CONVERSION OF RAPD MARKERS TO CO-DOMINANT BASED SEQUENCE CHARACTERIZED AMPLIFIED REGION (SCAR) MARKERS IN THREE *Paphiopedilum* SPECIES

THUN EVONNE

PERPUSTAKAAN UNIVERSITI MALAYSIA SAPAT

THIS THESIS IS SUBMITTED TO FULFILL THE REQUIREMENT TO OBTAIN A BACHELOR OF SCIENCE DEGREE WITH HONOURS

PLANT TECHNOLOGY PROGRAM SCHOOL OF SCIENCE AND TECHNOLOGY UNIVERSITI MALAYSIA SABAH

MARCH 2007



	100.00			1000		
10	17.1	5.1	ß	· CH	CT-	
- 5-	1.1	C (1.7	- 54	91	

UNIVERSITI MALAYSIA SABAH

	BORANG P	PENGESAHAN STATUS TESIS@	
JUDUL: Conversion	OF RAPD M	larkers to Co-dominant-based Sequence	
Charade rize	ed Amplified	Region (SCAR) markers in Three Paphiopedilum	Speares
Tjazah: BSc. CHON	() Plant T	echnology	
	SESI PI	ENGAJIAN: 2006/07	
Saya THUN EVONN	E		
		(HURUF BESAR)	
Malaysia Sabah dengan		na/Doktor Falsafah)* ini disimpan di Perpustakaan Universiti gunaan seperti berikut:	
1. Tesis adalah hakmili	k Universiti Mal	avsia Sahah	
		pah dibenarkan membuat salinan untuk tujuan pengajian sahaja.	
which is the second sec	rkan membuat sa	linan tesis ini sebagai bahan pertukaran antara institusi pengajian PERPUSTAKAAN	
tinggi. 4. **Sila tandakan ()	1	UNIVERSITI MALAYSIA SABAH	
Sha tandakan ((Mengandungi maklumat yang berdarjah keselamatan atau	
St	ЛIТ	kepentingan Malaysia seperti yang termaktub di dalam AKTA RAHSIA RASMI 1972)	
		AKTA KADIA KASIMI (972)	
TE TE	ERHAD	(Mengandungi maklumat TERHAD yang telah ditentukan oleh organisasi/badan di mana penyelidikan dijalankan)	
		(Mengandungi maklumat TERHAD yang telah ditentukan	
	ERHAD DAK TERHAD	(Mengandungi maklumat TERHAD yang telah ditentukan	
		(Mengandungi maklumat TERHAD yang telah ditentukan oleh organisasi/badan di mana penyelidikan dijalankan)	
π	DAK TERHAD	(Mengandungi maklumat TERHAD yang telah ditentukan oleh organisasi/badan di mana penyelidikan dijalankan)	
(TANDATANGAN)	DAK TERHAD PENULIS)	(Mengandungi maklumat TERHAD yang telah ditentukan oleh organisasi/badan di mana penyelidikan dijalankan) Disahkan oleh	
(TANDATANGAN)	DAK TERHAD PENULIS) n Empat	(Mengandungi maklumat TERHAD yang telah ditentukan oleh organisasi/badan di mana penyelidikan dijalankan) Disahkan oleh (TANDATANGAN PUSTAKAWAN) DR VIJAY KUMAR	
(TANDATANGAN) (TANDATANGAN) Alamat Tetap: 88, Jala Aorpang Baru, 31	DAK TERHAD PENULIS) n Empat	(Mengandungi maklumat TERHAD yang telah ditentukan oleh organisasi/badan di mana penyelidikan dijalankan) Disahkan oleh (TANDATAN POSTAKAWAN)	
(TANDATANGAN) (TANDATANGAN) Alamat Tetap: 88, Jala	DAK TERHAD PENULIS) n Empat	(Mengandungi maklumat TERHAD yang telah ditentukan oleh organisasi/badan di mana penyelidikan dijalankan) Disahkan oleh (TANDATANGAN PUSTAKAWAN) DR VIJAY KUMAR	

** Jika tesis ini SULIT atau TERHAD, sila lampirkan surat daripada pihak berkuasa/organisasi berkenaan dengan menyatakan sekali sebab dan tempoh tesis ini perlu dikelaskan sebagai SULIT dan TERHAD.

@ Tesis dimaksudkan sebagai tesis bagi Ijazah Doktor Falsafah dan Sarjana secara penyelidikan, atau disertasi bagi pengajian secara kerja kursus dan penyelidikan, atau Laporan Projek Sarjana Muda (LPSM).



DECLARATION

I hereby declare that this thesis is fully my own work except the quotation that I have clearly stated the sources.

12 MARCH 2007

PERPUSTAKAAN UNIVERSITI MALAYSIA SABAH

was.

THUN EVONNE HS 2004-2366



CERTIFIED BY

1. SUPERVISOR

Signature

(Dr. Vijay Kumar)

2. EXAMINER

(Ms. Chee Fong Tyng)

PERPUSTAKAAN UNIVERSITI MALAYSIA SABAH

3. DEAN

SHON

(SUPT/KS Prof. Madya Dr. Shariff A. K. Omang)



ACKNOWLEDGEMENT

Firstly, I would like to give my deepest gratitude to Dr. Vijay Kumar for giving me a chance to work on this interesting project and providing his guidance throughout the development of this project.

Next, my sincere gratitude goes to my seniors, Kenneth Rodrigues, Melvin Kinsuat and Thien Yong Nam for their support, guidance and valuable opinions throughout this project.

Besides, I would like to thank Yeoh Keat Ai and Lee Ting Ting who are doing similar projects on their assistance and discussion. Not forgetting are all the seniors, staffs and lab mates in BRI lab for their guidance and assistance during the whole progress of this project.

Last but not least, I would like to thank my family for their love, trust and neverending support to me.

Without the help and support from all of you, I would not be able to finish this project. May God bless you all.



ABSTRACT

The genus Paphiopedilum is endangered species protected under Convention on International Trade in Endangered Species (CITES). However, the discrimination between the species is difficult due to their similar morphological characters. Therefore, RAPD assay were applied to detect polymorphisms between the three species: P. rothschildianum, P. dayanum and P. lowii. Diagnostic fragments identified for discrimination for each species were converted into SCAR markers by cloning and sequencing the diagnostic fragments and designing SCAR primer pairs to amplify the diagnostic fragments. The reproducibility of these markers was analysed by performing diagnostic PCRs. In this study, PCR amplification with RAPD primers was successful in detecting polymorphisms between species. Three diagnostic fragments were converted into SCAR markers and all primer pairs amplified single distinct bands whose sizes were the same as expected fragments. One of the SCAR primers amplified the expected product size in P. dayanum and in one of the P. rothschildianum samples. When digested with restriction enzyme DpnII found polymorphisms in fragment length between the two samples. In the other two cases, the products of the expected fragments were amplified in P. lowii and all samples of P. rothschildianum and subsequent RE digestion failed to detect polymorphisms between the species. The overall findings had shown discovery of two SCAR markers to identify P. rothschildianum and P. lowii apart from P. dayanum and one SCAR marker to identify P. dayanum and its possible hybrids. Hence, more studies and screenings need to be done in order to develop species-specific markers for the Paphiopedilum species.



ABSTRAK

PENGUBAHAN PENANDA RAPD KEPADA KO-DOMINAN PENANDA SEQUENCE CHARACTERIZED AMPLIFIED REGION (SCAR) DALAM TIGA SPESIS Paphiopedilum

Genera Paphiopedilum merupakan spesis terancam yang dilindungi di bawah Convention on International Trade in Endangered Species (CITES). Walau bagaimanapun, pengecaman antara spesis adalah sukar disebabkan persamaan ciri-ciri morfologi. Oleh itu, teknik RAPD digunakan untuk mengesan polimorfisma antara tiga spesis: P. dayanum, P. lowii dan P. rothschildianum. Fragmen diagnostik yang dikesan untuk pengecaman setiap spesis akan diubah menjadi penanda SCAR melalui cara pengklonan dan sequencing fragmen diagnostik serta mencipta pasangan primer SCAR untuk mengamplifikasi fragmen-fragmen diagnostik tersebut. Kebolehan penanda yang dicipta manghasilkan fragmen diagnostik diuji melalui PCR diagnostik. Dalam kajian ini, amplifikasi PCR menggunakan primer-primer RAPD telah berjaya mengesan polimorfisma antara spesis. Tiga fragmen diagnostik telah diubah menjadi penanda SCAR dan semua pasangan primer mengamplifikasi satu fragmen spesifik di mana saiz fragmen adalah sama dengan fragmen yang dijangkakan. Satu daripada pasangan primer telah mengamplifikasi saiz fragmen yang dijangkakan dalam spesis P. dayanum dan salah satu sampel P. rothschildianum. Apabila dipotong dengan enzim pembatasan DpnII, didapati terdapat polimorfisma antara saiz fragmen antara kedua-dua sampel. Dalam dua kes yang lain, produk untuk saiz yang dijangkakan telah diamplifikasikan dalam P. lowii dan semua sampel P. rothschildianum serta diikuti pembatasan telah gagal untuk mengesan polimorfisma antara spesis. Keputusan keseluruhan mendapati penemuan dua penanda SCAR untuk membezakan P. rothschildianum dan P. lowii daripada P. dayanum dan satu penanda SCAR untuk mengesan P. dayanum dan hibridnya. Maka, lebih kajian dan pengesanan perlu dijalankan untuk menghasilkan penanda yang spesifik untuk mengesan spesis-spesis Paphiopedilum.



CONTENT

DECI	LARATION	i
CERT	TIFIED BY	ii
ACKI	NOWLEDGEMENT	iii
ABST	TRACT	iv
ABST	TRAK	v
CON	TENT	vi
LIST	OF TABLES	viii
LIST	OF FIGURES	ix
LIST	OF ABBREVIATIONS & SYMBOLS	x
СНА	PTER 1 INTRODUCTION	1
1.1	Introduction	1
1.2	Objectives	3
СНА	PTER 2 LITERATURE REVIEW	5
2.1	Genus Paphiopedilum	5
	2.1.1 Background	5
	2.1.2 Distribution and Ecology	6
	2.1.3 Vegetative Morphology	7
	2.1.4 Floral Morphology	8
2.2	Paphiopedilum Species	9
	2.2.1 Paphiopedilum rothschildianum	9
	2.2.2 Paphiopedilum lowii	10
	2.2.3 Paphiopedilum dayanum	12
2.3	Paphiopedilum on CITES	13
2.4	Distinguishing between Wild and Artificially Propagated Paphiopedilum	15
2.5	Random Amplified Polymorphic DNA (RAPD)	16
	2.5.1 Application of RAPD Analysis	18
2.6	Sequence Characterized Amplified Region (SCAR)	22
CHA	APTER 3 METHODOLOGY	25
3.1	Plant Materials	25



3.2	Methods	26	
	3.2.1 DNA Extraction	26	
	3.2.2 Polymerase Chain Reaction (PCR)	27	
	3.2.3 Gel Extraction	28	
	3.2.4 DNA Cloning	29	
	3.2.5 Plasmid Mini-prep	30	
	3.2.6 Sequence Analysis & Data Analysis	31	
	3.2.7 SCAR Primers Designing	31	
	3.2.8 SCAR Primers Testing	32	
CHA	APTER 4 RESULTS AND DATA ANALYSIS	34	
4.1	DNA Extraction	34	
4.2	PCR Amplification	36	
4.3	Gel Extraction	43	
4.4	DNA Cloning	45	
4.5	Plasmid Mini-prep	46	
4.6	Sequence Analysis		
4.7	Data Analysis	52	
4.8	SCAR Primers Design	54	
4.9	SCAR Primers Testing	55	
СНА	APTER 5 DISCUSSION	60	
5.1	DNA Extraction	60	
5.2	PCR Amplification	61	
5.3	Gel Extraction	62	
5.4	DNA Cloning for Diagnostic Fragments	63	
5.5	Plasmid Mini-prep	64	
5.6	Sequence Analysis	64	
5.7	Data Analysis		
5.8	SCAR Primers Design	66	
5.9	SCAR Primers Testing	67	
CHA	APTER 6 CONCLUSION	71	
REF	ERENCES	74	
APP	ENDIXES	79	



LIST OF TABLES

Table No.

2.1	Differences between wild and artificial Paphiopedilum	16
3.1	The incubation period and temperature of restriction enzyme DpnII	33
	used with its recognition sites	
4.1	Band coring and reproducibility rate for RAPD-PCR of OPA2	38
4.2	Band coring and reproducibility rate for RAPD-PCR of OPA12	40
4.3	Band coring and reproducibility rate for RAPD-PCR of OPA18	42
4.4	The count of colonies in blue-white screening and efficiency rate	45
4.5	DNA sequence of diagnostic fragment of P. rothschildianum	49
4.6	DNA sequence of diagnostic fragment of P. dayanum	50
4.7	DNA sequence of diagnostic fragment of P. lowii	51
4.8	Sequence definition and their respective accession number in GenBank	52
4.9	Putitive alignment of nucleotide sequence encoded by PRO2-1,	53
	PDO12-20 and PLO18-1 using Blastn algorithm	
4.10	Primer sequences for the SCARs and their annealing temperature	55



LIST OF FIGURES

Figure No. Page 4.1 Electrophoresis analysis of DNA extracted from three 35 Paphiopedilum species Electrophoresis analysis of banding profiles of RAPD-PCR of OPA2 39 4.2 4.3 Electrophoresis analysis of banding profiles of RAPD-PCR of OPA12 41 Electrophoresis analysis of banding profiles of RAPD-PCR of OPA18 41 4.4 Electrophoresis analysis of gel purification of extracted PCR products 44 4.5 Electrophoresis analysis of plasmid mini-prep for P. rothschildianum 4.6 47 diagnostic band 4.7 Electrophoresis analysis of plasmid digestion with restriction enzyme 47 EcoRI Electrophoresis analysis of plasmid mini-prep for P. dayanum 4.8 48 diagnostic band 4.9 Electrophoresis analysis of plasmid mini-prep for P. lowii diagnostic band 48 4.10 Electrophoresis analysis of SCAR-PCR amplification of SPRO2-1 56 4.11 Electrophoresis analysis of RFLP with DpnII for SPRO2-1 56 amplification products 4.12 Electrophoresis analysis of SCAR-PCR amplification of SPDO12-20 58 4.13 Electrophoresis analysis of RFLP with DpnII for SPDO12-20 58 amplification products Electrophoresis analysis of SCAR-PCR amplification of SPLO18-1 4.14 59 4.15 Electrophoresis analysis of RFLP with DpnII for SPLO18-1 59 amplification products



ix

LIST OF ABBREVIATIONS & SYMBOLS

bp	base pair	
g	gram	
kb	kilo base	
М	molar	
ng	nanogram	
mM	millimolar	
μl	microlitre	
rpm	revolution per minute	
pmol	picomole	
%	percent	
°С	degree Celcius	
dH ₂ O	de-ionized distilled water	
DNA	Deoxyribonucleic	
dNTP	Deoxynucleoside triphosphate	
EDTA	Ethylenediamine tetraacetic acid	
MgCl ₂	Magnesium chloride	
min	minutes	
NaCl	Natrium chloride	
PCR	Polymerase Chain Reaction	
RAPD	Random Amplified Polymorphic DNA	
RE	Restriction Enzymes	
RFLP	Restriction Fragment Length Polymorphism	
RNase	Ribonuclease	
S	seconds	
SCAR	Sequence Characterized Amplified Region	
SDS	Sodium dodecyl sulphate	
Taq	Thermus aquaticus	
TBE	Tris Boric EDTA	
TE	Tris-HCl EDTA	
Tris	Tris (hydroxymethyl) aminomethane	



Tris HCl Tris (hydroxymethyl) aminomethane hydrochloride

unit

U

V volt

xi



•

CHAPTER 1

INTRODUCTION

1.1 Introduction

The genus *Paphiopedilum* belongs to the family Orchidaceae and subfamily Cypripedioideae. *Paphiopedilum* is known as the slipper orchids because their lip resembles a lady's slipper. The range of *Paphiopedilum* species are found from India eastward across southern China to the Philippines and throughout south-east Asia and the Malay Archipelago to New Guinea and the Solomon Islands (Cribb, 1998). In Malaysia, the *Paphiopedilum* or slipper genus, one of the rarest orchids, is found only in a remote part of Mount Kinabalu.

Paphiopedilum orchids are listed on Appendix I of the Convention on International Trade in Endangered Species (CITES), under which trade is banned or regulated with special permits. This increases their desirability of the Paphiopedilum species and causes it to be hunted by orchid collectors. These collectors are obsessed with the Paphiopedilum species to the extend of committing crime by smuggling. For instance, in the year 2004, the head of research and development of Medpharm Ltd.,



a pharmaceutical company, was sentenced to 50 months in jail by the British court for smuggling 126 rare and protected slipper orchids, including the rare Sabah species, into Britain (John & Kaur, 2006)

The *Paphiopedilum* genus is protected under local legislation as well. In Sarawak, all orchids are protected species under the Wildlife Protection Ordinance established at year 1998. Meanwhile, in Sabah, the *Paphiopedilum* species are totally protected under the Wildlife Conservation Enactment established at year 1997. The penalty of those in possession of the species is a maximum fine of RM50,000, five years' jail or both. However, local communities continued to collect and sell the *Paphiopedilum* species for as low as RM5 to the tourists which are illegal and violate the value of the *Paphiopedilum* species.

Leaf shape in orchids is relatively uniform for many genera where most orchids have simple, linear, lanceolate, oblanceolate, ovate, or elliptic leaves. The *Paphiopedilum* species also have regular leaf morphology similar to the ordinary orchid genus when the plant is not flowering. Therefore, it is hard to determine the *Paphiopedilum* species morphologically. Hence, it is difficult to stem smuggling.

There have been a few studies using Random Amplified Polymorphic DNA (RAPD) analysis to determine the genetic diversity and relationship between *Paphiopedilum* and *Phragmipedium* species and cultivars (Chung *et al.*, 2006) and a preliminary analysis of the level and apportionment of genetic diversity in *Paphiopedilum micranthum* using RAPD analysis (Li *et al.*, 2002). Thus far, there is no study has been made for species identification for *Paphiopedilum* species.



Therefore, in this study, RAPD markers are used to create species-specific SCAR markers for differentiation of each of the *Paphiopedilum* species.

RAPD markers work by amplifying random sequences from a DNA template with a single arbitrary 10-mer primer. RAPD markers are dominant because polymorphisms are detected as the presence or absence of bands. Thus, it could not differentiate between homologous and heterozygous loci. Therefore, RAPD markers are transformed into a codominant marker called Sequence Characterized Amplified Region (SCAR) which allows for a reproducible amplification of a single RAPD fragment. SCAR markers are PCR-based markers that represent single, genetically defined loci that are identified by PCR amplification of genomic DNA with pairs of specific oligonucleotide primers. The sequence of the SCAR marker is designed from the amplified RAPD products that were cloned and sequenced.

In this study, species specific codominant RAPD-SCAR markers will be designed for three *Paphiopedilum* species: *P. rothschildianum*, *P. lowii* and *P. dayanum* for identification of each of these *Paphiopedilum* species.

1.2 Objectives

a. To determine specific Random Amplified Polymorphic DNA (RAPD) polymorphic markers among *Paphiopedilum lowii*, *P. dayanum* and *P. rothschildianum*.



- b. To convert the diagnostic RAPD marker for *P. lowii*, *P. dayanum* and *P. rothschildianum* into species-specific Sequence Characterized Amplified Region (SCAR) markers by performing cloning, sequencing and primer design.
- c. To test for reproducibility of the species-specific SCAR markers on *P. lowii*, *P. dayanum* and *P. rothschildianum* by PCR amplification.



CHAPTER 2

LITERATURE REVIEW

2.1 Genus Paphiopedilum

2.1.1 Background

Paphiopedilum derives from the Greek 'Paphio', an epithet for Aphrodite (the goddess known as Venus to the Romans), and 'pedilon' meaning slipper. Thus, the genus *Paphiopedilum* is also known as the slipper orchids, so called for the shape of the deeply saccate lip of their flower, represent a small but remarkable offshoot of the main line of orchid evolution (Cribb, 1998).

Paphiopedilum is the largest genus of slipper orchids with some 80 species occurring in the Asian tropics from southern India to New Guinea and the Philippines. They can be found growing on the ground, on rock and cliff surfaces and attached to trees and other vegetation. Most are ground-growing, growing in leaf litter or in cracks in rocks containing organic matter. They occur in a wide range of habitats from



branches of large trees in the rainforests of Thailand to the harsh serpentine soils on Mount Kinabalu (McGough *et al.*, 2006).

2.1.2 Distribution and Ecology

The range of *Paphiopedilum* extends from India eastward across southern China to the Philippines and throughout south-east Asia and the Malay Archipelago to New Guinea and the Solomon Islands. Of the seven Indian species, six are confined to the north-east, along the foothills of the Himalaya from eastern Nepal to the Naga Hills, and also in the Khasia Hills. Meanwhile, there are eighteen species have been reported from China (Pridgeon *et al.*, 1999).

The genus is well represented in the south-east Asia, notably in Thailand and Vietnam which can boast 12 species but with few in adjacent countries. In Borneo, there are twelve species of which seven are endemic. Meanwhile, Sumatra has ten species and six endemics, whereas Java has only three species and one endemic (Cribb, 1998).

The Philippines are another rich centre of diversity of the genus with eight species, seven of which are endemic. To the south and east the number of species drops rapidly. Four species have been recorded from Sulawesi and the Moluccas, one from each being endemic. In New Guinea four species, all endemic, are found with two reported from the Solomons (Pridgeon *et al.*, 1999).



Five species of *Paphiopedilum* have been reported as growing epiphytically; *P. parishii*, *P. lowii* and *P. villosum* are usually found growing on trees while *P. hirsutissimum* and *P. glanduliferum* are facultative epiphytes. The remaining species are either terrestrial or lithophytic (Cribb, 1998).

2.1.3 Vegetative Morphology

Paphiopedilum leaves are leathery with a prominent middle rib. The leaves are V shaped in cross-section. Leaves may be short and strap-like or oblong to linear. The leaves are usually short, less than 20 centimetres in length. An exception to this rule is the multiflowered group which includes species such as Paphiopedilum sanderianum, P. rothschildianum and P. lowii. Leaf colour ranges from plain or glossy green to mottled purple and can be quite useful in identification (McGough et al., 2006).

The leaves of the *Paphiopedilum* are often distinctive and taxonomically useful at the subgeneric and even in some cases species level. Leaf shape varies from linearligulate in sections *Paphiopedilum* and *Coryopedilum* to elliptic-oblong in most of the species of subgenus *Brachypetalum*. The upper surface of the leaves is uniformly green in all the species of sections *Paphiopedilum* and *Coryopedilum*. Paler margin are characteristic of some species. Mottling or tessellation of the upper surface is characteristic of the species of subgenera *Parvisepalum* and *Brachypetalum*. The tessellation of the leaves of section *Barbata* species is distinct from that of *Brachypetalum*. Meanwhile, faint tessellations can be seen on the upper surface of *P*. *victoria-regina* and *P*. *victoria-mariae* in section *Cochlopetalum* but not at all in other species. The lower surface of the leaves is also distinctively marked in many species.



In most species of subgenera *Parvisepalum* and *Brachypetalum*, the leaves are densely purple-spotted beneath (Cribb, 1998).

2.1.4 Floral Morphology

The most distinctive features of the *Paphiopedilum* plant are to be found in its flower. The flower has an inferior unilocular ovary with parietal placentation; a conspicuous upper or dorsal sepal and a less obvious synsepal formed by the united lateral sepals; two spreading petals that are usually deflexed or reflexed and often twisted; a deeply saccate ventral pouched lip, usually with more or less obvious side lobes; and a central short column bearing a ventral stalked trilobed stigma, two lateral and ventral anthers and an apical, usually shield-shaped, staminode (Cribb, 1998).

Petal shape, marking and indumentum are all taxonomically useful characters. In subgenera *Parvisepalum* and *Brachypetalum* the petals are more or less elliptic to circular in shape and usually less than twice as long as broad. Petals that taper from base to apex are found in all the species of section. In *Cochlopetalum* the petals are more or less linear, spreading and spirally twisted. The petal margins are also markedly ciliate, the ciliae being multicellular hairs. The remaining species in the genus have oblong to spathulate petals, the latter feature being particularly marked in species such as *P. lowii*, *P. haynaldianum*, *P. bullenianum*, *P. appletonianum* and *P. hookerae*.

The third petal in the flower of *Paphiopedilum* is highly modified to form a slipper-shaped lip. The lip is effectively three-lobed with mid-lobe deeply saccate and



the side-lobes infolded to give the lip an apparent claw. Four types of lips can be distinguished in the genus. An inflated, thin-textured and brightly coloured lip is characteristic of subgenus *Parvisepalum*. In subgenus *Brachypetalum* the lip is smaller, ovoid, and more solid-textured. In both these subgenera the apical margin is involute in the manner of many *Cypripedium* species and of *Phragmipedium besseae* and *P. schlimii*.

In section *Barbata*, *Cochlopetalum*, *Paphiopedilum* and *Pardalopetalum* the margin of the mid-lobe is not involute but the side-lobes are well developed and incurved to form a tube at the base of the lip. Finally, in section *Coryopedilum*, the side-lobes are much reduced and represented only by acute ear-like lobes that point into the saccate mid-lobe of the lip. In both the latter cases the form is paralleled in the genus *Phragmipedilum* (Cribb, 1998).

2.2 Paphiopedilum Species

2.2.1 Paphiopedilum rothschildianum

The *P. rothschildianum* belongs to the subgenus of *Paphiopedilum* in section *Coryopedilum*. This orchid is endemic to Mount Kinabalu in north-east Borneo (Pridgeon *et al.*, 1999). Of all the species in the genus, *P. rothschidianum* is one of the rarest in nature. It has been located in only three sites on the lower slopes around Mount Kinabalu, in one of which it is certainly now extinct. It usually grows on ledges on steep slopes and cliffs of ultra-basic rock where it seems to thrive in the open as well



as in shaded places. Fortunately, *P. rothschildianum* grows only inside the Kinabalu Park and is afforded some protection.

P. rothschildianum is a terrestrial or lithophytic herb often growing in large clumps. The leaves of this specie are several, linear to narrowly oblanceolate, acute, up to 60 cm long, 4 to 5 cm wide, sparsely ciliate at base and green in colour. The inflorescence is 2 to 4-flowered and erect; the peduncle is up to 45 mm long and purplish while the bracts are ovate-elliptic, obtuse, up to 5.5 cm long, ciliate and hairy on midvein, pale green or yellow in colour with purple-striped. The flowers are very large within 14 to 30 cm in diameter while the ovary can be up to 7.5 cm long and pale green with sparsely spotted purple. The dorsal sepal is ovate, 5.4 to 6.8 cm long, 3.3 to 4.8 cm wide and in the shade of ivory-white or yellow with maroon veins. The synsepal is similar but smaller compare to dorsal sepal with 4.8 to 6 cm long and 3.2 to 4.4 cm wide. Petals are narrowly tapering to rounded apex, 8 to 14 cm long, 0.7 to 1.5 cm wide, ciliate, papillose towards apex and in the shade of yellow or ivory-white marked with maroon. The lip is subporrect, grooved on back, 5.3 to 6 cm long, 2.2 to 2.5 cm wide and golden in colour with heavily purple-suffused. The staminode is linear, bifid at apex, geniculate, 14 to 16 mm long, 4 to 5 mm wide, densely glandular-pubescent on margins and at the base and pale yellow-green in colour (Cribb, 1998).

2.2.2 Paphiopedilum lowii

The *P. lowii* belongs to the subgenus of *Paphiopedilum* in the section of *Pardalopetalum*. This specie is the most widespread of the multiflowered species, found throughout the Malay Peninsular, Sumatra, Java, Borneo and the Celebes (Sulawesi)



(Pridgeon *et al.*, 1999). It is also one of the few epiphytic species in the genus, although it may also occasionally be found growing on rocks. *P. lowii* can be found growing at altitudes between 250 m and 1600 m in riverside, lower mountain and mountain rainforest either as an epiphyte on the trunks and branches of trees or as a lithophyte in moss- or humus-filled hollows of rocks especially limestone and of boulders.

P. lowii is an epiphytic herb that grows on trees. Its leaves are commonly about four to six, linear-ligulate, unequally roundly bilobed at apex 22 to 40 cm long, 2.8 to 6 cm wide and mid-green in colour. The inflorescence is erect-arcuate and 3 to 7-flowered; the peduncle is green, mottled purple, shortly pubescent and up to 70 cm long while the bracts are elliptic, obtuse, 2 to 4.5 cm long, 2.2 cm wide and in the shade of yellow with purple marks. The flowers are 9 to 16.5 cm across while pedicel and ovary is 4.5 to 7 cm long and greenish in shade. The dorsal sepal is elliptic-ovate, obtuse, 3 to 5.5 cm long, 2.5 to 3.5 cm wide, undulate and ciliate on margins, pale green colour and mottled dull purple in basal half with recurved basal margins. The synsepal is elliptic, obtuse, 2.2 to 4.8 cm long, 2 to 2.8 cm wide, 2-keeled on outer surface and pale green in colour. The petals often once-twisted in middle, spatulate, subacute to obtuse, 5 to 9.3 cm long, 1.5 to 3.1 cm wide, ciliate and pale yellow in colour with a purple apical third and maroonspotted in basal two-thirds. The lip is 3.5 to 6 cm long, 2.1 to 3.1 cm wide and in the shade of dull ochre-brown. The staminode is obovate, apically three-toothed with a long erect hook at the base, 10 mm long, 7 mm wide and pale ochre to brownish green in colour (Cribb, 1998).



REFERENCES

- Altschul, S. F., Madden, T. L., Schäffer, A. A., Zhang, J., Zhang, Z., Miller, W. & Lipman, D. J. 1997. Gapped BLAST and PSI-BLAST: A new generation of protein database search programs. *Nucleic Acids Research* 25, pp. 3389–3402.
- Bowtell, D. & Sambrook, J. 2003. DNA Microarrays: A Molecular Cloning Manual. Cold Spring Harbor Laboratory Press, New York.
- Cheghamirza, K., Koveza, O., Konovalov, F. & Gostimzky, S. 2002. Identification of RAPD markers and their use for molecular mapping in pea (*Pisum sativum L.*). Cellular & Molecular Biology Letters 7, pp. 649-655.
- Chung, S. Y., Choi, S. H., Kim, M. J., Yoon, K. E., Lee, G. P., Lee, J. S. & Ryu, K. H. 2006. Genetic relationship and differentiation of *Paphiopedilum* and *Phragmepedium* based on RAPD analysis. *Scientia Horticulture* 109, pp. 153-159.
- Coruzzi, G. & Puigdornènech, P. 1994. NATO ASI Series, Vol. H 81 Plant Molecular Biology. Inside: Tingey, S. V., Rafalski, A. & Hanafey, K. 1994. Genetic Analysis with RAPD markers. Springer-Verlag, Berlin Heidelberg, pp. 491-498.
- Cox, A. V., Pridgeon, A. M., Albert, V. A. & Chase, M. W. 1997. Phylogenetics of the slipper orchids (Cypripedioideae, Orchidaceae): nuclear rDNA ITS sequences. *Plant Systematics and Evolution* 208, pp. 197-223.
- Cribb, P. 1998. The Genus Paphiopedilum. 2nd ed. Natural History Publications, Kota Kinabalu.
- Das, M., Bhattacharya, S. & Pal, A. 2005. Generation and characterization of SCARs by cloning and sequencing of RAPD products: A strategy for species-specific marker development in Bamboo. *Annals of Botany* 95, pp. 835-841.



- Dellaporta, S. L., Wood, J. & Hicks, J. B. 1983. A plant DNA minipreparation. Plant Molecular Biology Reports 1, pp. 19-21.
- Fang, G., Hammar, S. & Grumet, R. 1992. A quick and inexpensive method for removing polysaccharides from plant genomic DNA. *Biofeedback* 13, pp. 52-54.
- Goh, M. W. K., Kumar, P. P., Lim S. H. & Tan, H. T. W. 2005. Random amplified polymorphic DNA analysis of the moth orchids, *Phalaenopsis (Epidendroideae:* Orchidaceae). Euphytica 141, pp. 11–22.
- Gosselin, I., Zhou, Y., Bousquet, J. & Isabel, N. 2002. Megagametophyte-derived linkage maps of white spruce (*Picea glauca*) based on RAPD, SCAR and ESTP markers. *Theoretical and Applied Genetics* 104, pp. 987-997.
- Hernández, P., de la Rosa, R., Rallo, L., Dorado, G. & Martin, A. 2001. Development of SCAR markers in olive (*Olea europaea*) by direct sequencing of RAPD products: applications in olive germplasm evaluation and mapping. *Theoretical and Applied Genetics* 103, pp. 788-791.
- Hul, J., Lil, G. Struss, D. & Quirosl, C. F. 1999. SCAR and RAPD markers associated with 18-carbon fatty acids in rapeseed, *Brassica napus*. *Plant Breeding* 118, pp. 145-149.
- John, E. & Kaur, J. 2006. Beauty and the thief. The New Straits Times. 12 March, pp. F1-F4.
- Joshi, S. P., Ranjekar, P. K. & Gupta, V.S. 1999. Molecular markers in plant genome analysis. *Current Science* 77, pp. 230-240.
- Li, A. & Ge, S. 2006. Genetic variation and conservation of *Changnienia amoena*, an endangered orchid endemic to China. *Plant Systemics and Evolution* 258, pp. 251-260.



- Li, A., Luo, Y. B., Xiong, Z. T. & Song, G. E. 2002. A preliminary study on conservation genetics of three endangered orchid species. *Acta Botanica Sinica* 44, pp. 250-252.
- Lim, S. H., Teng, P. C. P., Lee, Y. H. & Goh, C. J. 1999. RAPD analysis of some species in the genus Vanda (Orchidaceae). Annals of Botany 83, pp. 193–196.
- Manfield, I. W., Pavlov, V. K., Li, J., Cook, H. E., Hummel, F. & Gilmartin, P. M. 2005. Molecular characterization of DNA sequences from the *Primula vulgaris* S-locus. *Journal of Experimental Botany* 56, pp. 1177-1188.
- McGough, H. N., Roberts, D. L., Brodie, C. & Kowalczyk, J. 2006. CITES and Slipper Orchids – An Introduction to Slipper Orchids Covered by the Convention on International Trade in Endangered Species. Royal Botanic Gardens, Kew, United Kingdom.
- Mitchelson, K. R., Drenth, J., Hong, D. & Chaparro, J. X. 1999. Direct Sequencing of RAPD fragments using 3'-extended oligonucleotide primers and dye terminator cycle-sequencing. *Nucleic Acids Research* 27, pp. e28 i - e28 iv.
- Ohmori, T., Murata, M. & Motoyoshi, F. 1996. Molecular characterization of RAPD and SCAR markers linked to the Tin-1 locus in tomato. *Theoretical and Applied Genetics* 92, pp. 151-156.
- Paran, I. & Michelmore, R. W. 1993. Development of reliable PCR-based markers linked to downy mildew resistance genes in lettuce. *Theoretical and Applied Genetics* 85, pp. 985-993.
- Porebski, S., Bailey, L. G., & Baum, B. R. 1997. Modification of a CTAB DNA extraction protocol for plants containing high polysaccharide and polyphenol components. *Plant Molecular Biology Reporter* 15, pp. 8-15.



- Pridgeon, A. M., Cribb, P. J., Chase, M. W. & Rasmussen, F. N. 1999. Genera Orchidacearum Volume 1. Oxford University Press, New York.
- Reiter, R. S., Williams, J. G. K., Feldmann, K. A., Rafalski, A., Tinger, S. V. & Scolnik, P. A. 1992. Global and local genome mapping in *Arabidopsis thaliana* by using recombinant inbred lines and random amplified polymorphic DNAs. *Proceeding* of the National Academy of Science 89, February 1992, USA, pp. 1477-1481.
- Sambrook. J., Fritsch, E. P. & Moniatis, T. 1989. Molecular Cloning: A Laboratory Manual. Cold Spring Laboratory Press, New York.
- Sun, M. & Wong, K. C. 2001. Genetic structure of three orchid species with contrasting breeding systems using RAPD and allozyme markers. *American Journal of Botany* 88, pp. 2180-2188.
- Vidal, J. H., Delavault, P., Coarer, M. & Defontaine, A. 2000. Design of grapevine (Vitis vinifera L.) cultivar specific SCAR primers for PCR fingerprinting. Theoretical and Applied Genetics 103, pp. 1194–1201.
- Wang, B. & Porter, A. H. 1998. On the feasibility of developing a RAPD-based, codominant marker system for evolutionary studies. *Journal of Clinical Microbiology* 36, pp. 2057-2062.
- Wang, C., Zhang, P., Ma, Z., Zhang, M., Sun, G. & Ling, D. 2004. Development of a genetic marker linked to a new thermo-sensitive male sterile gene in rice (*Oryza* sativa L.) Euthytica 140, pp. 217-222.
- Welsh, J. & McClelland, M. 1990. Fingerprinting genomes using PCR with arbitrary primers. Nucleic Acids Research 24, pp. 7213-7218.



- Williams, J. G., Kubelik, A. R., Livak, K. J., Rafalski, J. A. & Tingey, S. V. 1990. DNA polymorphisms amplified by arbitrary primers are useful as genetic markers. *Nucleic Acids Research* 18, pp. 6531–6535.
- Wong, K. C. & Sun, M. 1999. Reproductive biology and conservation genetics of Goodyera procera (Orchidaceae). American Journal of Botany 86, pp. 1406-1413.
- Ye, Q., Qiu, Y. X., Quo, Y. Q. Chen, J. X., Yang, S. Z. Zhao, M. S. & Fu, C. X. 2006. Species-specific SCAR markers for authentication of *Sinocalycanthus chinensis*. *Journal of Zhejiang University SCIENCE B* 7, pp. 868-872.

