtained indicate that *trnL-trnF* intergenic spacer is more reliable compared to the *trnH-psbA* intergenic region for distinguishing between the three *Paphiopedilum* species. However, more research should to be conducted on *trnH-psbA* intergenic spacer as it exhibit sufficient nucleotide variations which proved that it is also a potentially good marker for phylogenetics studies and DNA barcoding of other plants.





## ABSTRAK

Dua *intergenic spacer*, *trnH-psbA* dan *trnL-trnF* dari DNA kloroplas dikaji untuk menentukan sama ada dua *intergenic spacer* ini sesuai digunakan untuk membezakan antara tiga spesies *Paphiopedilum* yang saling berkait rapat iaitu *P. rothschildianum*, *P. lowii* dan *P. dayanum*. DNA genomik diekstrak daripada daun tiga spesies *Paphiopedilum* tanpa menggunakan fenol kloroform. Kemudian, dua *spacer* tersebut diamplifikasikan menggunakan dua pasangan primer (*psbAF* – *trnHR* dan *trnLF* – *trnFR*). Produk PCR yang terhasil diklon dengan vektor pCR 2.1 – TOPO and ditransformasikan dalam TOP 10 *competent cell*. Selepas itu, plasmid diekstrak dan penjujukan DNA dilakukan. Analisis data dilakukan dengan menggunakan jujukan DNA yang diperolehi. Bagi *trnH-psbA intergenic spacer*, sejumlah 29 kawasan tumpuan mutasi dengan 95 *indels* telah diperhatikan dalam kawasan *spacer* yang bersaiz dalam lingkungan 924 hingga 1000 pasangan bes bagi ketiga-tiga spesies *Paphiopedilum*. Manakala bagi *trnL-trnF intergenic spacer* pula, 11 kawasan tumpuan mutasi dengan 48 *indels* telah dikenalpasti daripada tiga spesies yang bersaiz dalam lingkungan 462 hingga 490 pasangan bes. Persamaan dan perbezaan genetik diantara spesies-spesies ditentukan dengan mengira perbezaan nukleotid secara berpasangan antara tiga spesies tersebut bagi kedua-dua *intergenic spacer*. Keputusan yang diperolehi menunjukkan bahawa *P. lowii* dan *P. rothschildianum* mempunyai persamaan yang paling ketara berbanding *P. dayanum*. *Single nucleotide polymorphisms (SNP)* kemudiannya ditentukan dan didapati *trnH-psbA intergenic spacer* mempunyai bilangan kawasan informatif yang lebih banyak yakni 96 berbanding *trnL-trnF intergenic spacer* yang mempunyai 48 kawasan informatif. Bilangan kawasan informatif yang lebih banyak dalam *trnH-psbA intergenic spacer* menunjukkan *spacer* ini mempunyai lebih variasi berbanding *trnL-trnF intergenic spacer*. Hubungan filogenetik diantara tiga spesies *Paphiopedilum* dikaji dengan menggunakan pokok filogenetik dan kemudian dengan analisis *bootstrap*. Hasil kajian menunjukkan bahawa *trnL-trnF intergenic spacer* adalah lebih sesuai digunakan untuk membezakan antara tiga spesies *Paphiopedilum*. Namun, lebih banyak kajian keatas *trnH-psbA spacer* patut dijalankan kerana, *spacer* ini juga menunjukkan variasi nukleotid yang tinggi dan berpotensi untuk dijadikan penanda molekul untuk kajian filogenetik and barkod DNA bagi tumbuhan lain.



## TABLE OF CONTENTS

	Page
DECLARATION	ii
VERIFICATION	iii
ACKNOWLEDGEMENT	iv
ABSTRACT	v
ABSTRAK	vi
TABLE OF CONTENTS	vii
LIST OF TABLES	x
LIST OF FIGURES	xii
LIST OF ABBREVIATIONS	xiv
<b>CHAPTER 1            INTRODUCTION</b>	
1.1    Preface	1
1.2    Research Objectives	5
<b>CHAPTER 2            LITERATURE REVIEW</b>	
2.1 <i>Paphiopedilum</i> Orchids	6
2.1.1 <i>Paphiopedilum rothschildianum</i>	8
2.1.2 <i>Paphiopedilum dayanum</i>	9
2.1.3 <i>Paphiopedilum lowii</i>	10
2.2    Chloroplast DNA (cpDNA)	11
2.2.1    Gene Map of Chloroplast DNA	13
2.2.2    Coding and Non-coding Regions of cpDNA	15
2.2.3    Defining <i>trnH</i> , <i>psbA</i> , <i>trnL</i> and <i>trnF</i> Genes and Other Genes in Chloroplast DNA	16
2.3    DNA Barcoding and Barcode of Life	17
2.4    Occurrence of Indels in <i>trnH-psbA</i> and <i>trnL-trnF</i> Intergenic Spacers	20





### CHAPTER 3                      METHODOLOGY

3.1	Sampling of Plant Material	22
3.2	DNA Extraction	23
3.3	Agarose Gel Electrophoresis and DNA Quantification	24
3.4	PCR Primers for Amplification of <i>trnH-psbA</i> and <i>trnL-trnF</i> Intergenic Spacers	26
3.5	Screening of <i>trnH-psbA</i> and <i>trnL-trnF</i> Primers using PCR	27
3.6	Cloning of PCR Product ( <i>trnH-psbA</i> and <i>trnL-trnF</i> Intergenic Spacers)	29
3.6.1	Insertion and Transformation of Vector into <i>E. coli</i> Cells	29
3.6.2	Plating of Competent Cells	30
3.6.3	Blue White Screening and Sub-culturing of Colonies with Transformed Cells	30
3.6.4	Culture of White Colonies	31
3.6.5	Preparing Solution I, II and III for Plasmid Miniprep	32
3.6.6	Plasmid Miniprep	32
3.7	Restriction Enzyme (RE) Digestion of Plasmid DNA	34
3.8	DNA Sequencing	35
3.9	Submission of Sequences to GenBank	35
3.10	Data Analysis	36

### CHAPTER 4                      RESULTS

4.1	DNA Extraction	39
4.2	Analysis of PCR Amplified Intergenic Regions of <i>trnH-psbA</i> and <i>trnL-trnF</i>	40
4.3	Cloning of PCR Product	42
4.4	Analysis of Positive Clones	44
4.5	DNA Sequences	53
4.6	Homology Test and Comparison using Basic Local Alignment Search Tool (BLAST)	59
4.7	Multiple Sequence Alignment and Single Nucleotide Polymorphisms in <i>trnH-psbA</i> and <i>trnL-trnF</i> Intergenic Spacer	61
4.8	Calculation of Single Nucleotide Polymorphisms	69
4.9	Phylogenetic Analysis of the <i>trnH-psbA</i> and <i>trnL-trnF</i> Intergenic Spacers Sequences and Indels	70



<b>CHAPTER 5</b>	<b>DICUSSION</b>	
5.1	Leaf Sample Preservation and DNA Extraction	81
5.2	Primer Screening and PCR Optimization	84
5.3	Cloning of PCR Products	87
5.4	Plasmid Minipreparation and Restriction Enzyme (RE) Digestion	89
5.5	DNA Sequencing of <i>trnH-psbA</i> and <i>trnL-trnF</i> Intergenic Spacers	91
5.6	Homology Test and Comparison using Basic Local Alignment Search Tool (BLAST)	94
5.7	Multiple Sequence Alignment	96
5.8	Pairwise Nucleotide Differences between the <i>Paphiopedilum</i> species	97
5.9	Single Nucleotide Polymorphisms Calculations	99
5.10	Phylogenetic Relationship of the <i>Paphiopedilum</i> species using based on both Intergenic Spacers	100
<b>CHAPTER 6</b>	<b>CONCLUSION</b>	102
REFERENCES		105
APPENDIX		113



## LIST OF TABLES

No.	Table	Page
3.1	Nucleotide sequences of the PCR primers for amplification of <i>trnH-psbA</i> intergenic spacer	26
3.2	Nucleotide sequences of the PCR primers for amplification of <i>trnL-trnF</i> intergenic spacer	27
3.3	Calculations for each component of the RE reaction mixture	34
4.1	Taxa used in the data analysis, with the place of origin and accession number for each sequence	54
4.2	Complete DNA Sequence of <i>trnH-psbA</i> Intergenic Spacer of <i>P. dayanum</i> (1000 bp)	55
4.3	Complete DNA Sequence of <i>trnH-psbA</i> Intergenic Spacer of <i>P. lowii</i> (924 bp)	56
4.4	Complete DNA Sequence of <i>trnH-psbA</i> Intergenic Spacer of <i>P. rothschildianum</i> (926 bp)	57
4.5	Complete DNA Sequence of <i>trnL-trnF</i> Intergenic Spacer of <i>P. dayanum</i> (462 bp)	58
4.6	Complete DNA Sequence of <i>trnL-trnF</i> Intergenic Spacer of <i>P. lowii</i> (484 bp)	58
4.7	Complete DNA Sequence of <i>trnL-trnF</i> Intergenic Spacer of <i>P. rothschildianum</i> (490 bp)	59





4.8	Result of homology test of <i>P. dayanum</i> , <i>P. lowii</i> and <i>P. rothschildianum</i> based on the sequences obtained for <i>trnH-psbA</i> and <i>trnL-trnF</i> intergenic spacers	60
4.9	Multiple sequence alignment of the nucleotide sequences of the intergenic spacer between the <i>trnH</i> (GUG) and <i>psbA</i> gene	61
4.10	Pairwise nucleotide differences calculation between <i>P. dayanum</i> , <i>P. lowii</i> and <i>P. rothschildianum</i> in <i>trnH-psbA</i> intergenic spacer	64
4.11	Occurrence of indels in <i>trnH-psbA</i> intergenic spacer of three <i>Paphiopedilum</i> species	65
4.12	Multiple sequence alignment of nucleotide sequences of the intergenic spacer between the <i>trnL</i> (UAA) and <i>trnF</i> (GAA) gene	66
4.13	Pairwise nucleotide differences calculation between <i>P. dayanum</i> , <i>P. lowii</i> and <i>P. rothschildianum</i> in <i>trnL-trnF</i> intergenic spacer	67
4.14	Occurrence of indels in <i>trnL-trnF</i> intergenic spacer of three <i>Paphiopedilum</i> species	68
4.15	Comparison of phylogenetic information from variable sites for two intergenic spacers, <i>trnH-psbA</i> and <i>trnL-trnF</i>	69
4.16	Genetic distance obtained using Kimura distances model and standard errors for the chloroplast <i>trnH-psbA</i> intergenic spacer of <i>P. dayanum</i> , <i>P. lowii</i> and <i>P. rothschildianum</i>	71
4.17	Genetic distance obtained using Kimura distances model and standard errors for the chloroplast <i>trnL-trnF</i> intergenic spacer of <i>P. dayanum</i> , <i>P. lowii</i> , and <i>P. rothschildianum</i>	72



## LIST OF FIGURES

No.	Figure	Page
2.1	Gene map of tobacco chloroplast DNA and the locations of the <i>trnH-psbA</i> and <i>trnL-trnF</i> intergenic spacers	14
4.1	Profile of extracted <i>Paphiopedilum</i> DNA	40
4.2	PCR amplification of the <i>trnH-psbA</i> intergenic regions of the three <i>Paphiopedilum</i> species	41
4.3	PCR amplification of the <i>trnL-trnF</i> intergenic regions of the three <i>Paphiopedilum</i> species	42
4.4	The formation of blue white colonies observed in one of the cloning process. (cloning of <i>P. lowii</i> for <i>trnH-psbA</i> intergenic region)	43
4.5a	Plasmids derived from the white colonies (expected with inserts) after plasmid miniprep (Plasmids that carry the insert of <i>trnH-psbA</i> intergenic region of <i>P. dayanum</i> )	45
4.5b	Digested plasmid using <i>EcoRI</i> restriction enzyme (Plasmids that carry the insert of <i>trnH-psbA</i> intergenic region of <i>P. dayanum</i> )	46
4.6a	Plasmids derived from the white colonies (expected with inserts) after plasmid miniprep (Plasmids that carry the insert of <i>trnH-psbA</i> intergenic region of <i>P. lowii</i> )	47
4.6b	Digested plasmid using <i>EcoRI</i> restriction enzyme (Plasmids that carry the insert of <i>trnH-psbA</i> intergenic region of <i>P. lowii</i> )	48



4.7a	Plasmids derived from the white colonies (expected with inserts) after plasmid miniprep (Plasmids that carry the insert of <i>trnH-psbA</i> intergenic region of <i>P. rothschildianum</i> )	49
4.7b	Digested plasmid using <i>EcoRI</i> restriction enzyme (Plasmids that carry the insert of <i>trnH-psbA</i> intergenic region of <i>P. rothschildianum</i> )	50
4.8	Plasmid from the cloning of <i>trnL-trnF</i> intergenic region of <i>P. lowii</i> and the <i>EcoRI</i> digested plasmid	51
4.9	Plasmids derived from the white colonies (expected with inserts) after plasmid miniprep (Plasmids that carry the insert of <i>trnL-trnF</i> intergenic region of <i>P. dayanum</i> )	52
4.10	Example of cloning failure observed for cloning of <i>trnL-trnF</i> intergenic spacer of <i>P. rothschildianum</i>	53
4.11	Neighbor-joining tree generated from Kimura distances model for the <i>trnH-psbA</i> intergenic spacer sequences of <i>P. rothschildianum</i> , <i>P. lowii</i> , <i>P. dayanum</i> and 10 other plant species used for comparison using NTSYS	74
4.12	Neighbor-joining tree generated from Kimura distances model for the <i>trnL-trnF</i> intergenic spacer sequences of <i>P. rothschildianum</i> , <i>P. lowii</i> , <i>P. dayanum</i> and 10 other plant species used for comparison using NTSYS	77
4.13	Consensus tree generated by PHYLIP bootstrap method with 1000 replicates for analyzing the confidence of genetic relationship between three <i>Paphiopedilum</i> species ( <i>trnH-psbA</i> intergenic spacer)	79
4.14	Consensus tree generated by PHYLIP bootstrap method with 1000 replicates for analyzing the confidence of genetic relationship between three <i>Paphiopedilum</i> species ( <i>trnL-trnF</i> intergenic spacer)	80





## LIST OF ABBREVIATIONS

WCE	Sabah Wildlife Conservation Enactment
CBOL	The Consortium for the Barcode of Life
CITES	Convention on International Trade in Endangered Species
RM	Malaysian Ringgit
<i>COI</i>	<i>Cytochrome c oxidase subunit I</i> gene
DNA	Deoxyribonucleic acid
gDNA	Genomic deoxyribonucleic acid
mtDNA	Mitochondrial deoxyribonucleic acid
cpDNA	Chloroplast deoxyribonucleic acid
rRNA	Ribosomal ribonucleic acid
tRNA	Transfer ribonucleic acid
RNase	Ribonuclease
DNase	Deoxyribonuclease
BLAST	Basic Local Alignment Search Tool
SNP	Single nucleotide polymorphisms
SSR	Simple Sequence Repeat/ Microsatellite
RFLP	Restriction Fragment Length Polymorphism
RAPD	Random Amplified Polymorphism DNA
AFLP	Amplified Fragment Length Polymorphism
PCR	Polymerase Chain Reaction
NaCl	Sodium chloride
NaOH	Sodium hydroxide
EDTA	Ethylenediamine tetraacetic acid
SDS	Sodium dodecyl sulphate
KCl	Potassium acetate
TBE	Tris-Borate-EDTA
LB	Luria-Bertani
BSA	Bovine Serum Albumin
<i>E. coli</i>	<i>Escherichia coli</i>
X-Gal	5-bromo-4-chloro-3-indolyl- $\beta$ -D-galactopyranoside
UV	ultra-violet



A	Adenine
T	Thymine
C	Cytosine
G	Guanine
g	gram
rpm	round per minute
min	minute (s)
s	second (s)
kb	kilo base pair
bp	base pair
M	molar
mM	milimolar
mmol	milimol
pmol	picomol
ml	mililiter
$\mu$ l	microliter
L	liter
V	volt
U	unit
%	percent
$^{\circ}$ C	degree Celsius



## CHAPTER 1

### INTRODUCTION

#### 1.1 Preface

Malaysia is a home to about 3000 species of exotic, attractive and unique orchid species and many of these are native to Malaysia. Orchids usually grow well in the jungles, forests, mountains and islands. On mountainous areas, the orchids usually grow at the altitude of 1000 to 6000 meters above sea level. For instance, the largest species of all orchids, the *Grammatophyllum* can be found in the mountainous areas of the Main Range of Peninsular Malaysia (Fadelah *et al.*, 2001). In Sabah and Sarawak, unique orchid species such as the *Paphiopedilum* species orchids can be found.

The genus *Paphiopedilum* belongs to the family of Orchidaceae and subfamily of *Cypripedioideae*. In fact, *Paphiopedilum* orchids are closely related to *Cypripedium*, *Phragmipedium* and *Selenipedium*. They are all referred to as *Cypripedium* or 'Cyps' prior to being separated to their respective genera by Rolfe in 1896. The distribution of *Paphiopedilum* orchids extend from India to Burma, Southeast Asia, South China,





Malaysia, Java, Borneo, Philippines and Papua New Guinea to the Solomon Islands (Schoser, 1981). There are about 50 species of *Paphiopedilum* that are native to tropical Asia and five of which are found growing in Malaysia.

The name of *Paphiopedilum* is derived from a Greek word *paphos*, which means the 'island with a temple dedicated to Venus' and also *pedilon* which means 'sandal' (Teo, 1985). The common name of the ladyslipper orchid is given due to the appearance of the lip of the *Paphiopedilum* flower which resembles a lady's slipper. Due to the unique appearance of the flowers, *Paphiopedilum* species are always in high demand and popular among orchid lovers.

In Sabah, *Paphiopedilum* is zealously protected by the Mount Kinabalu National Park since the number of *Paphiopedilum* plants that are growing in the wild has been decreasing drastically and is facing extinction. The *Paphiopedilum* orchids in Sabah are facing enormous threats because the Kinabalu National Park borders are constantly being encroached by mining, lumbering, agricultural projects and also the practice of shifting agriculture by illegal squatters (Cribb, 1998). There are nine species of *Paphiopedilum* orchids that are listed under the Sabah Wildlife Conservation Enactment (WCE). The penalty for owning one of the species listed can be a fine of a maximum of RM 50 000, five years jail or both (Kaur and John, 2006).

Other than threats caused by development activities as mentioned, many orchid sellers and some handicrafts owners are not aware of the value of the endangered *Paphiopedilum* orchids. These orchids are planted or collected from the wild to be sold to tourists and orchid lovers at very low price rather than the actual



price which they are worth. On the other hand, most of the orchid lovers are aware of the penalty for owning the *Paphiopedilum* species orchid. However, the protected status of these orchids has only stir up more interest in them for possessing the plant. It has also been reported that these precious orchids are smuggled out of Sabah and to be sold in other countries (Kaur and John, 2006). The *Paphiopedilum* orchids are in very high demand especially in the temperate countries and they can fetch up to very high price (Cribb, 1998).

The continuous smuggling activities of *Paphiopedilum* have proved that the protection by the Kinabalu National Park and the penalty implemented are insufficient to protect the “Paph” orchids from being removed from its natural habitat. Efforts to prevent smuggling of the protected *Paphiopedilum* species are often not effective as the smugglers of these precious orchids can easily get away from the law as they can deny for possessing or selling the endangered *Paphiopedilum* plants. This is due to the reason as some of these endangered *Paphiopedilum* species has similar appearance like the ones which are non-protected species. In most cases, people are unable to distinguish between the endangered *Paphiopedilum* species and the non-protected ones.

Even at present, most of the orchid experts and experienced researchers are still differentiating *Paphiopedilum* orchids by the physical characteristics on the plants as there are no established or convincing methods to distinguish between similar looking species. Due to this reason, the use of DNA barcoding, in particular the intergenic region *trnH-psbA*, could be used to determine whether it is able to





discriminate between morphologically similar or closely allied *Paphiopedilum* species.

Using the concept of DNA barcoding, a uniform region of a genome in animals or plants is used to distinguish between different animals or plants species. Generally, the *cytochrome c oxidase subunit I (COI)* gene is used to tag animals whereas the *trnH-psbA* intergenic spacer is has been used in tagging angiosperms. DNA barcoding enable different animals and plants species to be clearly distinguished. Thus, at the same time it can also help in solving problems regarding closely related animals or plants species.

Applying the DNA barcoding concept in this research, the *trnH-psbA* intergenic spacer can be used to determine whether it would serve as a good marker to discriminate between few closely allied *Paphiopedilum* species. The *Paphiopedilum* orchid species that will be used are *Paphiopedilum rothschildianum*, *P. dayanum*, and *P. lowii* which are currently available at Sabah Park Orchid Conservatory. Based on the concept of DNA barcoding, the nucleotide sequence in the *trnH-psbA* intergenic region in all the three species of *Paphiopedilum* will be determined. Each species of *Paphiopedilum* species is expected to produce a set of unique nucleotides sequence of their *trnH-psbA* intergenic region.

The specific sets of nucleotide sequences of the *trnH-psbA* intergenic spacer will serve as barcodes and tag each of the species, thus help in solving ambiguities between closely related *Paphiopedilum* species. Moreover, should any problem arise in the future whether a person is possessing or selling endangered *Paphiopedilum*





orchids, the uncertainty can be solve by checking the barcodes of the *Paphiopedilum* plants. Other than tagging closely related *Paphiopedilum*, knowing the sequences of the *trnH-psbA* intergenic spacer will also show the relationship between the species. Generally, a species is said to be closely allied to another species if there is high percentage of nucleotide sequences similarity between the two species.

## 1.2 Research Objectives

The objectives of this research are:

- 1.2.1 To obtain the DNA nucleotide sequences of the *trnH-psbA* intergenic regions of *Paphiopedilum dayanum*, *P. lowii* and *P. rothschildianum*, respectively.
- 1.2.2 To perform multiple sequence alignment of the *trnH-psbA* intergenic spacer and to determine the number of single nucleotide polymorphism in *P. dayanum*, *P. lowii* and *P. rothschildianum*.
- 1.2.3 To examine the genetic relationship of the three *Paphiopedilum* species by constructing a phylogenetic tree based on the genetic distances obtained from pairwise nucleotide differences.

## CHAPTER 2

### LITERATURE REVIEW

#### 2.1 *Paphiopedilum* Orchids

The *Paphiopedilum* orchids are sympodial plants with narrow and elongated leaves. The leaves are green color or mottled and the surface of the leaves always appear waxy. New buds and plantlets often arise from the rhizome of the plant. Other than that, the ladyslipper orchids are also pseudobulbless and stemless. Even if they have stems, the stems are too short and it stops growing after some time. Generally, the *Paphiopedilum* species are able to flower throughout the year. Flowers of *Paphiopedilum* can be single or multiple and the flowers are usually in wide variety of form and color and can last for a very long period.

The *Paphiopedilum* orchids often grow well in the mountain forest where the grounds were covered with fallen leaves and twigs, limestone cliffs and on the granite cliffs. These habitats provide essential requirement for the *Paphiopedilum* roots to grow. *Paphiopedilum* plants should not get completely dry or too wet either as they are unable to store water. Most of the *Paphiopedilum* are often found growing best in



the monsoon areas which have dry and wet season instead of the tropical rain forests which receive plenty of rain throughout the year.

Temperature plays a very important role in determining the blossom of *Paphiopedilum* flowers. Generally, *Paphiopedilum* species grow best between the temperature range of 15 °C to 30 °C. However, to be more specific, the origin of each *Paphiopedilum* species determines the optimum temperature for the particular species to grow well. Often, the ones that originated from the lowland need higher temperatures like the tropical temperatures and as for the highland species, they need lower temperatures.

This orchid is a protected genus under Malaysia legislation and also listed on Appendix I of the Convention on International Trade in Endangered Species (CITES). Therefore, the trade of *Paphiopedilum* is strictly banned and ownership of these orchids is regulated with permits (Kaur and John, 2006). The endangered and protected *Paphiopedilum* species such as *Paphiopedilum rothschildianum* are found growing well on Mount Kinabalu, Sabah. Other than *P. rothschildianum*, other *Paphiopedilum* species orchids that were found grow on Mount Kinabalu since 1980s include *P. dayanum*, *P. lowii*, *P. volonteatum*, *P. petri*, *P. stonei*, *P. virens*, *P. hookerae*, *P. philippinense* and many other species.





### 2.1.1 *Paphiopedilum rothschildianum*

The *P. rothschildianum* orchid which originated from Borneo was named after Baron Ferdinand de Rothschild (Fowlie and Lamb, 1983). This orchid was introduced by Frederick Sander in 1888 and was crowned the 'King of Orchids' (Bechtel, 1983). *P. rothschildianum* population is one of the rarest in the *Paphiopedilum* genus as it is a species that only can be found in Mount Kinabalu, Sabah and they are protected by the Mount Kinabalu National Park. Unfortunately, this orchid species is facing great threats as their habitat is seriously destructed by development activities such as logging, mining, agriculture projects and illegal squatters that carry out shifting agriculture as part of their living (Cribb, 1998). Recently, it is also reported that this precious species was sold to tourists with extremely low price by the local handicraft owners (Kaur and John, 2006).

*P. rothschildianum* grow well at the slopes of Mount Kinabalu at about 1500 feet to 3000 feet above sea level and perform best in cool and shady conditions. In nature, it is seen in the steep ravines in the streams or on the ultrabasic rocks (Fowlie and Lamb, 1983). Seedlings of *P. rothschildianum* often sprout on lithophytic mosses and when mature, they are rooted lithophytically onto the cliff trees.

The leaves of mature *P. rothschildianum* can grow up to two feet long by 2½ inches in breadth. These hard and long-petal leaves are droopier looking compared to the leaves of *P. lowii*. The leaves catch leaf litters and debris which adds to the compost for the adult plants. Normally, the fallen leaves of other trees, for instance,



## REFERENCES

- Aldrich, J., Cherney, B. W. & Merlin, E. 1988. The role of insertion/ deletion in the evolution of the intergenic region between *psbA* and *trnH* in the chloroplast genome. *Current Genetics* **14**, 137-146.
- Attwood, T. K. & Parry-Smith, D. J. 1999. *Introduction to Bioinformatics*. Addison Wesley Longman Limited, London, pg. 108-131.
- Ausubel, F. M., Brent, R., Kingston, R. F., Moore, D. D., Seidman, J. G., Smith, J. A. & Struhl, K. (eds) 2002. *Short Protocols in Molecular Biology*. 5<sup>th</sup> Ed. John Wiley & Sons, Inc., Canada, pg. 2-1 and 2-13.
- Bechtel, P. G. 1983. *Paphiopedilum rothschildianum* and its hybrids (a guide to success with temperamental orchids). *The Orchid Digest* **47**, 165-169.
- Bedbrook, J. R. & Bogorad, L. 1976. Endonuclease recognition sites mapped on *Zea mays* chloroplast DNA. *Proceedings of the National Academy of Science USA* **73**, 4309-4313.
- Benson, D. A., Karsch-Mizrachi, I., Lipman, D. J., Ostell, J., Rapp, B. A. & Wheeler, D. L. 2002. GenBank. *Nucleic Acid Research* **30**, 17-20.
- Booth, F. L. 1982. The hard, strap-leaved *Paphiopedilum* species and hybrids. *The Orchid Digest* **46**, 45-48.
- Bremer, B., Bremer, K., Heidari, N., Erixon, P., Olmstead, R. G., Anderberg, A. A., Källersjö, M. & Barkhordarian, E. 2002. Phylogenetics of asterids based on 3 coding and 3 non-coding chloroplast DNA markers and the utility of non-coding DNA at higher taxonomic levels. *Molecular Phylogenetics and Evolution* **24**, 274-301.
- Brownlee, C. 2004. DNA bar codes: life under the scanner. *Science News* **166**, 360-361.





- Chandler, G. T., Bayer, R. J. & Crisp, M. D. 2001. A molecular phylogeny of the endemic Australian genus *Gastrolobium* (Fabaceae: Mirbelieae) and allied genera using chloroplast and nuclear markers. *American Journal of Botany* **88**, 1675-1687.
- Cribb, P. 1998. *The Genus Paphiopedilum*. 2<sup>nd</sup> Ed. Natural History Publications (Borneo), Kota Kinabalu, pg. 177-182, 162-168 and 349-353.
- Chris Teo, K. H. 1985. *Native Orchids of Peninsulas Malaysia*. Times Books International, Singapore, pg. 66-69.
- Chung, S. Y., Choi, S. H., Kim, M. J., Yoon, K. E., Lee, G. P., Lee, J. S. & Ryu, K. H. 2006. Genetic relationship and differentiation of *Paphiopedilum* and *Phragmepedium* based on RAPD analysis. *Scientia Horticulturae* **109**, 153-159.
- Csaikl, U. M., Bastian, H., Brettschneider, R., Gauch, S., Meir, A., Schauerte, M., Scholz, F., Sperisen, C., Vornam, B. & Ziegenhagen, B. 1998. Comparative analysis of different DNA extraction protocols: a fast, universal maxi-preparation of high quality plant DNA for genetic evaluation and phylogenetic studies. *Plant Molecular Biology Reporter* **16**, 69-86.
- Dellaporta, S. L., Woods, J. & Hicks, J. B. 1983. A plant DNA mini-preparation Version II. *Plant Molecular Biology Reporter*, pg. 19-21.
- Deprés, L., Gielly, L., Redoutet, B. & Taberlet, P. 2003. Using AFLP to resolve phylogenetic relationships in a morphologically diversified plant species complex when nuclear and chloroplast sequences fail to reveal variability. *Molecular Phylogenetics and Evolution* **27**, 185-196.
- Eckert, K. A. & Kunkel, T. A. 1990. High fidelity DNA synthesis by the *Thermus aquaticus* DNA polymerase. *Nucleic Acids Research* **18**, 3739-3744.





- Fadelah, A. A., Rozaily, H., Nuraini, I., Tan, S. L. & Hamidah, S. 2001. *Orchids: The Living Jewels of Malaysia*. Malaysian Agricultural Research and Development Institute, Kuala Lumpur, pg. 19-30.
- Felsenstein, J. 1985. Confidence limits on phylogenies: an approach using the bootstrap. *Evolution* **39**, 783-79.
- Fowlie, J. A. 1981. Part XVIII: The occurrence of *Paphiopedilum virens* (Rchb.f.) Pfitz on granite boulders with leaf detritus on Mt. Kinabalu, Sabah, North Borneo. *The Orchid Digest* **45**, 85-90.
- Fowlie, J. A. & Lamb, T. 1983. Part XXIV: *Paphiopedilum dayanum* and *Paphiopedilum rothschildianum* in serpentine cliffs on Mount Kinabalu, eastern Malaysia. *The Orchid Digest* **47**, 175-186.
- Fowlie, J. A. 1985. Part XXIX: Rediscovering the habitat of *Paphiopedilum dayanum* on serpentine cliffs on Mount Kinabalu in eastern Malaysia (formerly north Borneo). *The Orchid Digest* **49**, 125-129.
- Hajibabaei, M., deWaard, J. R., Ivanova, N. V., Ratnasingham, S., Dooh, R. T., Kirk, S. L., Mackie, P. M. & Hebert, P. D. N. 2005. Critical factors for assembling a high volume of DNA barcodes. *Philosophical Transactions of the Royal Society: B series*. **360**, 1959-1967.
- Hajibabaei, M., Janzen, D. H., Burns, J. M., Hallwachs, W. & Hebert, P. D. N. 2005. DNA barcodes distinguish species of tropical Lepidoptera. *Proceedings of the National Academy of Science USA* **103**, 968-971.
- Ham, R. C. H. J. van, Hart, Henk't, Mes, T. H. M., Sandbrink, J. M. 1994. Molecular evolution of noncoding regions of the chloroplast genome in the Crassulaceae and related species. *Current Genetics* **25**, 558-566.
- Haslam, E. 1998. *Practical Polyphenolics: From Structure to Molecular Recognition and Physiological Action*, Cambridge University Press, New York, pg. 184.



- Hebert, P. D. N., Cywinska, A., Ball, S. L. & deWaard, J. R. 2003. Biological identifications through DNA barcodes. *Proceedings of the Royal Society of London Series B* **270**, 313-320.
- Hebert, P. D. N. & Hogg, I. D. 2004. Biological identification of springtails (Hexapoda: Collembola) from the Canadian Arctic, using mitochondrial DNA barcodes. *Canadian Journal of Zoology* **82**, 749-754.
- Henry, R. J. (eds) 2001. *Plant Genotyping: The DNA Fingerprint of Plants*. CABI Publishing, New York, pg. 242 and 243.
- Holderegger, R. & Abbott, R. J. 2003. Phylogeography of the Arctic-Alpine *Saxifraga oppositifolia* (Saxifragaceae) and some related taxa based on cpDNA and ITS sequence variation. *American Journal of Botany* **90**, 931-936.
- Ingvarsson P. K., Ribstein, S. & Taylor, D. R. 2003. Molecular evolution of insertions and deletion in the chloroplast genome of *Silene*. *Molecular Biology and Evolution* **20**, 1737-1740.
- Gross, G. G., Hemingway, R. W., Yoshida, T. & Branham, S. J. (eds) 1999. *Plant Polyphenols 2: Chemistry, Biology, Pharmacology, Ecology*. Kluwer Academic/ Plenum Publishers, New York, pg. 30.
- Grundmann, M., Schneiden, H., Russell, S. J. & Vogel, J. C. 2006. Phylogenetic relationships of the moss genus *Pleurochaete* Lindb. (Bryales: Pottiaceae) based on chloroplast and nuclear genomic markers. *Organisms, Diversity & Evolution* **6**, 33-45.
- Karp, G. 2003. *Cell and Molecular Biology: Concepts and Experiments*. 3<sup>rd</sup> Ed. John Wiley & Sons, Inc., USA, pg. 407 and 763.
- Kaur, J. & John, E. 2006. Beauty and the thief. *New Sunday Times*, 12 March, F1-F4.





- Kimura, M. 1980. A simple method for estimating evolutionary rates of base substitutions through comparable studies of nucleotide sequences. *Journal of Molecular Evolution* **16**, 111-120.
- Klug, W. S., Cummings, M. R. & Spencer, C. A. 2006. *Concepts of Genetics*. 8<sup>th</sup> Ed. Pearson Education, Inc., New Jersey, pg. 219-220.
- Kress, W. J., Wurdack, K. J., Zimmer, E. A., Weigt, L. A. & Janzen, D. H. 2005. Use of DNA barcodes to identify flowering plants. *Proceedings of the National Academy of Science USA* **102**, 8369-8374.
- Marshall, E. 2005. Will DNA bar codes breathe life into classification? *Science* **307**, 1037.
- Mast, A. R. & Givnish T. J. 2002. Historical biogeography and the origin of stomatal distributions in *Banksia* and *Dryandra* (Proteaceae) based on their cpDNA phylogeny. *American Journal of Botany* **89**, 1311-1323.
- Richlen, M. L. & Barber, P. H. 2005. A technique for the rapid extraction of microalgal DNA from single live and preserved cells. *Molecular Ecology Notes* **5**, 688-691.
- Rodrigues, K. F., Yeoh, K. A. & Kumar, S. V. 2006. Molecular genetic approaches for the conservation of endangered orchids. *Bulletin of the Genetics Society of Malaysia* **12**, 13-15.
- Rodrigues, K. F. & Kumar, S. V. 2006. Isolation of 43 microsatellite loci from *Paphiopedilum rothschildianum*, an endangered species of slipper orchid. *Orchids*, June, 448-451.
- Sager, R. & Ishida, M. R. 1963. Chloroplast DNA in *Chlamydomonas*. *Proceedings of the National Academy of Science USA* **50**, 725-730.





- Saitou, N. & Nei, M. 1987. The neighbor-joining method: a new method for reconstructing phylogenetic trees. *Molecular Biology and Evolution* **4**, 405-425.
- Sambrook, J. & Russell, D. W. 2001. *Molecular Cloning: A Laboratory Manual*. 3<sup>rd</sup> Ed. Cold Spring Harbor Laboratory Press, New York, pg. 1.32-1.34.
- Sang, T., Crawford, D. J. & Stuessy, T. F. 1997. Chloroplast DNA phylogeny, reticulate evolution, and biogeography of *Paeonia* (Paeoniaceae). *American Journal of Botany* **84**, 1120-1136.
- Sanger, F., Nicklen, S. & Coulson, A. R. 1977. DNA sequencing with chain-terminating inhibitors. *Proceedings of the National Academy of Science USA* **74**, 5463-5467.
- Scarafoni, A. & Duranti, M. 2001. An approach to critical assessment of the experimental conditions in practical molecular biology: isolation of plant DNA **29**, 21-23.
- Schindel, D. E. & Miller, S. E. 2005. DNA barcoding a useful tool for taxonomists. *Nature* **435**, 17.
- Schoser, G. 1981. Cultural suggestions for the *Paphiopedilum*: orchids for every admirer. *The Orchid Digest* **45**, 185-189.
- Shuman, S. 1994. Novel approach to molecular cloning and polynucleotide synthesis using *Vaccinia* DNA topoisomerase. *Journal of Biology Chemistry* **269**, 32678-32684.
- Small, R. L., Lickey, E. B., Shaw, J. & Hauk, W. D. 2005. Amplification of noncoding chloroplast DNA for phylogenetic studies in lycophytes and monilophytes with a comparative example of relative phylogenetic studies utility from Ophioglossaceae. *Molecular Phylogenetics and Evolution* **36**, 509-522.



- Soliva, M., Kocyan, A. & Widmer, A. 2001. Molecular phylogenetics of the sexually deceptive orchid genus *Ophrys* (Orchidaceae) based on nuclear and chloroplast DNA sequences. *Molecular Phylogenetics and Evolution* **20**, 78-88.
- Stappen, J. V., Weltjens, I., Munaut, F. & Volckaerta, G. 1999. Interspecific and progeny relationships in the genus *Stylosanthes* inferred from chloroplast DNA sequence variation. *Life Sciences* **322**, 481-490.
- Sugiura, M. 2003. History of chloroplast genomics. *Photosynthesis Research* **76**, 371-377.
- Taberlet, P., Gielly, L., Pautou, G. & Bouvet, J. 1991. Universal primers for amplification of three non-coding regions of chloroplast DNA. *Plant Molecular Biology* **17**, 1105-1109.
- Tsai, L. C., Yu, Y. C., Hsieh, H. M., Wang, J. C., Linacre, A. & James Lee, C. I., 2006. Species identification using sequences of the *trnL* intron and the *trnL-trnF* IGS of chloroplast genome among popular plants in Taiwan. *Forensic Science International* **164**, 193-200.
- Wakasugi, T., Sugita, M., Tsudzuki, T. & Sugiura, M. 1998. Updated gene map of tobacco chloroplast DNA. *Plant Molecular Biology Reporter* **16**, 231-241.
- Watson, J. D., Baker, T. A., Bell, S. P., Gann, A., Levine, M. & Losick, R. 2004. *Molecular Biology of the Gene*. 5<sup>th</sup> Ed. Pearson Education, Inc., Singapore, pg. 654-655.
- Will, K. W. & Rubinoff, D. 2004. Myth of the molecule: DNA barcodes for species cannot replace morphology for identification and classification. *Cladistics* **20**, 47-55.



Williams, J. F. 1989. Optimization strategies for the polymerase chain reaction. *Biotechniques* 7, 762–769.

Yamashiro, T., Fukuda, T., Yokoyama, J. & Maki, M. 2004. Molecular phylogeny of *Vincetoxicum* (Apocynaceae-Asclepiadoideae) based on the nucleotide sequences of cpDNA and nrDNA. *Molecular Phylogenetics and Evolution* 31, 689–700.

