

DEVELOPMENT OF A PROTOCOL FOR THE IMMOBILIZATION OF PYRODINIUM
BAHAMENSE VAR. COMPRESSUM FOR USE AS A BIOSENSOR FOR HARMFUL
ALGAL BLOOMS

JASON YEOH

THIS DISERTATION IS BEEN PRESENTED TO FULLFILL PART OF THE
REQUIREMENT TO OBTAIN BACHELOR OF SCIENCE WITH HONOURS

PERPUSTAKAAN
UNIVERSITI MALAYSIA SABAH

BIOTECHNOLOGY PROGRAMME
SCHOOL OF SCIENCE AND TECHNOLOGY
UNIVERSITY MALAYSIA SABAH

MARCH 2006



UMS
UNIVERSITI MALAYSIA SABAH

UNIVERSITI MALAYSIA SABAH

BORANG PENGESAHAN STATUS TESIS@

JUDUL: Development of protocol for the immobilization of pyroclinumBahan uji var. compressum for use as a biosensor for lambIJAZAH: Sarjana muda dgn kejuruanSAYA Jacob YeohSESI PENGAJIAN: 2003/2006

(HURUF BESAR)

Mengaku membenarkan tesis (LPSM/Sarjana/Doktor Falsafah) ini disimpan di Perpustakaan Universiti Malaysia Sabah dengan syarat-syarat kegunaan seperti berikut:-

1. Tesis adalah hakmilik Universiti Malaysia Sabah.
2. Perpustakaan Universiti Malaysia Sabah dibenarkan membuat salinan untuk tujuan pengajian sahaja.
3. Perpustakaan dibenarkan membuat salinan sebagai bahan pertukaran antara institusi pengajian tinggi.
4. Sila tandakan (/)

PERPUSTAKAAN
UNIVERSITI MALAYSIA SABAH☐

SULIT

(Mengandungi maklumat yang berdarjah keselamatan atau Kepentingan Malaysia seperti yang termaktub di dalam AKTA RAHSIA RASMI 1972)

☒

TERHAD

(Mengandungi maklumat TERHAD yang telah ditentukan oleh organisasi/badan di mana penyelidikan dijalankan)

☐

TIDAK TERHAD

Disahkan Oleh

(TANDATANGAN PENULIS)

(TANDATANGAN PUSTAKAWAN)

Alamat Tetap: 62 Taman Emas
Bt 9 1/2 jalan peninsular
93250 Kuching S.W.M.

Prof. Datuk Dr. Kanar Zaman
Nama Penyelia

Tarikh: 26/4/06

Tarikh: _____

CATATAN:- *Potong yang tidak berkenaan.

**Jika tesis ini SULIT atau TERHAD, sila lampirkan surat daripada pihak berkuasa / organisasi berkenaan dengan menyatakan sekali sebab dan tempoh tesis ini perlu dikelaskan sebagai SULIT dan TERHAD.

@Tesis dimaksudkan sebagai tesis bagi Ijazah Doktor Falsafah dan Sarjana secara penyelidikan atau disertai bagi pengajian secara kerja kursus dan Laporan Projek Sarjana Muda (LPSM).



DECLARATION

I affirm that this paper is my own work effort, except for the references and summaries, which have been, cited clearly its source.

PERPUSTAKAAN
UNIVERSITI MALAYSIA SABAH

22 March 2006



JASON YEOH
HS2003-2807

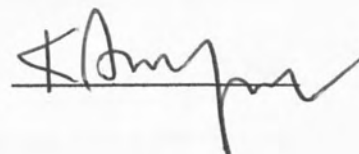


UMS
UNIVERSITI MALAYSIA SABAH

CONFIRMATION BY

Signature

Professor Datuk Dr. Kamaruzaman Ampon
(SUPERVISOR)

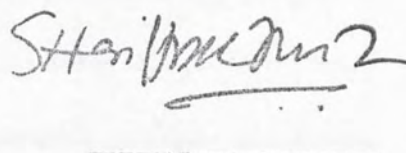


Dr. Ivy Wong Nyet Kui
(EXAMINER-1)



PERPUSTAKAAN
UNIVERSITI MALAYSIA SABAH

Assc. Professor Dr. Shariff A.K. Omang
(DEAN)



ACKNOWLEDGEMENT

First of all, I would like to give thanks to the Almighty Father for all His blessings and glory that I am able to finish this project. I would like to thank my project supervisor, Professor Datuk Dr. Kamaruzaman Ampon for giving me a chance to do my final year project under his supervision and also for giving me useful advises along the project period was conduct. Then I would like to thank Professor Datin Dr. Ann Anton for giving me the permission to use to the facilities in the Phycology lab and also for giving me some useful information regarding the HAB species that I dealing with for this project.

I also would like to thank the Dean of SST and all Biotechnology program's lecturers for understanding all the difficulties that I had while doing this project. Then I would like to give thanks to Mr. Chong Tong Seng for his valuable guidance and lessons that he taught to me. To Mr Ang Yee Kai, Miss Azima Azmi, Miss Tan Su Hui, Mr Lum Mok Sum, Mr. Musbah, and Puan Radizah, I also would like to thank you all for your corporation to me. To all of my fellow friends, thanks to you all for always supporting me during my ups and down as a Biotechnology student of University Malaysia Sabah.

And to my family, I would like to give million of thanks to you all for your undivided love and support to me especially to my beloved mother for always keep encouraging me to appreciate the beauty of knowledge and science. Last but not least I would like to thank every body that helped in my project in terms of moral support or thoughts. May God bless you all.

ABSTRACT

Harmful Algal Bloom (HAB) algae species that is found in Sabah, *Pyrodinium bahamense* var. *compressum* was immobilized and cultured in 4 types of different culture media in order to determine the growth rate after it had been immobilized. *Pyrodinium bahamense* var. *compressum* cells were immobilized through gel-entrapment technique by using sodium alginate (3% w/v). Beads with the diameter from 4 ± 0.33 mm until 5 ± 0.33 mm is formed with 10 ml of algae-alginate mixture were able to produce 100 beads. These beads were cultured in 4 types of media that were f/2, f/50, L1 10 dan L1 50 at the temperature of 25 ± 3 °C and 14:10 light:dark cycle with light intensity of 3000 lux for 21 days. Difference between growth rate of free algal cells culture and immobilized algal cells culture was observed through cell density per day value and OD reading at 680 nm wavelength per day value with the growth rate of immobilized algae cells was slower compared to the growth rate of free algal cells. Overall, cell density value is proportional to OD reading at 680 nm wavelength value for all culture media for immobilized algal cells culture with the growth rate of immobilize algal cells were optimum when cultured in f/2 media.

CONTENT

	Page
FRONT PAGE	i
DECLARATION	ii
CONFIRMATION	iii
ACKNOWLEDGEMENT	iv
ABSTRACT	v
ABSTRAK	vi
CONTENT	vii
LIST OF FIGURES	x
LIST OF PHOTOS	xiii
LIST OF ABBREVIATION	xiv
 CHAPTER 1 INTRODUCTION	 1
1.1 Introduction	1
1.2 Research Scope	3
1.3 Research Objectives	3
 CHAPTER 2 LITERATURE REVIEW	 4
2.1 Dinoflagellates	4
2.2 <i>Pyrodinium bahamense</i> var. <i>compressum</i>	6
2.3 Harmful algal bloom (HAB)	9
2.4 Paralytic Shellfish Poisoning (PSP)	14
2.5 Saxitoxin	15
2.6 Algal Cell Immobilization	18



CHAPTER 3	METHODOLOGY	21
3.1	Chemical Reagents	21
3.2	Apparatus/Glassware	22
3.3	Algae Source	23
3.4	Algae Culture Media Preparation	24
	3.4.1 f/2 and f/50 Media Preparation	24
	3.4.2 L1 10 and L150 Media Preparation	25
3.5	Culturing The Algae	27
3.6	Algae Immobilization	28
CHAPTER 4	RESULT	30
4.1	The Culture Condition	30
4.2	Size Of The Immobilize Beads	31
4.3	The Structure Of The Free Cells And The Immobilize Cells	32
4.4	Cell Density Of The Free Cells And The Immobilize Cells	34
4.5	Optical Density (OD) reading Of The Free Cells And The Immobilize Cells	39
4.6	The Relationship Between The Algae Cells Density With The OD reading	44
CHAPTER 5	DISCUSSION	49
5.1	The Immobilize Beads	49
5.2	The Cell Structure Of The Free Cell And The Immobilized Cells	51
5.3	The Cell's Growth Rate	52
5.4	Optical Density Analysis	56



CHAPTER 6	CONCLUSSION	58
REFERENCES		60
APPENDIX 1		64
APPENDIX 2		65
APPENDIX 3		66
APPENDIX 4		67
APPENDIX 5		68
APPENDIX 6		69
APPENDIX 7		70
APPENDIX 8		71



LIST OF FIGURES

Figure No.		Page
2.1	<i>Pyrodinium bahamense</i> var. <i>bahamense</i>	7
2.2	<i>Pyrodinium bahamense</i> var. <i>compressum</i>	8
2.3	Saxitoxin Chemical Structure	16
4.1	Growth curves of free <i>Pyrodinium bahamense</i> var. <i>compressum</i> in f/2 media	34
4.2	Growth curves of free <i>Pyrodinium bahamense</i> var. <i>compressum</i> in f/50 media	35
4.3	Growth curves of free <i>Pyrodinium bahamense</i> var. <i>compressum</i> in L1 10 media	35
4.4	Growth curves of free <i>Pyrodinium bahamense</i> var. <i>compressum</i> in L1 50 media	36
4.5	Growth curves of immobilized <i>Pyrodinium bahamense</i> var. <i>compressum</i> cells in f/2 media culture	36



Figure No.		Page
4.6	Growth curves of immobilized <i>Pyrodictinium bahamense</i> var. <i>compressum</i> cells in f/50 media culture	37
4.7	Growth curves of immobilized <i>Pyrodictinium bahamense</i> var. <i>compressum</i> cells in L1 10 media culture	37
4.8	Growth curves of immobilized <i>Pyrodictinium bahamense</i> var. <i>compressum</i> cells in L1 50 media culture	38
4.9	OD reading of free <i>Pyrodictinium bahamense</i> var. <i>compressum</i> cells in f/2 media culture	39
4.10	OD reading of free <i>Pyrodictinium bahamense</i> var. <i>compressum</i> cells in f/50 media culture	40
4.11	OD reading of free <i>Pyrodictinium bahamense</i> var. <i>compressum</i> cells in L1 10 media culture	40
4.12	OD reading of free <i>Pyrodictinium bahamense</i> var. <i>compressum</i> cells in L1 50 media culture	41
4.13	OD reading of immobilized <i>Pyrodictinium bahamense</i> var. <i>compressum</i> cells in f/2 media culture	41
4.14	OD reading of immobilized <i>Pyrodictinium bahamense</i> var. <i>compressum</i> cells in f/50 media culture	42
4.15	OD reading of immobilized <i>Pyrodictinium bahamense</i> var. <i>compressum</i> cells in L1 10 media culture	42
4.16	OD reading of immobilized <i>Pyrodictinium bahamense</i> var. <i>compressum</i> cells in L1 50 media culture	43



Figure No.		Page
4.17	Cell density of free <i>Pyrodinium bahamense</i> var. <i>compressum</i> cells against OD reading in f/2 media culture.	44
4.18	Cell density of free <i>Pyrodinium bahamense</i> var. <i>compressum</i> cells against OD reading in f/50 media culture.	45
4.19	Cell density of free <i>Pyrodinium bahamense</i> var. <i>compressum</i> cells against OD reading in L1 10 media culture.	45
4.20	Cell density of free <i>Pyrodinium bahamense</i> var. <i>compressum</i> cells against OD reading in L1 50 media culture.	46
4.21	Cell density of immobilized <i>Pyrodinium bahamense</i> var. <i>compressum</i> cells against OD reading in f/2 media culture.	46
4.22	Cell density of immobilized <i>Pyrodinium bahamense</i> var. <i>compressum</i> cells against OD reading in f/50 media culture.	47
4.23	Cell density of immobilized <i>Pyrodinium bahamense</i> var. <i>compressum</i> cells against OD reading in L1 10 media culture.	47
4.24	Cell density of immobilized <i>Pyrodinium bahamense</i> var. <i>compressum</i> cells against OD reading in L1 50 media culture.	48



LIST OF PHOTOS

Photo No.		Page
4.1	Cultures of free cells and immobilized cells in f/2 media	30
4.2	Size of the immobilize beads for f/2 media	31
4.3	Free <i>Pyrodictum bahamense</i> var. <i>compressum</i> chain cells	32
4.4	Free <i>Pyrodictum bahamense</i> var. <i>compressum</i> single cells	32
4.5	Immobilized <i>Pyrodictum bahamense</i> var. <i>compressum</i> single cells	33
4.6	Immobilized <i>Pyrodictum bahamense</i> var. <i>compressum</i> chain cells	33



LIST OF ABBREVIATION

HAB	Harmful Algal Bloom
PSP	Paralytic Shellfish Poisoning
ASP	Amnesic Shellfish Poisoning
NSP	Neurotoxic Shellfish Poisoning
DSP	Diarrhetic Shellfish Poisoning
STX	saxitoxin
Na ⁺	sodium ion
Ca ⁺	calcium ion
MU	mouse unit
MUm ^g ⁻¹	mouse units per milligram
HPLC	High Performance Liquid Chromatography
PSII	Photosystem II
APA	Alkaline phosphatase activity
Sub-ppb	sub parts per billion
BSA	Bovine serum albumin
rpm	rotation per minute
ml	millilitre
mgml ⁻¹	milligram per millilitre
gml ⁻¹	gram per millilitre
gL ⁻¹	gram per litre
N	Normality



$\text{FeCl}_3 \cdot 6\text{H}_2\text{O}$	ferum (II) chloride hexahydrate
$\text{Na}_2\text{EDTA} \cdot 2\text{H}_2\text{O}$	sodium edetate
$\text{NiSO}_4 \cdot 6\text{H}_2\text{O}$	Nickel (II) sulfate hexahydrate
Na_3VO_4	sodium orthovanadate
K_2CrO_4	potassium chromate



CHAPTER 1

INTRODUCTION

1.1) Introduction

Harmful Algal Bloom (HAB), commonly known as “Red tide” is a phenomenon where certain phytoplankton species contain reddish pigments, and “bloom” such that the water appears to be coloured red. This term red tide is no longer used because it is a misnomer. The blooms are not always red but can be yellow, orange, brown, pink or reddish brown. Besides that, blooms also are not dependent on the tides. Therefore, the term HAB is used to describe a bloom phenomenon that contains toxins or a bloom that cause negative impacts to the environment (Anderson, 2000) In Malaysia, harmful algal blooms were first reported in 1976 in the west coast of Sabah. It is caused by a toxic dinoflagellates known as *Pyrodinium bahamense* var. *compressum* which produce toxin known as saxitoxin which are well known for causing paralytic shellfish poisoning (PSP) (Lim, 2002). PSP is a group of heterocyclic guanidines. In marine context, it is known as product of dinoflagellates red tides that accumulate in shellfish to cause PSP when contaminated shellfish are consumed. Saxitoxin cause PSP by inhibiting nerve conduction

by blocking neuronal sodium channel thus causing oral facial numbness decreased arterial blood pressure, lost of voluntary muscle coordination and inhibition of respiration. By mass, saxitoxin is 1000 times more potent than cyanide and 50 times stronger than curare (Katırcıoğlu *et al.*, 2004)

Harmful algal bloom occurs throughout the world, and Scandinavian and Japanese fisheries are occasionally drastically affected, but in Malaysian the damage to industry is not major. However, it had give an unhealthy impact to fishing sectors in affected areas such as Sabah especially to the fisherman and the fish mongers as the public demands towards seafood are affected every time the bloom outbreak occur (Utusan Malaysia, 4 April 2000).

1.2 Research Scope

This project scope is to immobilize the toxic-producing algae based on the immobilization technique that is available for immobilizing cells.

1.3 Objective

The research objective is to immobilize *Pyrodictum bahamense* var. *compressum* growth based on the immobilization technique that been used for immobilizing cells such as cell entrapment. From this project, it is hope that we will be able to:

- To immobilize the toxic-producing algae in gel
- To compare the growth condition of the immobilized algae grown in two types of culture media, f/2 media and L1 media in various concentration.
- To determine the algae viability after immobilization.



CHAPTER 2

LITERATURE REVIEW

2.1 Dinoflagellates

The dinoflagellates were firstly defined by Otto Bütschli in 1885. At that time it was defined as the flagellate order Dinoflagellida. Botanists classified them under the division of Pyrrophyta and class of Dinophyceae. They are an important group of phytoplankton in marine and fresh water because of their roles in the aquatic food chain. Their adaptation to wide variety of environment is reflected by a tremendous diversity in form and nutrition and an extensive fossil record dating back few hundreds million years (*Dinoflagellate*, Wikipedia).

Some dinoflagellates produce toxins that are dangerous to man, marine mammals, fish, seabirds, and other components of the marine food light. Besides that, the dinoflagellates are large group of flagellate protests. Most of them are marine plankton but they are common in fresh habitats as well. About half of them are photosynthetic, thus making them the largest group of algae aside from diatoms (Hacket *et al.*, 2004).



Most dinoflagellates are unicellular forms with two dissimilar flagella. One of these extends towards the posterior, known as the longitudinal flagellum, while the other forms a lateral circle, called the transverse flagellum. The transverse flagellum provides most of the force propelling the cell and other imparts to it a distinctive whirling motion, which is what gives the dinoflagellates refers to. In Greek, *dinos* means whirling (*Dinoflagellate*, Wikipedia).

The dinoflagellates are photosynthetic organisms that are known to the capability of bioluminescence behaviour. It is widely accepted that the dinoflagellates account for much of the planktonic bioluminescence in the ocean (Kelly *et al.*, 1978). The physiology and the biochemistry of dinoflagellate are relatively well understood. The bioluminescence system of dinoflagellate consists of enzyme luciferase, its substrate luciferin, and a protein that binds luciferin. Nearly all luminescent organisms in the ocean emit light with a peak wavelength near 490 nm, and dinoflagellates are no exception (Hacket *et al.*, 2004).

Most dinoflagellates are haploid and reproduce primarily through fission, but sexual reproduction also occurs. This takes place by fusion of two individuals to form a zygote, which may remain mobile in typical dinoflagellate fashion or may form a resting cyst, which later undergoes meiosis to produce new haploid cells (*Dinoflagellate*, Wikipedia).

Dinoflagellates sometimes bloom in concentration of more than a million per cells per millilitre. Some species produce toxins that in such quantities will kill fish and accumulate in filter feeders such as shellfish, which in turn may pass them on people who consumed the affected fish or shellfish. This phenomenon is called a red tide or harmful algal bloom that will be discussed further.

2.2 *Pyrodinium bahamense* var. *compressum*

Pyrodinium bahamense var. *compressum* is one of two variety of *Pyrodinium bahamense*. Another variety of *Pyrodinium bahamense* is *Pyrodinium bahamense* var. *bahamense*. *Pyrodinium bahamense* in both varieties are bioluminescent. Except for the appearance, *Pyrodinium bahamense* var. *compressum* is longitudinally compressed and forms long chains of cells while *Pyrodinium bahamense* var. *bahamense* is round and found individually or in pairs (Refer Figure 2.1 and Figure 2.2)

Pyrodinium bahamense var. *compressum* can be found in the Indo-Pacific Ocean. In Malaysia, *Pyrodinium bahamense* var. *compressum* is found only the South China Sea in the coastal area of West Sabah, where it cause harmful algal bloom almost every year since it first outbreak reported in 1976 (Anton. *et al.*, 2000).

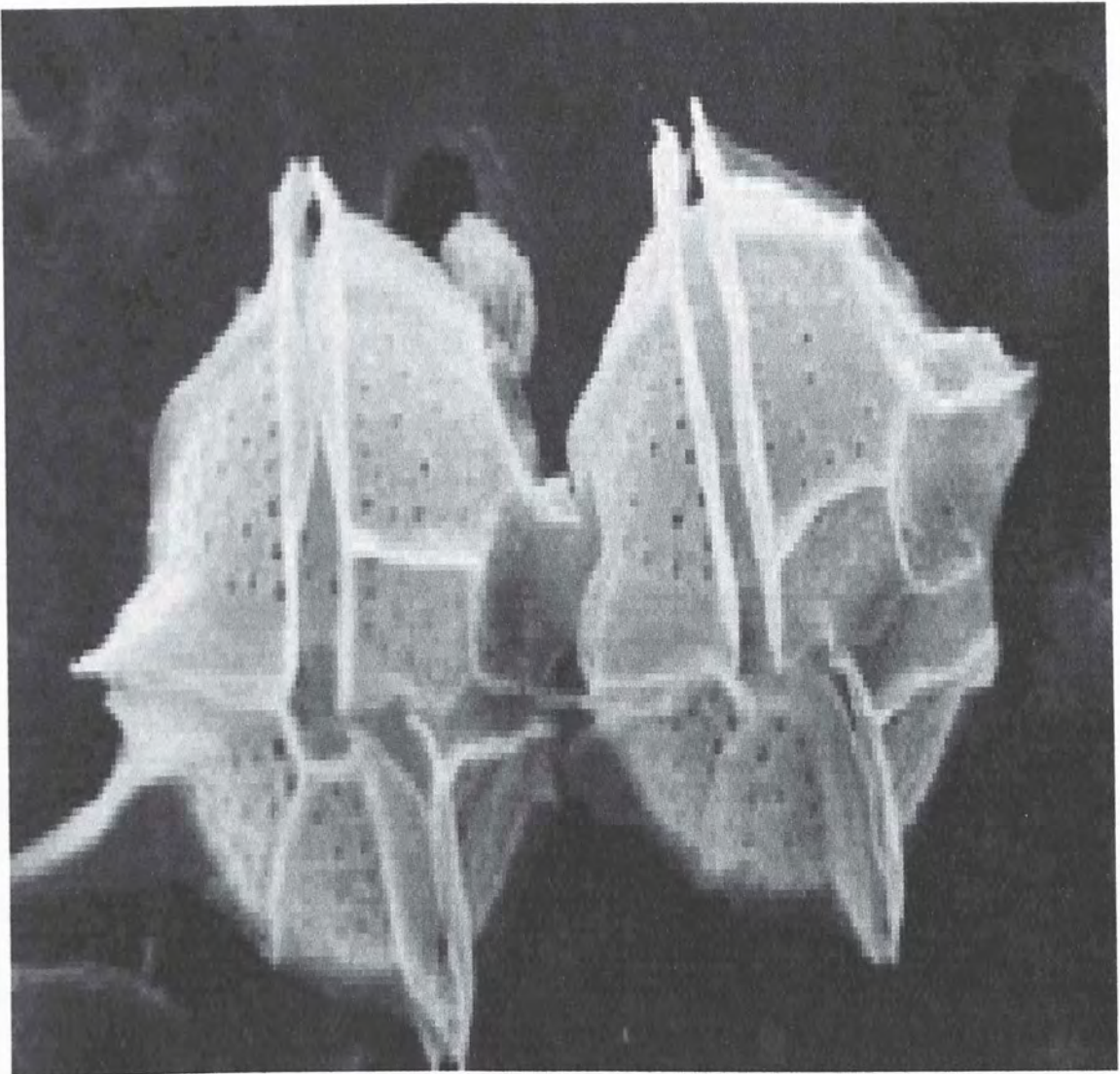


Figure 2.1 *Pyrodinium bahamense* var. *bahamense* (Phlips *et al.*, 2004)

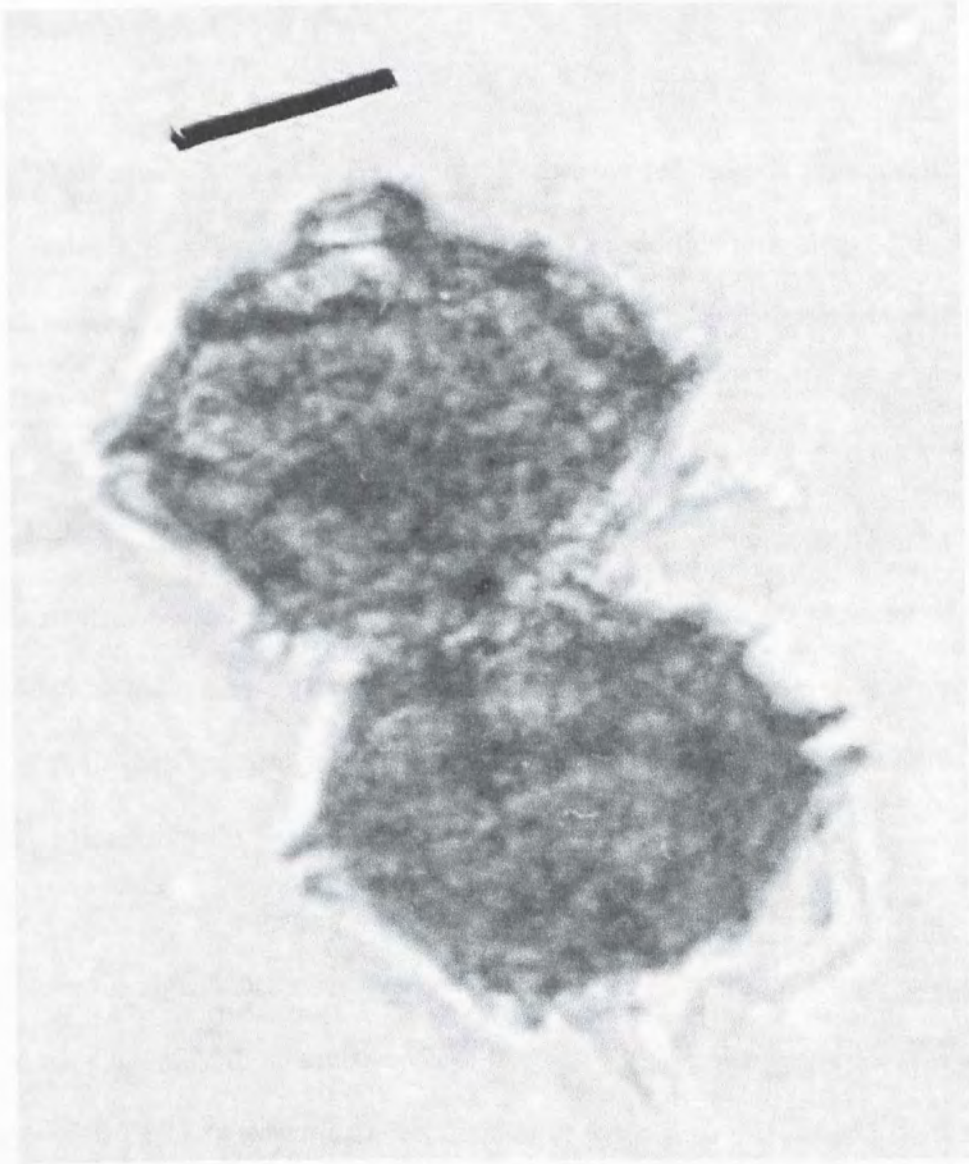


Figure 2.2 *Pyrodinium bahamense* var. *compressum* (UMS, 2005)

2.3 Harmful algal bloom (HAB)

Harmful algal bloom (HAB) is commonly known as red tide. It is associated with a massive multiplication or “bloom” of algae cells. The red tide term derived from species by which phytoplankton reproduce a high concentration of cell that made water visibly turns a red or dark brown colour (*Algal Bloom*, Wikipedia).

However, this term is not so correct to define the phenomena as red coloured water can also be caused by many non-toxic algae species. Besides that, most occurrences of toxic marine algae are not accompanied by any visible change in the colour of the water and also the occurrence of red tides has almost nothing to do with ocean tides (Baeir, 2000).

Harmful algal bloom are characterized by one important feature, which is they cause harm. Generally there are two ways that algal bloom can cause harm. Firstly, the algal bloom cause harm through the production of toxin such as paralytic symptom toxin (PST) or secondly, in non-toxic means that is physical bumping into organisms that cause damage, depletion of oxygen, and etc. Most algal blooms cycle are “investigate” by some limiting nutrient that is the growth of an algal bloom will be halted by the consumption of the available supply of the limiting nutrient. In freshwaters, this limiting nutrient tends to be phosphorous meanwhile in salt-water environment; the limiting nutrients that prevent the formation of an algal bloom are usually nitrogen and iron (Anderson *et al.*, 2002). It can rapidly increase to very high concentrations. Some blooms of this phytoplankton can

REFERENCES

- Anderson, D.M., Gilbert P.M., and Burkholder, J.M., 2002. Harmful algal blooms and eutrophication: nutrient sources, composition and consequences. *Estuaries*, **25(4b)**: 704-726
- Anderson, D., 2000. *The Harmful Algae Page*.
<http://www.redtide.whoi.edu/hab.htm>
- Anton, A, Alexander, J, Estim, A (2000). Harmful algal blooms in Malaysia: Revisiting Kimanis Bay. *Abstract Ninth International Conference on Harmful Algal Blooms, 7–11 February 2000, Hobart, Tasmania, Australia*, 78.
- Baeir C., 2000. Red Tide and Shellfish Poisoning: Toxic Products of Marine Algae, *Principal of Environmental Toxicology*. University of Idaho.
<http://www.agls.uidaho.edu/etoxweb/resources/Case%20Study/REDTIDE2.PDF>
- Blackburn, S.I., and Oshima, Y., 1989. Review of culture methods for *Pyrodinium bahamense*. In: Hallegraef, G.M and Maclean J.L (eds.) *Biology, Epidemiology, and Management of Pyrodinium Red Tides*. ICLARM, Manila, 257-272
- Bomber, J.W., Morton S.I., Babinchak J.A., Norris D.R. and Morton J.G., 1988. Epiphytic dinoflagellates of drift algae – another toxigenic community in the ciguatera food chain. *Bull. Mar. Sci.* **43**, 204-214
- Chouteau C., Dzyadevych S., Chovelon J.M., and Durrieu C., 2004. A bi-enzymatic whole cell conductometric biosensor for heavymetal ions and pesticides detection in water samples. *Articles in Press; Biosensors and Bioelectronics xxx (2004) xxx–xxx (not publish yet)*
- Chouteau C., Dzyadevych S., Chovelon J.M., and Durrieu C., 2004. Development of novel conductometric biosensors based on immobilized whole cell *Chlorella vulgaris* microalgae. *Biosensors and Bioelectronics* **19** (2004) 1089–1096.

- Chen Y.C., 2001. Immobilized microalga *Scenedesmus quadricauda* (Chlorophyta, Chlorococcales) for long-term storage and for application for water quality control in fish culture. *Aquaculture* **195**/1-2: 71-80.
- Curtis, K.M., Trainer, V.L., and Shumway, S.E., 2000. Paralytic shellfish toxins in geoduck clams (*Panope abrupta*): variability, anatomical distribution, and comparison of two toxin detection methods. *J. Shellfish Res.* **19**(1): 313-319.
- Glibert, P.M., and Pitcher, G. (eds), 2001. *Global Ecology and Oceanography of Harmful Algal Blooms, Science Plan*. Scientific Commission on Oceanic Research, Paris.
- Guillard R.R.L. and Ryther J.H., 1962. Studies of marine planktonic diatoms. I. *Cyclotella nana* Hustedt and *Detonula confervacea* Cleve. *Can. J. Microbiol.* **8**: 229-239
- Hackett J.D., Anderson D.M., Erdner D.L., and Bhattacharya D., 2004. Dinoflagellates: A Remarkable Evolutionary Experiment. *American Journal of Botany* **91**(10): 1523–1534.
- Hallegraeff, G.M., 1992. On the global spreading of toxic dinoflagellates. *SPC Ciguatera Information Bulletin* #2, July, 4-5
- Haque, S.M., and Onoue, Y., 2002. Effects of Salinity on Growth and Toxin Production of a Noxious Phytoflagellate, *Heterosigma akashiwo* (Raphidophyceae) *Botanica Marina* Vol. **45**, 356–363
- Jian Chin, 2005. *Bio - Hydrocarbons from Algae: impacts of temperature, light and salinity on algal growth*. Rural Industries Research and Development Corporation, Adelaide, 26 pgs
- Jeong, M.L., Gillis J.M., Hwang J.Y., 2003. Carbon Dioxide Mitigation by Microalgal Photosynthesis. *Bull. Korean Chem. Soc.* 2003, Vol. **24**, (12) , pp 1763-1766
- Kao, C. Y., 1993. Paralytic shellfish poisoning. In: Falconer, I. R. (ed) *Algal Toxins in Seafood and Drinking Water*, Academic Press, San Diego, 75-86.

- Katircioğlu H., Akın B. S., and Atıcı T., 2004. Microalgal toxin(s): characteristics and importance. *African Journal of Biotechnology* Vol. 3 (12), pp. 667-674.
- Kelly, M. G., and Tett P., 1978. Bioluminescence in the ocean. In: P. J. Herring [ed.], *Bioluminescence in action*, Academic Press, London, UK., 399-417.
- Laupa Junus, 2002. 'Air pasang merah' jadi baja. *Utusan Malaysia*, 14 September, 32
- Lim, P. T., Leaw, C. P. and Usup, G., 2002. First incidence of paralytic shellfish poisoning on the east coast of Peninsula Malaysia. *Proceeding Asia Pacific Conference on Marine Science and Technology*. Kuala Lumpur.
- Lehane, L., 2000. *Paralytic Shellfish Poisoning: A review*. National Office of Animal and Plant Health, Agriculture, Fisheries and Forestry, Canberra, 56 pgs.
- Martinez P.T., Gonzalez E.B., Lorenzo S.M., D. Rodriguez P. Micro-columns packed with *Chlorella vulgaris* immobilized on silica gel for mercury speciation. *Articles in Press; Talanta xxx (2005) xxx-xxx xxx (not publish yet)*
- Oshima, Y., 1995. Post column derivatization liquid chromatographic method for paralytic shellfish toxins. *Journal of AOAC International* 78 (2): 528-32.
- Phlips, E.J., Badylak, S., Youn, S., and Kelley K., 2004. The occurrence of potentially toxic dinoflagellates and diatoms in a subtropical lagoon, the Indian River Lagoon, Florida, USA. *Harmful Algae* 3 (2004) 39-49.
- RaLonde R., 1996. Paralytic Shellfish Poisoning: The Alaska Problem, Vol. 8(2). University of Alaska Sea Grant, Fairbanks, AK.
- Robinson P. K., Dainty A. L., Goulding K. H., Simpkins I., and Trevan M. D., 1985. Physiology of alginate-immobilized *Chlorella*. *Enzyme and Microbial Technology* Vol 7 (5):212-216
- Scholin, C.A., Gulland, F., Doucette, G.J., Benson, S., Busman, M., Trainer, V., and Dolah, F.M.V., 2000. Mortality of sea lions along the central California coast linked to a toxic diatom bloom. *Nature* 403:80-84.

Shuler M.L. and Kargi F., 2002. *Bioprocess Engineering: Basic Concepts 2nd Edition*. Prentice Hall PTR, United States of America, 535 pgs.

Taylor, F.J.R., and Fukuyo Y., 1989. Morphological features of the motile cell of *Pyrodinium bahamense*. In: Hallegraef, G.M and Maclean J.L (eds.) *Biology, Epidemiology, and Management of Pyrodinium Red Tides.*, ICLARM, Manila, 207-213

Taylor, F.J.R., Fukuyo Y., Larsen J., and Hallegraef, G.M., 2004. Morphological features of the motile cell of *Pyrodinium bahamense*. In: Hallegraef, G.M., Anderson D.M., and Cembella A.D., (eds.) *Manual On Harmful Marine Microalgae*. UNESCO Publishing, France, 207-213

Utusan Malaysia, 2000. UMS tubuh pasukan kaji peningkatan alga beracun. *Utusan Malaysia*, 4 April, 12.

University Malaysia Sabah, 2005. *Pyrodinium bahamense* var. *compressum*. <http://www.ums.edu.my/sst/redtide/Pyro.html>

Védrine C , Leclerc J.C., Durrieu C., and Minh C.T., 2003. Optical whole-cell biosensor using *Chlorella vulgaris* designed for monitoring herbicides. *Biosensors and Bioelectronics* **18** (2003) 457_463.

Wikipedia the free encyclopedia, 2005. *Algal Bloom*. http://en.wikipedia.org/wiki/Algal_bloom

Wikipedia the free encyclopedia, 2005. *Dinoflagellate*, <http://en.wikipedia.org/wiki/Dinoflagellate>

Willaert, R., and Baron, G., 1996. Gel entrapment and micro-encapsulation: methods, applications and engineering principles. *Reviews in Chemical Engineering* **12**: 5-205.

Wright J.L.C., 1995. Dealing with seafood toxins: present approaches and future options. *Food Research International*, Vol. **28**, No. 4, pp. 341-358.