

**THE DEVELOPMENT OF LIGHT ORGAN AND BACTERIAL INFECTION IN
SLIP-MOUTHS (LEIOGNATHIDAE)**

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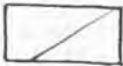
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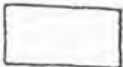
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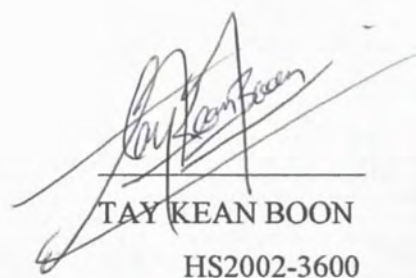
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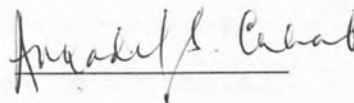


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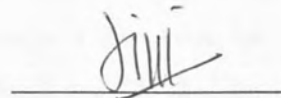
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ABSTRACT

To date, there are no previous studies to describe the development of the light organ in slip-mouths and at the stage at which bacterial infection occurs. Thirty four juveniles of *Leiognathus rapsoni*, a new record for Sabah, were captured by using modified triple layering gill net at Karambunai Lagoon (NE 06° 04' 39.5", E 116° 06' 59.7") for this investigation. Six juveniles of *Leiognathus splendens* were bought from roadside along Jalan Sulaman for culturing the bacterial presence in a leiognathid. Light organs from six of *L. rapsoni* (3.8 to 5.2 cm SL) were histologically examined. Results show that the light organ increases in height, width and thickness with increasing size of the leiognathids (3.8 to 5.2 cm SL). Furthermore, the pattern of development of the light organs might develop in two different ways. There are single sided growing pattern with a ring-liked surrounding, and dual sided growing pattern with a croissant-liked surrounding. All *L. splendens* juveniles (6.9 to 7.9 cm SL) have bacteria, *P. leiognathi*, that are gram negative. It is apparent from the results that the light organ in *L. rapsoni* develops and gets infected by *P. leiognathi* in individuals smaller than 3.8 cm. Since luminous bacteria can be bioindicators of the water quality, these findings indicate that the Karambunai Lagoon is not polluted by toxicants.



ABSTRAK

Sehingga kini, tiada kajian menghuraikan pembangunan organ bercahaya dalam slip-mouths dan daripada peringkat mana infeksi bakteria berlaku. Tiga puluh empat juvenil *Leiognathus rapsoni*, catatan sebuah rekod baru bagi Sabah, telah ditangkap dengan menggunakan sebuah pukut insang berlapisan tiga yang telah dimodifikasi di Teluk Karambunai (NE 06° 04' 39.5", E 116° 06' 59.7") untuk pemeriksaan. Enam juvenil *Leiognathus splendens* telah dibeli daripada tepi jalan sepanjang Jalan Sulaman untuk penentuan kehadiran kultur bakteria dalam leiognathid. Organ bercahaya daripada enam individu *L. rapsoni* (3.8 hingga 5.2 cm SL) telah dijalani pemeriksaan histologi. Keputusan menunjukkan bahawa organ bercahaya bertambah dalam ketinggian, lebar dan ketebalan dengan pertambahan saiz leiognathid (3.8 hingga 5.2 cm SL). Tambahan ini, corak perkembangan organ bercahaya mungkin berkembang dalam dua jenis yang berlainan, iaitu corak pertumbuhan sisian-bujang dengan lingkaran bentuk cincin, serta corak pertumbuhan sisian-dedua dengan lingkaran bentuk roti berbentuk anak bulan. Semua juvenil *L. splendens* (6.9 to 7.9 cm SL) mempunyai bakteria *P. leiognathi* yang bergram negatif. Daripada keputusan yang diperolehi, adalah jelas bahawa organ bercahaya dalam *L. rapsoni* berkembang dan dapat infeksi oleh *P. leiognathi* dalam individu yang lebih kecil daripada 3.8 cm. Oleh disebabkan bakteria yang bersinar boleh menjadi sebuah penunjuk biologi bagi kualiti air, maka kajian ini telah menunjukkan bahawa Teluk Karambunai adalah bebas daripada pencemaran toksik.

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4.19 *Photobacterium leiognathi*

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LIST OF SYMBOLS

%	percentage
°C	degree of celcius
g	gram
ml	mililitre
m	metre
cm	centimetre
mm	milimetre
µm	micrometer
s	second(s)
min	minute(s)
h	hour(s)
kPa	kilo Pascal
kCal	kilo Calories
<	less than
>	more than



LIST OF ABBREVIATIONS

Approx.	approximately
SL	standard length
SD	standard deviation



CHAPTER 1

INTRODUCTION

1.1 BACKGROUND

Natural bacteria in the marine environment are important to the ecology of fishes but this natural population may be disturbed by pollution. Slipmouths (Leiognathidae), which often known as ponyfish are bioluminescent fishes, having a culture of bioluminescent fishes in a light organ around the esophagus. The bacteria is from the natural, free-living *Photobacterium leiognathi* (Nealson and Hastings, 1979). No study has been done on the development of the light-organ and culture of *Photobacterium leiognathi*.

Leiognathidae is mainly habitat marine though some species enter freshwater. It occurs in estuarine and coastal waters of the Indo-West Pacific (Pauly, 1976). It feeds on benthic invertebrates, therefore, it is commonly found in shallow coastal waters and tidal creeks. It can be easily caught by gill net during adult, nevertheless, obviously not so easy juvenile. Thereafter, Karambunai Lagoon, Sabah is to be chosen to set as a spot (divided into several stations-close or mid of sea grass region) to collect specimens.



Karambunai Lagoon suits the best to all required factors for habitat and living environment for Leionathidae.

1.1.1 The Development of Light Organ of Leionathids

Of the more than 30 families of bioluminescent marine fishes, a dozen or more rely on luminous bacteria for light production (Nealson and Hastings, 1979). Among these bacterially bioluminescent fishes, leionathids is found sufficiently abundant and accessible that extensive behavioral studies of their luminescence are feasible. Bioluminescent in leionathids is produced by an apparently pure culture of *Photobacterium leionathi* (Hastings and Mitchell, 1971) housed in a circumesophageal, gland-like light organ located just anterior to the stomach. This internal position of the light organ allows for the incorporation of other parts of the fishes' anatomy in the expression and control of luminescence. Proximal control of light expression is by the operation of muscular shutters on the light organ itself. Therefore, to study the slowly development of the light organ, histology method is in use.

1.1.2 The Development of Bacterial Infection in Leionathids

Symbiotic luminescence is known in many species of fishes in shallow marine environments. These unique bacteria-animal interactions are unusual in that the hosts harbor monospecific cultures of one species of luminous bacteria at high density as their light source in specialized organs. To maintain this specific interaction between the hosts

and their symbiotic bacteria over ensuing generations, the symbiotic must be passed on to the offspring with fidelity (Haneda and Tsuji, 1976). In this project, the colonization and establishment of the bacterial population within the light organ are to be directly assessed by Gram staining from the larval to juvenile leiognathids.

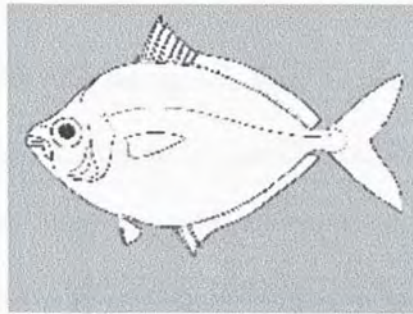


Figure 1.1 Draft of ponyfish (Shiino, 1976)

Table 1.1 Taxonomic Hierarchy

Family	Leiognathidae	-- ponyfishes
Genus	<u>Gazza</u>	
Genus	<u>Leiognathus</u>	
Genus	<u>Secutor</u>	

(Shiino, 1976)

1.2 OBJECTIVES

There are mainly two objectives to this project: firstly, to describe the development of light organ in slipmouths (*Leiognathidae*); and secondly, to determine when bacterial (*Photobacterium leiognathi*) infection occurs.

By having these two objectives succeeded, there is one method for field work, and two methods of laboratory work have to be carried out. In field work, specimens have to be captured by sampling. Sampling consist three basic steps, briefly, vary net capturing, data labeling and formalin preserving. Whereas, in laboratory work, methods of histology (Samoilys and Roelofs, 2000) and Gram staining (Standard procedure introduced by Brock, 2000) are to be chosen to describe and determine the objectives. These two methods are closely guided and followed by standard procedures.



1.3 HYPOTHESIS

There are two sets of hypotheses: hypothesis null and two hypothesis alternatives. Whereby, first hypothesis is that the light organ develops slowly in the out-pocketing of the esophagus, with the hypothesis alternative of the light organ does not develop slowly in the out-pocketing of the esophagus. Secondly, the hypothesis null is that the bacterial infection occurs at 5mm long in leiognathidae, with the hypothesis alternative of the bacterial infection does not occur at 5mm long in Leiognathidae.



REFERENCES

- Blaber, S. J. M., Brewer, D. T. and Salini, J. P., 1995. Fish communities and the nursery role of a tropical bay in the Gulf of Carpentaria, Australia. *Estuarine, Coastal and Shelf Science* **40**, 177-188.
- Brock, K., 2000. Gram Stain. *Microbiology 120L in General Bacteriology Laboratory Manuals*. 46-50.
- Boettcher, K. J. and Ruby, E. G., 1990. Depressed light emission by symbiotic *Vibrio fischeri* of the sepiolid squid *Euprymna scolopes*. *Journal of Bacteriol* **172**, 3701-3706.
- Boisvert, H., Chatelain, R. and Bassot, J. M., 1967. Luminous bacteria in Leionathidae. *Annles Institute of Pasteur* **112**, 520-524.
- Buck, J., 1988. Synchronous rhythmic flashing of fireflies. *Quarterly Review of Biology* **63**, 265-289.
- Chan E. H. and Liew H. C., 1986. Characteristics of an exploited tropical shallow-water demersal fish community in Malaysia. In: Maclean, J.L., Dizon, L B. and Hosillos, L.V.(eds) *Proceedings of the First Asian Fisheries Forum*. Asian Fisheries Society, 349 – 352.
- Fitzgerald, J. M., 1977. Classification of luminous bacteria from the light organ of the Australian pinecone fish, *Cleidopus gloriamaris*. *Archeological Microbiology* **112**, 153-156.



- Haddock, S. H. D. and Case, J. F., 1999. Bioluminescence spectra of shallow and deep-sea gelatinous zooplankton: ctenophores, medusae, and siphonophores. *Marine Biology* **133**, 571-582.
- Haddock, S. H. D., Rivers, T. J. and Robison, B. H., 2003. Can coelenterates make coelenterazine? Dietary requirement for luciferin in cnidarian bioluminescence. *Proceeding of National Academia Science* **98**, 11148-11151.
- Haddock, S. H. D. and Ling, H. Y., 1997. The enclosing latticed sphere of *Tuscaridium cygneum* (Murray), a eurybathyal phaeodarian Radiolaria, from the North Pacific. *Paleontological Research* **1**, 144-149.
- Haddock, S. H. D., McDougall, C. M. and Case, J. F., 2004. "The Bioluminescence Web Page", <http://lifesci.ucsb.edu/~biolum/>
- Hajisamae, S. and Chou, L. M., 2002. Do shallow water habitats of an impacted coastal strait serve as nursery grounds for fish? *Estuarine, Coastal and Shelf Science* **56**, 281-290.
- Hall, M. A., 1996. On bycatches. *Reviews in Fish Biology and Fisheries* **6**, 319-352.
- Haneda, Y., 1940. On the luminescence of the fishes belonging to the family Leiognathidae of the tropical Pacific. *Palao trop. Boil. Stn Stud.* **2**, 29-39.
- Haneda, Y., 1950. Luminous organ of fish which emit light indirectly. *Pacific Science* **4**, 214-277.
- Haneda, Y., 1965. Observations on a luminous apogonid fish, *Siphamia versicolor* and on others of the same genus. *Sci Rep. Yokosuka City Mus* **11**, 1-12.



- Haneda, Y. and Tsuji, F. I., 1976. The luminescent systems of ponyfishes. *Journal of Morphology* **150**, 539-552.
- Harm, J. W., 1928. Bau und Entwicklung eines eigenartigen Leuchtorgans bei *Equula spec.* *Zeitschrift für Wissenschaftliche Zoologies* **131**, 157-179.
- Harris, S. A., Cyrus, D. and Beckley, L. E., 1999. The larval fish assemblages in the nearshore waters off the St. Lucia Estuary, South Africa. *Estuarine, Coastal and Shelf Science* **49**, 789-811.
- Harris, S. A., Cyrus, D. and Beckley, L. E., 2001. Horizontal trends in larval fish diversity and abundance along an ocean-estuarine gradient on the northern KwaZulu- Natal coast, South Africa. *Estuarine, Coastal and Shelf Science* **53**, 221-235.
- Hastings, J. W. and Mitchell, G., 1971. Endosymbiotic bioluminescent bacteria from the light organ of pony fish. *Biology Bulletin Marine* **141**, 261-268.
- Hastings, J. W., 1996. Chemistries and colors of bioluminescent reactions-a review. *Gene* **173**, 5-11.
- Hastings, J. W., 1998. Bioluminescence. In: Sperelakis, N. (ed) *Cell Physiology*, Academic Press, New York, 984-1000.
- Herring, P. J., 1987. Systematic distribution of bioluminescence in living organisms. *Journal of Bioluminescence and Chemiluminescence* **1**, 147-163.
- Herring and Morin, J.G., *Bioluminescence in action*. Academic Press, New York.



- Herring, P. J. and Morin, J. P., 1978. Bioluminescence in fishes. In: Herring, P. J. (ed) *Bioluminescence in action*, London, 273-330.
- Herring, P. J., and J. G. Morin., 1978. *Bioluminescence in fishes*. Academic Press, New York.
- Herring, P. J., Clarke M. R., Boletzky, S. V. and Ryan, K. P., 1981. The light organs of *Sepiolo atlantica* and *Spirula spirula* (Mollusca: Cephalopoda): bacterial and intrinsic systems in the order Sepioidea. *Journal of Marine Biological Association of United Kingdom* **61**, 901-916.
- Herring, P. J., 1977. Bioluminescence in marine organisms. *Nature* **267**, 788-793.
- Herring, P. J., 1978. *Bioluminescence in action*. Academic Press, London.
- Herring, P. J. and Widder, E. A., 2001. Plankton: Light production in marine organisms, mechanisms and functions. Academic Press, London.
- Ikejima, K., Tongnunui, P., Medej, T. and Taniuchi, T., 2002. Juvenile and small fishes in a mangrove estuary in Trang Province, Thailand: seasonal and habitat differences. *Estuarine, Coastal and Shelf Science* **56**, 447-457.
- Jayabalan, N., 1980. *Studies on silver-bellies (Pisces: Leiognathidae) and their associated bioluminescent bacteria of Porto Novo waters*. Ph.D. thesis. Annamalai University, India.
- Jayawardane, P. A. A. T. and Dayaratne, P., 1998. Reproductive Biology of Shortnase Ponyfish *Leiognathus brevirostris* (Valenciennes) from Portugal Bay in the Puttalam Estuary, Sri Lanka. *Asian Fisheries Science* **10**, 189-200.



- Jones, G., 1985. Revision of the Australian species of the fish family Leiognathidae. *Australian Journal of Marine and Freshwater Research* **36**, 559-613.
- Kühlmorgan-Hille, G., 1974. *Leiognathidae. FAO species identification sheets for fishery purposes, Eastern Indian Ocean (Fishing Area 57) and Western Central Pacific (Fishing Area 71)*. Food and Agriculture Organization of the United Nations.
- Kuiter, R. H. 1996. *Photo Guide to Fishes of the Maldives*. Atoll Editions, Apollo Bay, Australia.
- McFall-Ngai, M. J., 1981. New function for the teleost gasbladder: a source of oxygen for the bioluminescent bacteria in the leiognathid light organ. In: Obispo, S.L. *Western Society of Naturalists*, California.
- McFall-Ngai, M. J., 1998. The development of cooperative associations between animals and bacteria: establishing détente between domains. *Am. Zoology* **38**, 3-18.
- McFall-Ngai, M. J. and Dunlap, P. V., 1983. Three New Modes of Luminescence in the Leiognathid Fish *Gazza minuta*: Discrete Projected Luminescence, Ventral Body Flash, and Buccal Luminescence. *Marine Biology* **73**, 227-237.
- Mc-Fall-Ngai, M. J. and Dunlap, P. V., 1984. External and internal sexual dimorphism in leiognathid fishes: morphological evidence for sex-specific bioluminescent signaling. *Journal of Morphological* **182**, 71-83.
- Michael, F. C. and Paul, V. D., 2000. Aposymbiotic culture of the sepiolid squid *Euprymna scolopes*: Role of the symbiotic bacterium *Vibrio fischeri* in host animal growth, development, and light organ morphogenesis. *Journal of Experimental Zoology* **286**, 280-296.



- Montgomery, M. K., and McFall-Ngai, M. J., 1995. The inductive role of bacterial symbionts in the morphogenesis of a squid light organ. *Am. Zoology* **35**, 372-380.
- Morin, J. G., 1982. Bioluminescent patterns in shallow tropical marine fishes. In: Gomez, E. *The Reef and Man, Proceedings of the Fourth International Coral Reef Symposium, Manila* **2**, 569-574.
- Masachika Maeda, 1997. Introduction of detrital food webs into an aquaculture system by supplying single cell algal detritus produced from *Laminaria japonica* as a hatchery diet for *Anemia nauplii*. *Journal of Aquaculture*. **154**. 125-137.
- Nealson, K. H. and Hastings, J. W., 1991. *The luminous bacteria*.
- Nelson, K. H. and Hastings, J. W., 1979. Bacterial bioluminescence: its control and ecological significance. *Microbiology Review* **43**. 496-518.
- Pauly, D., 1976. The Leiognathidae (teleosts): their stock and fishery in Indonesia, with notes on the biology of *Leiognathus splendens*. In: *Ecology and management of some tropical shallow eater communities* **17**.
- Pfister, C. A., 1999. Recruitment of intertidal fishes. In: Horn, M.H., Martin, K.L.M. and Chotkowski, M.A. (eds), *Intertidal fishes life in two worlds*, Academic Press, London.
- Reichelt J. L. and Baumann, P., 1975. *Photobacterium mandapamensis* Hendrie et al. a later subjective synonym of *Photobacterium leiognathi* Boisvert et al. *Int J Syst Bacteriol* **25**, 208-209



- Ramaiah, N. and Chandramohan, D., 2003. Ecological and Laboratory studies on the role of luminous bacteria and their luminescence in coastal pollution surveillance. *Marine Pollution Bulletin* **26**, 190-201.
- Reichelt, J. L., Neelson, K. H. and Hastings, J. W., 1977. The specificity of symbiosis: pony fish and luminescent bacteria. *Arch. Microbiology* **112**, 157-161.
- Ruby, E. G. 1996. Bacterial-animal association: the *Vibrio fischeri*-*Euprymna scolopes* light organ symbiosis. *Annual Review Microbiology* **50**, 591-624.
- Ruby, E. G. and Neelson, K. H., 1977. Pyruvate production and excretion by the luminous marine bacteria. *Application Environmental Microbiology* **34**, 164-169.
- Ruby, E. G., and Lee, K. H., 1998. The *Vibrio fischeri* *Euprymna scolopes* light organ association: current ecological paradigms. *Application Environmental Microbiology* **64**, 805-812.
- Ruby, E. G., and Asato, L. M., 1993. Growth and flagellation of *Vibrio fischeri* during initiation of the sepiolid squid light organ symbiosis. *Arch. Microbiol.* **159**, 160-167.
- Samoilys, M. A. and Roelofs, A., 2000. Defining the reproductive biology of a large Serranid *Plectropomus leopardus*. *CRC Reef Research Technical Manual* **30**, 4-8.
- Santamaria, C. A., Mateo, M. M. D., Traveset, R., Sala, R., Grau, A., Pastor, E., Sarasquete, C. and Crespo, S., 2004. Larval organogenesis in common dentex *Dentex dentex* L. (Sparidae): histological and histochemical aspects. *Aquaculture*, **237**, 207-228.
- Shiino, S. M., 1976. List of common names of fishes of the world, those prevailing among English-speaking nations. *Science Report of Shiima Marineland* **4**, 262.



- Smoyer, W. E., Ransom, R., Harris, R. C., Welsh, M. J., Lutsch, G. and Benndorf, R., 1999. Ischemic acute renal failure induces differential expression of small heat shock proteins. *J. Am. Soc. Nephrol* **11**, 211-221
- Spencer V. and Nyholm, 2000. Establishment of an animal-bacterial association: Recruiting symbiotic vibrios from the environment. *Proceedings of the National Academy of Sciences of United States of America* **97**, 10231-10235
- Viviani, V. R., Hastings, J. W. and Wilson, T., 2002. Two bioluminescent Diptera: the North American *Orfelia fultoni* and the Australian *Arachnocampa flava*. Similar niche, different bioluminescence systems. *Photochemical Photobiology* **75**, 22-27.
- Wada, M., Azuma, N., Mizuno N. and Kurokura H., 1999. Transfer of symbiotic luminous bacteria from parental *Leiognathus nuchalis* to their offspring. *Journal of Marine Biology* **135**, 683-687.
- Wei, S. L. and Young, R. E., 1989. Development of symbiotic bacterial bioluminescence in a nearshore cephalopod, *Euprymna scolopes*. *Marine Biology* **103**, 541-546.
- West, G., 1990. Methods of assessing ovarian development in fishes: a review. *Australian Journal of Marine and Freshwater Research* **41**, 199-222.
- Widder, E. A., 1997. Bioluminescence-Shedding some light on plankton distribution patterns. *Sea Technology*, 33-39.
- Widder, E. A., 1998. *The Bioluminescence Coloring Book*. Harbor Branch Oceanographic Institution, Florida.



- Widder, E. A., 1999 Bioluminescence. In: Archer, S.N. (eds) *Adaptive mechanisms in the ecology of vision*, 555-581.
- Widder, E. A., in press. Bioluminescence and the pelagic visual environment. *Marine and Freshwater Behaviour and Physiology*.
- Widder, E. A., Johnsen, S., Bernstein, S. A., Case, J. F. and Neilson, D. J., 1999 Thin layers of bioluminescent copepods found at density discontinuities in the water column. *Marine Biology* **134**, 429-437.
- Wilson, T. and Hastings, J. W., 1998. Bioluminescence Annual Revision of Cell Developmental Biology **14**, 197-230.
- Winsor, L., 1991. Histological techniques for fish gonad studies. Coral trout and red-throat emperor age, growth and reproduction workshop. *Australian Institute of Marine Science*.
- Winsor, L., 1994. Tissue processing. In: Woods, A. E. and Ellis, R. C. (eds) *Laboratory histopathology: a complete reference* **4**.
- Woodland, D. J., Cabanban, A. S., Taylor, V. M. and Taylor, R. J., 2002. A synchronized rhythmic flashing light display by schooling *Leiognathus splendens* (Leiognathidae : Perciformes). *Journal of Marine and Freshwater Research* **53**, 159-162.
- Woodland, D. J., Premcharoen, S. and Cabanban, A. S., 2001. Family Leiognathidae. In: Carpenter, K. and Niem, V. *FAO Identification Guide to the Living Marine Resources of the Western Central Pacific* **5**, 2814-2815.

